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# *O***-Protecting groups as long-range stereocontrolling elements in the addition of acetylides to 4-substituted quinolines**

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**Abstract—**Addition of magnesium acetylides in the presence of chloroformate esters to racemic differently *O*-protected 2-(4 quinolyl)propanols and to enantiopure 2-(4-quinolyl)-1,3-dialkoxypropanes, prepared by a chemoenzymatic route, gives almost exclusive regioselective attack at C-2, with stereoselectivities from moderate to good, depending mainly on the bulkiness of the *O*-protecting group present. © 2002 Elsevier Science Ltd. All rights reserved.

## **1. Introduction**

The quinoline nucleus, and also its hydrogenated form are fragments frequently found in many natural products. $1-3$  Moreover, many quinoline derivatives are known for their interesting biological properties, such as antimalarial activity,  $4,5$  antagonist activity with respect to neurokinin-3,<sup>6</sup> glucagon<sup>7</sup> or cannabinoid<sup>5</sup> receptors, antiprotozoal<sup>8</sup> and antitumor<sup>9,10</sup> activity. Dynemicin A, a recently discovered<sup>11</sup> enediyne, containing a quinoline derivative, has attracted the attention of many research groups for its notable cytotoxicity and novel mode of action. Some syntheses of this compound have also been reported.<sup>12–14</sup> However, its complex structure and low stability has prompted many research groups to design simplified analogues, which still have their biological activity promoted by the presence of the enediyne moiety.<sup>15</sup> In the last years our group has been involved in the preparation of new artificial enediynes.<sup>16</sup> Among them, we aimed to synthesize a new family of analogues of natural Dynemicin A, namely compounds **1a**,**b**, and the corresponding hydroxymethyl derivatives **2a**,**b**.

In our strategy we planned to assemble the 10-membered enediyne ring, starting from highly functionalized quinoline-derived intermediates **3** and **4**, provided with

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both triple bonds. Because we envisaged the employment of the appropriate 4-substituted quinolines as the first stage of the synthesis, we had to study the regioand diastereoselective functionalization of the parent heterocycle, starting from compounds of general formula **5** and **6**, as summarized in Scheme 1. Preliminary results in this field have already been reported on racemic compounds **5**. <sup>17</sup> Herein, we report more details on diastereoselective additions to **5** and extend our study to optically active **6**.



**Scheme 1.**

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## **2. Results and discussion**

## **2.1. Preparation of quinolines (±)-5 and 6 and addition of acetylides to the heterocyclic ring**

For the preparation of (±)-**5a**, 4-ethylquinoline, obtained through a Skraup-like reaction (Scheme  $2$ ),<sup>18,19</sup> was heated under reflux in aqueous formaldehyde, following the same procedure employed for the preparation of 1,3-propanediols with 2- or 4-pyridyl<sup>20</sup> or quinoly $l^{21}$  substituents at C-2. In this case, however, a complex mixture of hemiacetal compounds of formaldehyde was formed. The mixture is best hydrolyzed under basic conditions, which afford the desired (±)-**5a** in high yield.

Several *O*-protected derivatives of  $(\pm)$ -5a, with different properties, have been prepared in satisfactory to excellent yields, following standard procedures, as reported in Scheme  $2<sup>22</sup>$  We also tried to introduce other protecting groups into  $(\pm)$ -**5a**, with the groups differing not only in their steric hindrance properties, but also in their ability to give chelation in reactions involving organometallic intermediates. Actually, in some cases this is a great opportunity for controlling diastereoselectivity.<sup>23</sup> However, all attempts to transform  $(\pm)$ -5a into a series of common ethers characterized by chelating properties, like benzyl, 4-methoxybenzyl, methoxymethyl (MOM), methoxyethoxyethyl (MEM) or 2,4-dinitrophenyl ether, employing different conditions,<sup>22</sup> failed. We observed unexpectedly low reactivity of (±)-**5a** and, in every case, the desired products could not be isolated since the starting alcohol usually did not react, while forcing reaction conditions only led to decomposition of the substrate.

The optically active quinolines **6b**–**g** were synthesized starting from monoacetate **6a**, which could be prepared in very high chemical yield, with enantiomeric excess up to 97.5% by asymmetrization of the corresponding diol catalyzed by lipase from *Candida antarctica*, as previ-



ously reported.<sup>21</sup> This compound was then protected varying the groups at the two oxygenated functionalities. For this purpose we planned to introduce two *O*-protecting groups with orthogonal properties, having in mind to manipulate them at different times after the addition reaction. As previously experienced with  $(\pm)$ -**5a**, we succeeded with the introduction of only a limited number of protecting groups: triphenylmethyl ether and their analogues, silyl ethers, anisyl ether, esters (Scheme 3). Protecting groups requiring the formation of the alcoholate of **6a** could not be introduced. Acetal protecting groups were the most troublesome: while the methoxymethyl ether could be prepared in moderate yield (40%) but with poorly reproducible results; we did not succeed, for example, in the introduction of the very versatile tetrahydropyranyl ether.

This was quite puzzling, since it prevented the study of the addition reaction on a system bearing at the same time a chelating and a 'non-chelating' group, $^{23}$  a condition that should influence the diastereoselectivity during the nucleophilic attack.

Having in hand a certain number of *O*-protected quinolines **5** and **6**, we started our studies on the regio- and diastereoselective addition of *C*-nucleophiles. The propensity of quinolines, pyridines and isoquinolines to undergo nucleophilic addition at C-2 and/or C-4 is well known, and has received increased attention after the discovery of Dynemicin A. For the synthesis of simplified analogues of Dynemicin A, the functionalization of the quinoline nucleus by means of *C*-nucleophiles is a milestone either when performed on achiral  $\frac{1}{24-28}$  or partially rigid quinolines fused with other rings and bearing at least two stereocenters.29–31 The asymmetric synthesis of 1,2- and 1,4-dihydroquinolines by nucleophilic addition to a chiral aminal (with  $C_2$ -symmetry) of quinoline-3-carbaldehyde is also known.<sup>32</sup> To the best of our knowledge only one example is known in which the reaction is performed on a quinoline with just one stereocenter.<sup>33</sup> In order to prepare the precursors of **1a**,**b** and **2a**,**b** it is important to fully understand the regio- and stereochemical course of the addition on such a conformationally flexible







quinolyl derivative with a pre-existing stereogenic center remote from the one generated during the reaction. Thus, every factor that can affect the regio- and diastereoselectivity as well as the yield, leading to compounds **8** and **9** (Scheme 4), was carefully investigated.<sup>17</sup>

Compound  $(\pm)$ -**5b** was first chosen as the starting substrate and some meaningful data are presented in Table 1.<sup>†</sup> The low electrophilicity of the heterocycle was enhanced by performing the addition of the nucleophile in the presence of a suitable alkyl or aryl chloroformate.<sup>34</sup> In this case, the best results were obtained using phenyl chloroformate, which allowed the addition to be complete in most cases at −78°C. In contrast, the reaction was very slow when benzyl or methyl chloroformate were used (entries 1 and 2) and, with the first reagent, we observed a decrease in the diastereomeric ratio. The reaction was always very regioselective, with



nucleophilic attack occurring only on C-2 (with the exception of entry 6 in which cerous acetylide also gave a small additional amount of attack on C-4). Changing the order of addition of the reagents, the reaction temperature, or the counterion of the acetylide affected only the yields, while the observed d.r. were not influenced in an appreciable manner. One exception was seen with lithium acetylide, which gave a complex reaction mixture, the addition products being present only in negligible amounts. The screening was performed on 100 mg of  $(\pm)$ -5**b** using 3 molar equiv. of a 0.5 M solution of acetylide in THF and 3 molar equiv. of chloroformate. When the reaction was scaled up to 10–14 mmol, the amount of reagents was reduced to not more than 1.5–2 molar equiv., without observing appreciable changes. In all cases, however, the d.r. was moderate, with **8** always predominating. This fact prompted us to study the influence of the protecting group of the primary alcohol function on the diastereoselectivity.

The same reaction was performed on the complete series of racemic quinolines **5**, reproducing the conditions of entry 3 (Table 1). The results are shown in Table 2.

Apart from acetyl as a protecting group, for which a diminished d.r. with respect to SiMe<sub>2</sub><sup>t</sup>-Bu was observed (entry 5), in all other cases (entries 1–4, 6–11) higher d.r. was noticed. In particular, the best results were obtained using the triphenylmethyl group; in this case d.r. up to 87:13 was observed (entry 8). This ratio can be viewed as excellent considering the distance between the newly formed and pre-existing stereogenic centers. Moreover, simple fractional crystallization allowed us **Scheme 4.** to increase the d.r. up to 99:1 in a very simple manner.

**Table 1.** Addition of acetylides to  $(\pm)$ -5b using different experimental conditions  $(R^6 = \text{SiMe}_2 t \cdot \text{Bu}; R^1 = \text{Me}; R^2 = \text{SiMe}_3)$ 

Entry	$R^3$	Nucleophile	Temperature $(^{\circ}C)$	Isolated yield $(\%)^a$	( $\pm$ )-8a–c:( $\pm$ )-9a–c <sup>b</sup>
	<b>B</b> n	$SiMe3C \equiv CMgBr$	$-78$ to $-2^{\circ}$	72 (69)	59:41 <sup>d</sup> (a)
2	Me	$SiMe3C \equiv CMgBr$	$0^{\circ}$	78 (76)	66:34(b)
	Ph	$SiMe3C \equiv CMgBr$	$-78$ to $-2^{\circ}$	77 (64)	$68:32^d$ (c)
4	Ph	$SiMe3C \equiv CMgBr$	$-78$ to 0 <sup>c</sup>	87 (84)	68:32(c)
5.	Ph	$SiMe3C \equiv CMgBr$	$-78^{\circ}$	66 $(63)^e$	67:33(c)
6	Ph	$SiMe3C=CCeCl2$	$0^{\circ}$	30 $(24)^f$	68:32(c)
	Ph	$SiMe3C=CMgCl$	$-78$ to $0^{\circ}$	35 (34)	68:32(c)

<sup>a</sup> Since by-products derived from chloroformate are always present and not easy to separate from **8**/**9**, the yield after purity determination by GLC analysis is reported in brackets.

<sup>b</sup> Determined by GLC, if not otherwise specified.

 $c$  Reaction conditions: [for each entry order of addition of reagents was reported (quinoline = a, nucleophile = b, chloroformate = c, temperature and reaction times were reported only if necessary); reactions were performed on 100 mg of  $(\pm)$ -5b, using 3 molar equiv. of nucleophile (0.5 M) sol. in THF) and 3 molar equiv. of chloroformate]. *E*1, *E*3, *E*7: a+b+c, *E*2: b+a+c (after 10 min), *E*4: b+c (−78-0°C, 1.5 h)+a (−78°C), *E*5: b+a (3 h)+c, *E*6: b+a+c.

<sup>d</sup> Determined by GC–MS (HP-1 column).

<sup>e</sup> This reaction was performed on a preparative scale (10 mmol) under these conditions, but using 2–3 molar equiv. of nucleophile and 1.5 molar equiv. of chloroformate to give, after 15 h, the mixture  $(\pm)$ -8,9 in 96% yield.

<sup>f</sup> Together with 4–5% products derived from addition of nucleophile on C-4.

<sup>†</sup> Complete data have already been reported (Ref. 17).





<sup>a</sup> See note (a), Table 1 (in this case GLC or HPLC).

<sup>b</sup> In all cases **8** prevailed.

<sup>c</sup> see note (c), Table 1: *E*1, *E*3–*E*17: a+b+c, *E*2: a+c (0°C, 30 min)+b (−78°C).

<sup>d</sup> Determined by HPLC.

<sup>e</sup> Determined by GLC.

f D.r. determined more precisely after Tr removal to give the corresponding alcohols  $(\pm)$ -8,9d.

<sup>g</sup> This reaction was performed on a preparative scale (14 mmol) under these conditions, but using 2–3 molar equiv. of nucleophile and 1.5 molar equiv. of chloroformate to give, after 18 h, the mixture  $(\pm)$ -8.9*j* in 99% yield.

<sup>h</sup> After removal of Tr.

<sup>i</sup> Determined by <sup>1</sup>H NMR.

<sup>j</sup> Determined by <sup>13</sup>C NMR after Tr removal.

<sup>k</sup> *p*-Methoxyphenyldiphenylmethyl.

<sup>l</sup> Di(*p*-methoxyphenyl)phenylmethyl.

Using different acetylides—compounds having an alkoxymethyl group instead of  $\text{SiMe}_3$ —we did not observe appreciable effects on the results (entries 10 and 11), since these groups are probably too far away from the zone responsible for asymmetric induction. In this case, the addition products could not be separated from the excess of alkyne employed, which meant that accurate characterization and precise determination of the d.r. could not be performed until after removal of the trityl group. For analytical purposes, the mixture of products could always be separated into pure (±)-**8** and  $(\pm)$ -9, with the exception of compounds  $(\pm)$ -8–9h,k,l. Anyway, the best separation in view of further synthetic elaborations was obtained after *O*-protecting group removal.

The additions to **6b**–**g** were performed using trimethylsilylmagnesium acetylide (entries 12–17, Table 2). With **6b** the chemical yield was excellent (entry 12), while the d.r. was considerably lower than with compound (±)-**5h**. Planning the forthcoming elaboration of **8**,**9**, we also tested the addition reaction with substituted trityl derivatives, that can, in principle, be removed under milder conditions.22 For this reason we employed both *p*-methoxyphenyldiphenylmethyl (MMTr) and di(*p*-methoxyphenyl)phenylmethyl (DMTr) ethers: with the first protecting group we

obtained **8**,**9** with the same d.r. as the trityl protected derivative, but the yield was only moderate (entry 13), while the DMTr group in **6c** proved to be unstable under the conditions utilized and, the disappearance of starting material was accompanied by the formation of many other products. Unexpected behavior was also observed with **6e**: the presence of a free hydroxyl function was actually responsible for extensive decomposition, which is in contrast with the results obtained with alcohol (±)-**5a** (entry 1). Triethylsilyl protected **6f** was a good substrate for the addition reaction (entry 16), giving indeed an inseparable mixture of diastereoisomers but in excellent overall yield, albeit with a diminished diastereoisomeric ratio. Curious behavior was seen with **6g**, since the reaction evolved in the usual manner, with the complete disappearance of the quinoline and the formation of the expected adducts. However, during purification of the crude mixture by chromatography, only the starting material **6g** was recovered in near-quantitative yield, demonstrating that in some cases this reaction can be reversible.

A rational interpretation of the factors affecting the stereoselectivity in the racemic series is not easy. It is clear from the experimental data that the attack of the nucleophile always takes place on the same face of the

heterocycle, since diastereoisomer **8** always prevailed. Use of force field calculations performed with CSC Chem 3D Plus for the simple trimethylsilyl ether derived from  $(\pm)$ -5a suggested that the C-4-substituent should preferentially adopt the conformation shown in Scheme 5. In it the benzylic hydrogen—the smaller substituent on the carbon bonded to the heterocycle—is directed toward the *peri* hydrogen, slightly tilted away from the aromatic plane.

Of the other two substituents, one is therefore nearly perpendicular to the aromatic ring. In this hypothesis it is reasonable that the bulkier  $-CH<sub>2</sub>OR$  group prefers to occupy this perpendicular position, and that the nucleophile will attack preferentially from the opposite side, as shown in transition state **10**. However, the extent of the d.r. cannot depend on steric effects alone. Actually, while the highest induction observed with bulky groups  $(SiPh_2t-Bu)$  and the best of all, triphenylmethyl) can be reasonably explained on steric grounds, it is not easy to understand the behavior of  $(\pm)$ -**5a** and  $(\pm)$ -**5g**. In the first case (entry 1) the reaction must occur on the alcoholate of  $(\pm)$ -**5a**, and this fact should be responsible for a different electronic environment in the transition state. On the other hand, either anisyl or *p*-nitrophenyl groups (entries 6 and 7) induced higher diastereoselectivities and this can probably not be attributed to steric factors, but only to stereoelectronic effects, that have not been clearly understood.

The diminished stereoselectivity observed in the additions to derivatives **6** is most likely a consequence of the fact that the two differently protected hydroxymethyl groups are more similar than the methyl and the –CH<sub>2</sub>OR of quinolines ( $\pm$ )-5, thus making the differentiation between the two faces of the heterocycle, based on steric hindrance, less pronounced. Also in this case stereoisomer **8** prevailed, with the bulkier  $-CH_2OCAr_3$  group lying nearly perpendicular to the aromatic ring.

Generally, although the stereoselectivity of these reactions is not impressive, it should be noted that they involve the formation of a new stereocenter that has a 1,4-relationship with the pre-existing one. Moreover, the first point of difference in the two oxygenated substituents of the stereogenic centers, that is the *O*protecting group, has a 1,7-relationship with the newly created center. This reaction therefore represents an example of long-range stereocontrol.<sup>35</sup>



The influence of the protecting groups on the stereoselectivity of nucleophilic additions to heterocycles is well known. Isobe's group observed good diastereoselectivity (up to 93:7) during the addition of acetylides to 4-substituted quinolines bearing an *O*protecting group in a 1,6-relationship with the newly created stereocenter.<sup>36</sup> Moreover, we obtained excellent results performing the same reaction on 3-substituted isoquinolines (with d.r. up to 95:5), bearing an *O*-protecting group in the same relationship with the newly formed stereogenic carbon at C-2.37

# **2.2. Relative configuration of compounds 8 and 9**

After our screening was complete we had to establish the relative configuration of compounds **8**,**9** and to correlate the addition products to demonstrate which epimer prevails during the addition of the acetylide.

To solve the first problem, we decided to transform both  $(\pm)$ -8*j* and  $(\pm)$ -9*j* into tricyclic derivatives, hopefully suitable for NOE studies. As a first goal we planned to synthesize a cyclic acetonide. For this purpose we exploited the epoxidation of  $(\pm)$ -8,9*j*, a reaction that is known to be highly stereoselective on this family of compounds, with the epoxide forming on the opposite side of the triple bond on C-2.38 Both compounds  $(\pm)$ -11,12 were obtained as a single diastereoisomer when (±)-**8j** and (±)-**9j** were separately treated with *m*-chloroperbenzoic acid (Scheme 6). Both derivatives were then reacted with *p*-toluenesulfonic acid in dry methanol. Under these conditions, the trityl group was removed and the oxirane underwent an acid catalyzed ring opening, with methanol acting as nucleophile. The structure of compounds  $(\pm)$ -13,14 was established on the basis of the following considerations: (1) the methoxy group must be bonded on benzylic carbon; actually, acetylating  $(\pm)$ -**13**, diacetate  $(\pm)$ -17 was isolated in which H<sub>3</sub> was shifted downfield as expected, passing from an alcoholic function to an acetoxy group (from 4.02 ppm as broad doublet to 5.30 ppm as doublet); (2) the *trans* relationship between OMe and OH was supposed on the basis of the opening mechanism of epoxides and confirmed later on acetonide  $(\pm)$ -16 and on lactones  $(\pm)$ -18,19; (3) the *trans* relationship between H<sub>2</sub> and  $H<sub>3</sub>$  was established by the coupling constants in the proton spectrum.

For the transformation of  $(\pm)$ -8**j** into a cyclic acetonide we first treated  $(\pm)$ -13 with NaHCO<sub>3</sub> in methanol to cleave the  $C$ -SiMe<sub>3</sub> bond. We were surprised to isolate as the major product, methyl carbamate  $(\pm)$ -15, which was transformed into acetonide (±)-**16** under standard conditions. Since we demonstrated that the transformation of phenyl carbamate into methyl carbamate does not occur during the treatment with acid, the only step responsible for the observed reaction must be the sec-**Scheme 5.** ond, although this reaction is peculiar just for this



# **Scheme 6.**

substrate.<sup> $\ddagger$ </sup> However, ( $\pm$ )-16 was found to be unsuitable for the determination of the relative configuration of all the four stereocenters using spectroscopic methods and this is most likely due to the fact that several conformations with comparable energy are present. NOEDIFF experiments on  $(t)$ -16 confirmed only that the junction between the two saturated rings must be *cis*. Finally, more useful information on the relative configuration of the stereocenters could be obtained from lactones (±)-**18**,**19**, obtained by selective oxidation of the primary alcoholic function of  $(\pm)$ -13 and  $(\pm)$ -14 with tetrapropylammonium perruthenate (TPAPP), to give a one-pot conversion into the tricyclic derivatives (±)-**18** and (±)-**19**. <sup>39</sup> Force field calculations on slightly simplified analogues, with H instead of  $\text{SiMe}_3$  and OMe instead of OPh (Scheme 7), helped us make some hypotheses on the three-dimensional structure of  $(\pm)$ -18,19, which were confirmed by <sup>1</sup>H NMR spectra (both chemical shift and coupling constants data) and NOEDIFF experiments. In particular, the information collected for both lactones, allowed us to make certain conclusions: (1) the  $C_{3a}$ -O bond is axial with respect to the nitrogen containing ring. A dihedral angle  $(C_{3a} - H_{3a} - C_4 - H_4)$  of 54–56° was predicted by calculations for both stereoisomers, which is in good agreement with the spectroscopic data obtained. Actually,  $H_4$  and  $H_{3a}$  gave two singlets at 5.10 and 5.73 ppm, respectively, in **18** and two doublets at 5.12 and 5.73 ppm, respectively, in **19** with a constant of 1.5 Hz; (2) the lactone ring is nearly planar; (3) one of the two substituents (H for **18** and Me for **19**) is directed toward the aromatic ring, causing a strong upfield shift



### **Scheme 7.**

<sup>‡</sup> The same reaction performed on an advanced intermediate in the synthesis of **1a**,**b** gave indeed the expected product (Ref. 45).

in proton spectrum with respect to the opposite situation [H (**18**): 2.52 ppm; H (**19**): 3.15 ppm; Me (**18**): 1.21 ppm; H (**19**): 0.77 ppm]; (4) a strong NOE is observed between  $H_{3a}$  and OMe as a consequence of the *cis* junction between the five- and the six-membered ring; (5) a moderate NOE effect is present between  $H_{3a}$  and Me  $\alpha$  to C=O in 18 and between H<sub>3a</sub> and H  $\alpha$  to C=O in **19**.

In conclusion, all the data obtained agree with the attribution of structure  $(\pm)$ -18 to the lactone derived from  $(\pm)$ -8**j** and  $(\pm)$ -19 to the lactone obtained from (±)-**9j**. The most striking evidence for the assignment of the relative stereochemistry to the carbon  $\alpha$  to C=O with respect to the other stereogenic centers is shown by the chemical shifts of the substituents attached to it. Since, during the transformation of  $(\pm)$ -8*j* and  $(\pm)$ -9*j* into  $(\pm)$ -18,19, the initial present stereogenic centers were not subjected to manipulations, the determination of the relative configuration of the two lactones allows us to attribute structure  $(\pm)$ -8j to the prevailing stereoisomer obtained by the addition of acetylides to  $(\pm)$ -5h and  $(\pm)$ -9j to the minor stereoisomer.

At this point we had only to correlate all major obtained diastereoisomers reported in Table 2 with (±)-**8j** and the minor diastereoisomers with (±)-**9j**. The correlation between  $(\pm)$ -8,9c,d,e and  $(\pm)$ -8,9i was quite simple and was based on the interconversion of protecting groups (Scheme 8). We established that  $(\pm)$ -8d can be obtained after trityl removal from (±)-**8j**, and (±)- **8c**,**e** were transformed into (±)-**8d**, by simple removal of the silyl protecting group.

Since a methyl ether requires drastic conditions for its removal, we preferred the methylation of (±)-**8d**, a reaction that was found to be unexpectedly troublesome. Many methods were tested (NaH, MeI; MeOTf, 2,6-lutidine;  $CH_2N_2$ ,  $SiO_2$ <sup>40</sup>, MeI,  $Ag_2O^{41}$  but ( $\pm$ )-8f was isolated in negligible amounts or did not form at all and, in the last case, only decomposition of the starting material was observed. Finally, using methyl triflate and 2,6-di-*t*-butyl-4-methylpyridine instead of 2,6-lutidine and working at 60°C, allowed the desired  $(\pm)$ -8f to be obtained in reasonable yield.<sup>42</sup>

Hydrolysis of the acetyl group in (±)-**8g** was not trivial, since basic conditions also cleaved the C-Si bond. The



alternative hydrolysis in the presence of a lipase (from *Candida antarctica* or porcine pancreas) was unsuccessful: while the latter enzyme did not accept this substrate at all, the former required the reaction to be carried out at 60°C or above§ before a very sluggish reaction began. For this reason we preferred to acetylate (±)-**8d** under standard conditions, thus obtaining (±)-**8g**.

Removal of the *p*-anisyl group under oxidative conditions (cerium ammonium nitrate) gave a faster competitive 1,2-oxidation, with the formation of a quinoline bearing the acetylenic moiety at C-2, followed by slower protecting group cleavage. In this case we correlated  $(\pm)$ -8h by transforming  $(\pm)$ -8d into the *O*-protected derivative under Mitsunobu conditions. However, the same reaction performed in order to prepare  $(\pm)$ -8**i** did not succeed and so the relative stereochemistry of that compound is assumed just by analogy with the very similar *p*-anisyl derivative. For compounds  $(\pm)$ -8–9a,**b** we did not demonstrate the relative configuration, although, on the basis of spectroscopic and chromatographic analogies, also in this case the relative stereochemistry of the prevailing diastereoisomer is most likely the same as the very similar (±)-**8c** derivative.

We chose to transform compounds (±)-**8**–**9k**,**l** into a common intermediate that can also be obtained from (±)-**8**,**9j** after homologation at the acetylenic carbon. Diastereomerically pure (±)-**8j** was transformed into alkyne (±)-**20**, which was treated with *t*-butylmagnesium chloride and paraformaldehyde (Scheme 9). The reaction was sluggish and very slow, affording, after trityl removal, diol  $(\pm)$ -26 in modest yield. On the other hand we tried to transform both diastereoisomeric mixtures (±)-**8**–**9k**,**l** into chromatographically separable diols (±)-**26**,**27**. Both epimeric mixtures were separately treated with boron trifluoride etherate in the presence of ethanethiol, a method known to give cleavage of benzyl-, *p*-methoxybenzyl- and trityl ethers.<sup>43</sup> While the mixture composed by (±)-**8**,**9l** was transformed, albeit in moderate yield into  $(\pm)$ -26,27, under the same conditions (±)-**8**,**9k** underwent only trityl cleavage in excellent yield. The benzyl group could be removed only forcing the conditions, that is working at reflux, but in this case the reaction was not clean. For  $(\pm)$ -8,9l a two-step sequence as summarized in Scheme 9 gave better results. Since  $(\pm)$ -26 and  $(\pm)$ -27 are readily separated, we ascertained that also in this series  $(\pm)$ -26, which has been also obtained from (±)-**8j**, prevailed.

As the final step we had to correlate compounds obtained by nucleophilic addition to **6b**,**c**,**f** with **8**,**9**. We easily demonstrated that the same diastereoisomer predominated, by cleaving the triethylsilyl ether of the mixture **8**,**9o** and acetylating the prevailing diastereoiso-

 $§$  This enzyme can usually work at this temperature without losing its activity (Ref. 37).



**Scheme 9.**

mer of the free alcohol, that is **28** to give **8m** (Scheme  $10$ .





On the other hand, the reductive cleavage of one of the two C-O bonds permitted conversion of **29** into (−)-**9j** (Scheme 11). This reaction was quite difficult and many usually employed conditions (NaBH<sub>4</sub>/DMSO, Red-Al®, Super-Hydride®) failed. Finally, the reaction succeeded under radical conditions with tri-*n*-butyltin hydride in the presence of sodium iodide. The choice of solvent was crucial: when commonly employed 1,2dimethoxyethane was used<sup>44</sup> 9*j* was not obtained, while using THF and working in a sealed tube at 85°C, compound **9j** could be isolated, albeit in moderate yield.



**Scheme 11.**

## **3. Conclusions**

In this paper we have reported a new example of diastereoselective functionalization of quinoline derivatives, in which the d.r. is mostly influenced by the nature of a *O*-protecting group. The reaction has been performed on racemic substrates like **5** and enantiomerically pure compounds like **6** to give polyfunctionalized derivatives, useful intermediates for more complex molecules of potential biological activity. Part of this project has already been realized in our group,<sup>45</sup> while other studies are currently under investigation and will be presented in due course.

#### **4. Experimental**

# **4.1. General**

NMR spectra were taken in CDCl<sub>3</sub>, at 200 MHz  $(^1H)$ , and 50 MHz  $(^{13}C)$ , using TMS as internal standard. Chemical shifts are reported in ppm ( $\delta$  scale), coupling constants are reported in hertz. Peak assignment in <sup>1</sup>H

<sup>¶</sup> The spectroscopic analogies between **8**,**9m** and **8**,**9n** allowed us to suggest **8n** as the most likely structure for the prevailing diastereoisomer obtained by acetylide addition to **6c**.

NMR spectra was also made with the aid of double resonance experiments. In ABX systems, the proton A is considered downfield and B upfield. Peak assignment in  $^{13}$ C spectra was made with the aid of DEPT experiments. GC–MS were carried out on a HP-5971A instrument, using an HP-1 column (12 m long, 0.2 mm wide), electron impact at 70 eV, and a mass temperature of about 170°C. Unless otherwise indicated analyses were performed with a constant He flow of 0.9 ml/min, init. temp. 100°C, init. time 2 min, rate 20°C/min, final temp. 260°C, final time 4 min, inj. temp. 250°C, det. temp. 280°C. Diastereomeric ratios were determined by GLC using a Carlo Erba FRACTOVAP gas chromatograph equipped with a RSL-150 or SE-54 (both 25 m long, 0.25 mm wide) column or by HPLC using a Waters mod. 6000 instrument with an absorbance detector mod. 440 (at 245 nm) and equipped with an Erbasil column (10  $\mu$ m, 250 mm×4.6 mm).  $R_t$  are in min. IR spectra were measured with a Perkin–Elmer 881 instrument as CHCl<sub>3</sub> solutions. Values of  $[\alpha]_D$  were determined on a Jasco DIP 181 polarimeter, in CHCl<sub>3</sub> (containing 0.75–1% EtOH) solution. Melting points were measured on a Büchi 535 apparatus and are uncorrected. TLC analyses were carried out on silica gel plates, which were developed by these detection methods: (**A**) UV; (**B**) dipping into a solution of  $(NH_4)_4MoO_4·4H_2O$  (21 g) and  $Ce(SO_4)_2·4H_2O$  (1 g) in  $H_2SO_4$  (31 ml) and  $H_2O$  (469 ml) and warming.  $R_f$  were measured after an elution of 7–9 cm. Chromatographies were carried out on 220–400 mesh silica gel (if not otherwise specified) using the 'flash' methodology. Petroleum ether (40–60°C) is abbreviated as PE. In extractive work-up, aqueous solutions were always reextracted thrice with the appropriate organic solvent. Organic extracts were washed with brine, dried over  $Na<sub>2</sub>SO<sub>4</sub>$  and filtered, before evaporation of the solvent under reduced pressure. All reactions employing dry solvents were carried out under a nitrogen atmosphere. The purity of all compounds was established by TLC, 1 H NMR, GC–MS. Lipase from recombinant *C*. *antarctica* was a kind gift from Novo Nordisk, while PLE was purchased from Fluka.

#### **4.2. 4-Ethylquinoline, 7**

A solution of 1-chloro-3-pentanone  $(5.00 \text{ g}, 85\%, 35.25)$ mmol) in nitrobenzene (10 ml) was treated with freshly distilled aniline (6.42 ml, 70.50 mmol) and concentrated hydrochloric acid (2.94 ml, 35.25 mmol) and then stirred at 100°C for 18 h. The solution was diluted with  $Et<sub>2</sub>O$  and extracted trice with 1N HCl. The aqueous extract was treated with 6N NaOH until pH 10 was reached. After filtration over a Celite pad, an usual extraction with  $Et<sub>2</sub>O$  was performed. A rapid filtration over silica gel with  $PE \rightarrow PE/Et_2O$ , 8:2, then  $Et_2O$ , followed by distillation under reduced pressure, gave compound **7** in 41–49% overall yield as a colorless oil. Bp: 83 $^{\circ}$ C, 0.9 mbar.  $R_f$  0.31 (PE/Et<sub>2</sub>O 1:1, A). GC–MS (usual conditions, but with init. temp.  $70^{\circ}$ C):  $R_{t}$  6.75; *m*/*z* 158 (12), 157 (M<sup>+</sup> , 100), 154 (11), 143 (5.5), 142 (52), 130 (5.9), 129 (7.3), 128 (8.4), 116 (5.5), 115 (22), 89 (5.1), 77 (7.8), 51 (5.4), 40 (6.3). <sup>1</sup> H NMR: 1.40 [3H, t, >CHCH<sub>3</sub>,  $J=7.6$ ]; 3.13 [2H, q,  $-CH_2CH_3$ ,  $J=7.6$ ];

7.26 [1H, d, H<sub>3</sub>, J = 4.7]; 7.56 [1H, ddd, H<sub>6</sub>, J = 1.4, 6.8, 8.3]; 7.70 [1H, ddd,  $H_7$ ,  $J=1.5$ , 6.9, 8.3]; 8.05 [1H, dd, *H*5, *J*=1.1, 8.4]; 8.12 [1H, dd, *H*8, *J*=1.1, 8.4]; 8.82  $[H, d, H<sub>2</sub>, J=4.5].$ 

## **4.3. 2-Quinolin-4-yl-propan-1-ol, 5a**

Ethylquinoline **7** (10.0 g, 63.61 mmol) was refluxed with aqueous formaldehyde (37%, 19 ml, 254.4 mmol) for 15.5 h. Excess formaldehyde was removed under reduced pressure and the residue was taken up with methanol and concentrated again. The residue was dissolved in MeOH and stirred under nitrogen at rt in the presence of anhydrous  $K_2CO_3$  (2.5 g) for 20 h. After filtration of the carbonate, the solution was concentrated under reduced pressure, diluted with water and extracted as usual with  $Et<sub>2</sub>O$ . After solvent removal, crude product was chromatographed with PE/AcOEt,  $2:8 \rightarrow$  AcOEt and then AcOEt/EtOH, 97:3, to give pure **5a** as a pale yellow oil (10.59 g, 89%).  $R_f$  0.16 (Et<sub>2</sub>O, A). IR:  $v_{\text{max}}$  3612, 2964, 1591, 1255, 1021. GC–MS (usual conditions, but with init. temp. 80°C):  $R_t$  8.21;  $m/z$  188 (5.6), 187 (M<sup>+</sup> , 42), 157 (42), 156 (100), 155 (11), 154 (30), 130 (7.5), 129 (8.6), 128 (14), 127 (7.8), 101 (5.2), 77 (6.4). <sup>1</sup> H NMR: 1.45 [3H, d, >CHC*H*3, *J*=6.5]; 3.82–4.04 [3H, m, >CHCH<sub>2</sub>OH]; 7.32 [1H, d,  $H_3$ ,  $J=$ 4.5]; 7.57 [1H, ddd,  $H_6$ ,  $J=1.1$ , 6.9, 8.2]; 7.70 [1H, ddd,  $H_7$ ,  $J=1.4$ , 6.9, 8.5]; 8.11 [1H, dd,  $H_5$ ,  $J=1.1$ , 7.9]; 8.13 [1H, dd,  $H_8$ ,  $J=1.1$ , 8.1]; 8.84 [1H, d,  $H_2$ ,  $J=4.6$ ]. <sup>13</sup>C NMR: 17.33 [>CH*C*H3]; 36.44 [>*C*HCH3]; 67.20 [–*C*H2O–]; 118.12 [*C*3]; 123.06 [*C*5]; 126.30 [*C*6]; 127.40  $[C_{4a}]$ ; 128.92 and 129.53 [2C,  $C_7$  and  $C_8$ ]; 147.67 [ $C_4$ ]; 149.44 [C<sub>2</sub>]; 150.9 [C<sub>8a</sub>].

## **4.4. Silylation of 5a**

A solution of **5a** (1.00 g, 5.34 mmol) in dry DMF (12 ml) was treated with imidazole (618 mg, 9.08 mmol) and  $R_2t$ BuSiCl and stirred at rt for 1.5–2.5 h. The solution was diluted with water/NaHCO<sub>3</sub> sat. solution  $(1:1)$  and extracted with Et<sub>2</sub>O. The crude product was purified by chromatography using  $PE/ACOE$  7:3 $\rightarrow$ 1:1 to give the corresponding products **5b** (87% yield) and **5c** (84% yield) as colorless oils.

**4.4.1. 4-{[2-(***tert***-Butyldimethylsilyloxy)-1-methyl]ethyl} quinoline, 5b.**  $R_f$  0.45 (PE/Et<sub>2</sub>O 1:1, A). IR:  $v_{\text{max}}$  2829, 2883, 1244, 1092. GC–MS: *R*<sup>t</sup> 8.57; *m*/*z* 301 (M<sup>+</sup> , 0.12), 246 (5.6), 245 (21), 244 (100), 214 (6.6), 170 (6.8), 154 (7.3), 89 (5.4), 75 (16), 73 (19), 59 (5.6). <sup>1</sup> H NMR: −0.07 and −0.08 [6H, 2 s, >Si(CH<sub>3</sub>)<sub>2</sub>]; 0.81 [9H, s, −C(CH)<sub>3</sub>]; 1.43 [3H, d, >CHC*H*3, *J*=6.6]; 3.72–3.94 [3H, m, >C*H*C*H*2O–]; 7.31 [1H, d, *H*3, *J*=4.7]; 7.56 [1H, ddd, *H*6, *J*=1.4, 6.8, 8.4]; 7.70 [1H, ddd, *H*7, *J*=1.4, 6.9, 8.2]; 8.12 [1H, dd,  $H_5$  or  $H_8$ ,  $J=1.1$ , 8.1]; 8.14 [1H, dd, *H*<sub>5</sub> or *H*<sub>8</sub>, *J*=1.1, 8.1]; 8.85 [1H, d, *H*<sub>2</sub>, *J*=4.6]. <sup>13</sup>C NMR: −5.51 and −5.48 [2C, >Si(CH<sub>3</sub>)<sub>2</sub>]; 17.24  $[\neg \text{CHCH}_3]$ ; 18.24  $[\neg \text{C}(\text{CH}_3)_3]$ ; 25.82  $[\text{3C}, \neg \text{C}(\text{CH}_3)_3]$ ; 36.25 [>*C*HCH3]; 68.04 [–*C*H2O–]; 118.36 [*C*3]; 123.20 [*C*5]; 126.18 [*C*6]; 127.48 [*C*4a]; 128.80 and 130.23 [2C, *C*<sup>7</sup> and *C*8]; 148.25 [*C*4]; 150.00 [*C*2]; 150.58 [*C*8a].

**4.4.2. 4-{[2-(***tert***-Butyldiphenylsilyloxy)-1-methyl]ethyl} quinoline, 5c.**  $R_f$  0.54 (PE/Et<sub>2</sub>O 2:8, A). IR:  $v_{\text{max}}$  2930, 2859, 1729, 1590, 1462, 1374, 1240, 1105. GC–MS: *R*<sup>t</sup> 12.67; *m*/*z* 425 (M<sup>+</sup> , 0.059), 370 (8.1), 369 (31), 368 (100), 290 (19), 199 (20), 184 (6.4), 181 (8.6), 154 (5.6), 135 (5.0). <sup>1</sup> H NMR: 0.95 [9H, s, –C(C*H*)3], 1.47 [3H, d,  $\triangleright$ CHC*H*<sub>3</sub>, *J*=6.4]; 3.75–3.91 [3H, m,  $\triangleright$ CHC*H*<sub>2</sub>O–]; 7.20–7.57 [12H, m, H<sub>3</sub>, H<sub>6</sub>, aromatics of TBDPS], 7.67 [1H, ddd,  $H_7$ ,  $J=1.3$ , 6.9, 8.3]; 7.85 [1H, d,  $H_5$ ,  $J=8.4$ ]; 8.10 [1H, d,  $H_8$ ,  $J=8.0$ ]; 8.82 [1H, d,  $H_2$ ,  $J=4.6$ ]. <sup>13</sup>C NMR: 17.08 [>CHCH<sub>3</sub>]; 19.18 [–C(CH<sub>3</sub>)<sub>3</sub>]; 26.75 [3C, –C(*C*H3)3]; 36.17 [>*C*HCH3]; 68.60 [–*C*H2O–]; 118.54 [*C*3]; 123.17 [*C*5]; 126.23 [*C*6]; 127.50 [*C*4a]; 127.61 and 127.67 [4C, *C meta* of TBDPS]; 128.80 and 130.24 [2C, *C*<sup>7</sup> and *C*8]; 129.61 and 129.71 [2C, *C para* of TBDPS]; 133.31 and 133.44 [2C, *C ipso* of TBDPS]; 135.50 and 135.61 [4C, *C ortho* of TBDPS]; 148.31 [*C*4]; 150.03  $[C_2]$ ; 150.27  $[C_{\text{sa}}]$ .

# **4.5. 4-[(2-Methoxy-1-methyl)ethyl]quinoline, 5d**

A solution of **5a** (520 mg, 2.78 mmol) in dry THF was cooled to  $0^{\circ}$ C and treated with methyl iodide (346  $\mu$ l, 5.55 mmol) and NaH (60% in mineral oil, 167 mg, 4.17 mmol). After 10 min the reaction was allowed to stir at rt for 24 h (after 3 h an equivalent amount of both reagents was added). The mixture was partitioned between  $NH<sub>4</sub>Cl$  sat. sol. and AcOEt and extracted as usual. Chromatography with  $PE/Et_2O$ ,  $8:2 \rightarrow Et_2O$  gave **5d** as a pale yellow oil (324 mg, 58%).  $R_f$  0.48 (Et<sub>2</sub>O, A). IR: max 2928, 2876, 1590, 1570, 1450, 1214, 1108. GC–MS:  $R_t$  6.68;  $m/z$  202 (6.5), 201 (M<sup>+</sup>, 46), 200 (27), 170 (18), 157 (8.0), 156 (25), 155 (5.4), 154 (15), 128 (8.1), 127 (5.2), 45 (100). <sup>1</sup> H NMR: 1.44 [3H, d, >CHC*H*3, *J*=6.8]; 3.36 [3H, s, –OC*H*3]; 3.57 and 3.72 [2H, AB part of ABX system,  $> CHCH_2O-, J_{AB}=9.2$ ,  $J_{AX}$  and  $J_{BX} = 5.8$ , 7.2]; 3.91 [1H, sextuplet, >C*H*CH2O–, *J*=6.7]; 7.31 [1H, d, *H*3, *J*=4.7]; 7.58 [1H, ddd,  $H_6$ ,  $J=1.5$ , 6.9, 8.3]; 7.71 [1H, ddd,  $H_7$ ,  $J=1.3, 6.9, 8.5$ ; 8.13 [2H, dd,  $H_5$  and  $H_8$ ,  $J=1.1, 8.4$ ]; 8.87 [1H, d,  $H_2$ ,  $J=4.7$ ]. <sup>13</sup>C NMR: 18.01 [>CHCH<sub>3</sub>]; 34.04 [>*C*HCH3]; 59.00 [–O*C*H3]; 77.28 [–*C*H2O–]; 118.08 [*C*3]; 122.93 [*C*5]; 126.40 [*C*6]; 127.26 [*C*4a]; 128.94 and 130.40 [2C,  $C_7$  and  $C_8$ ]; 148.43 [ $C_4$ ]; 150.19 [ $C_2$ ]; 150.25  $[C_{8a}]$ .

## **4.6. Acetic acid 2-quinolin-4-yl-propyl ester, 5e**

A solution of **5a** (537 mg, 2.87 mmol) in dry pyridine (3 ml) was stirred at rt in the presence of acetic anhydride  $(541 \mu l, 5.74 \mu)$  for 2.5 h. The solution was partitioned between water/ $Et<sub>2</sub>O$  and extracted as usual. After solvent removal, residue pyridine was azeotropically removed with heptane. Chromatography with PE/ AcOEt  $1:1 \rightarrow 4:6$  gave **5e** as a pale yellow oil (631 mg, 96%). *R<sub>f</sub>* 0.55 (PE/AcOEt 1:1, A). IR:  $v_{\text{max}}$  3005, 3962, 1728, 1591, 1374, 1191. GC–MS: *R*<sup>t</sup> 7.44; *m*/*z* 229 (M<sup>+</sup> , 14), 170 (11), 169 (66), 168 (15), 157 (35), 156 (91), 155 (15), 154 (79), 130 (6.6), 129 (7.7), 128 (11), 127 (8.8), 101 (6.5), 77 (7.3), 51 (5.6), 43 (100). <sup>1</sup> H NMR: 1.46 [3H, d, >CHC $H_3$ ,  $J=6.9$ ]; 2.00 [3H, s,  $-COCH_3$ ]; 4.02 [1H, sextuplet,  $>CHCH<sub>2</sub>O-$ ,  $J=6.9$ ]; 4.25 and 4.43 [2H, AB part of ABX system,  $>CHCH<sub>2</sub>O-$ ,  $J<sub>AB</sub>=10.9$ ,  $J<sub>AX</sub>$  and  $J_{\text{BX}}=6.6, 7.1$ ]; 7.32 [1H, d,  $H_3$ ,  $J=4.7$ ]; 7.60 [1H, ddd,  $H_6$ ,  $J=1.3$ , 6.9, 8.5]; 7.73 [1H, ddd,  $H_7$ ,  $J=1.4$ , 6.8, 8.1]; 8.15 [2H, dd,  $H_5$  and  $H_8$ ,  $J=1.6$ , 8.5]; 8.89 [1H, d,  $H_2$ ,  $J=4.6$ ]. <sup>13</sup>C NMR: 17.52 [>CHCH<sub>3</sub>]; 20.77 [–CO*C*H3]; 32.90 [>*C*HCH3]; 68.18 [–*C*H2O–]; 117.95  $[C_3]$ ; 122.62  $[C_5]$ ; 126.59  $[C_6]$ ; 127.14  $[C_{4a}]$ ; 129.08 and 130.41 [2C,  $C_7$  and  $C_8$ ]; 148.41 and 148.95 [2C,  $C_4$  and *C*8a]; 150.12 [*C*2]; 170.90 [*C*O].

## **4.7. Preparation of O-Ar derivatives of 5a under Mitsunobu's conditions**

A solution of  $5a$  (350 mg, 1.87 mmol) in dry CH<sub>2</sub>Cl<sub>2</sub> (15 ml) was treated at rt with PPh<sub>3</sub> (637 mg, 2.43) mmol), the appropriate phenol (*p*-methoxyphenol for **5f** and *p*-nitrophenol for **5g**, 5.60 mmol) and diethyl azodicarboxylate  $(382 \text{ µl}, 2.43 \text{ mmol})$ . The solution was stirred for 1 h (**5f**) and 24 h (**5g**) with the addition of an equivalent amount of the reagents after 7 h. After quenching with  $NH<sub>4</sub>Cl$  sat. sol. the mixture was extracted with AcOEt. The organic layers were rapidly washed with 1N aqueous NaOH and brine until  $pH \approx 8$ was reached. After solvent removal, chromatography with the appropriate  $PE/Et_2O$  mixture gave 5f (77%) yield, colorless oil) and **5g** (76% yield, pale yellow oil).

**4.7.1. 4-{[2-(***p***-Methoxyphenoxy)-1-methyl]ethyl}quinoline, 5f**. *R*<sub>f</sub> 0.49 (PE/AcOEt 1:1, A). IR:  $v_{\text{max}}$  2960, 1733, 1590, 1496, 1464, 1191, 1028. GC–MS: *R*<sup>t</sup> 10.52; *m*/*z* 294 (12), 293 (M<sup>+</sup> , 60), 171 (14), 170 (100), 168 (5.8), 155 (15), 154 (25), 124 (61), 123 (5.4, 109 (18). <sup>1</sup> H NMR: 1.55 [3H, d, >CHC*H*3, *J*=6.6]; 3.75 [3H, s, -OCH<sub>3</sub>]; 4.08 [1H, center of m, >CHCH<sub>2</sub>O–]; 4.17 and 4.23 [2H, AB part of ABX system,  $\geq$ CHCH<sub>2</sub>O–,  $J_{AB}$ = 6.8,  $J_{AX}$  and  $J_{BX}$  = 2.0, 4.1]; 6.80 [4H, s, *p*-anisyl]; 7.36 [1H, d,  $H_3$ ,  $J=4.4$ ]; 7.58 [1H, ddd,  $H_6$ ,  $J=1.6, 7.0, 8.4$ ]; 7.72 [1H, ddd,  $H_7$ ,  $J=1.2$ , 6.6, 8.4]; 8.15 [2H, d,  $H_5$  and  $H_8$ ,  $\bar{J}$ =8.8]; 8.88 [1H, d,  $H_2$ ,  $J$ =4.8]. <sup>13</sup>C NMR: 17.75 [>CH*C*H3]; 33.75 [>*C*HCH3]; 55.60 [–O*C*H3]; 72.62 [–*C*H2O–]; 114.58 [2C, *C ortho* to –OMe]; 115.50 [2C, *C meta* to –OMe]; 118.13 [C<sub>3</sub>]; 122.87 [C<sub>5</sub>]; 126.44 [C<sub>6</sub>]; 127.12 [*C*4a]; 128.95 and 130.33 [2C, *C*<sup>7</sup> and *C*8]; 148.35 and 149.55 [2C, C<sub>4</sub> and C<sub>8a</sub>]; 150.10 [C<sub>2</sub>]; 152.77 and 153.95 [2C, *C* quat. of *p*-anisyl].

**4.7.2. 4-[1-Methyl-2-(***p***-nitrophenoxy)ethyl]quinoline, 5g**. *R*<sub>f</sub> 0.67 (AcOEt 1:1, A). IR:  $v_{\text{max}}$  2966, 1592, 1497, 1342, 1193, 1111, 1016. GC–MS: *R*<sup>t</sup> 12.37; *m*/*z* 309 (6.1), 308 (M<sup>+</sup> , 30), 171 (13), 170 (100), 169 (6.7), 168 (7.5), 156 (19), 155 (25), 154 (35), 143 (7.4), 142 (6.5), 128 (8.4), 127 (5.0). <sup>1</sup> H NMR: 1.60 [3H, d, >CHC*H*3, *J*=6.5]; 4.07–4.41 [3H, m, >C*H*C*H*2O–]; 6.91 [2H, dt, *H* meta to  $NO_2$ ,  $J=2.8$ , 9.3]; 7.37 [1H, d,  $H_3$ ,  $J=4.5$ ]; 7.62 [1H, ddd,  $H_6$ ,  $J=1.4$ , 6.9, 8.3]; 7.75 [1H, ddd,  $H_7$ ,  $J=1.3$ , 6.8, 8.3]; 8.14 [2H, dd,  $H_5$  and  $H_8$ ,  $J=1.1$ , 8.2]; 8.18 [2H, dt, *H* ortho to NO<sub>2</sub>, *J*=2.8, 9.2]; 8.90 [1H, d,  $H_2$ ,  $J=4.6$ ]. <sup>13</sup>C NMR: 17.66 [>CHCH<sub>3</sub>]; 33.54 [>*C*HCH3]; 72.77 [–*C*H2O–]; 114.44 [2C, *C meta* to –NO2]; 118.15 [*C*3]; 122.63 [*C*5]; 125.89 [2C, *C ortho* to –NO2]; 126.77 [*C*6]; 128.91 [*C*4a]; 129.24 and 130.60 [2C, *C*<sup>7</sup> and *C*8]; 141.75 [CNO2]; 148.49 and 148.61 [2C, *C*<sup>4</sup> and *C*<sub>8a</sub>]; 150.18 [*C*<sub>2</sub>]; 163.58 [*C para* to -NO<sub>2</sub>].

## **4.8. 4-[1-Methyl-2-(trityloxy)ethyl]quinoline, 5h**

A solution of  $5a$  (493 mg, 2.63 mmol) in dry CH<sub>2</sub>Cl<sub>2</sub> (20 ml) was treated with dry pyridine  $(317 \mu l, 3.94)$ mmol) and trityl chloride (807 mg, 2.89 mmol) and stirred at rt for 22 h. The reaction was partitioned between NH<sub>4</sub>Cl sat. sol. and AcOEt and then extracted. Residue pyridine was removed azeotropically with heptane and the crude mixture was purified by chromatography with PE/AcOEt 7:3 $\rightarrow$ 6:4 to give a white foam  $(1.131 \text{ g}, 96\%)$ , which crystallized from Et<sub>2</sub>O to give a white solid. Mp 124.3–125.9°C (Et<sub>2</sub>O).  $R_f$  0.68 (PE/ AcOEt 1:1, **A**, **B**). IR:  $v_{\text{max}}$  2969, 1591, 1487, 1449, 1189, 1070. GC–MS: (usual conditions, but with fin. temp. 290°C)  $R_1$  14.21;  $m/z$  429 (M<sup>+</sup>, 0.20), 244 (22), 243 (100), 170 (8.1), 165 (24), 154 (6.2), 105 (7.9), 77 (5.1). <sup>1</sup> H NMR: 1.45 [3H, d, >CHC*H*3, *J*=6.9]; 3.34 and 3.41 [2H, AB part of ABX system,  $\geq$ CHC*H*<sub>2</sub>O–,  $J_{AB} = 9.1$ ,  $J_{AX}$  and  $J_{BX} = 6.2$ , 7.2]; 3.91 [1H, sextuplet, >C*H*CH<sub>2</sub>O–, *J*=6.7]; 7.17–7.34 [16H, m, *H*<sub>3</sub> and aromatics of Tr]; 7.52 [1H, ddd,  $H_6$ ,  $J=1.2$ , 6.9, 8.3]; 7.70 [1H, ddd,  $H_7$ ,  $J=1.4$ , 6.7, 8.3]; 8.02 [1H, d,  $H_5$  or  $H_8$ ,  $J=8.4$ ]; 8.12 [1H, d,  $H_5$  or  $H_8$ ,  $J=8.0$ ]; 8.80 [1H, d,  $H_2$ , *J*=4.7]. 13C NMR: 17.81 [>CH*C*H3]; 34.36 [>*C*HCH3]; 68.21 [–*C*H2O–]; 86.59 [–*C*Ph3]; 118.32 [*C*3]; 123.26 [*C*5]; 126.16 [*C*6]; 126.90 [3C, *C para* of Tr]; 127.42 [*C*4a]; 127.66 and 128.59 [12C, *C ortho* and *meta* of Tr]; 128.81 and 130.26 [2C, *C*<sup>7</sup> and *C*8]; 143.90 [3C, *C ipso* of Tr]; 148.32 [*C*4]; 150.06 [*C*2]; 150.58 [*C*8a].

## **4.9. Triarylmethyl derivatives of 6a**

The same procedure described above for the preparation of **5h** was followed, starting from **6a** and using TrCl at reflux, to give **6b** (100% yield) as a white foam and MMTrCl at rt to give **6c** (95% yield) as a white foam or DMTrCl at rt to give **6d** (85% yield) as a yellow foam.

**4.9.1. (***S***)-Acetic acid 2-(quinolin-4-yl)-3-(trityloxy) propyl ester, 6b**.  $R_f$  0.35 (PE/Et<sub>2</sub>O 2:8, A, B).  $[\alpha]_D =$ +31.4 (*c* 2.06, CHCl<sub>3</sub>). IR:  $v_{\text{max}}$  2961, 1736, 1449, 1194, 1070, 1033. GC–MS: (usual conditions, but with fin. temp. 290°C) *R*<sup>t</sup> 16.52; *m*/*z* 410 (M<sup>+</sup> −77, 0.095), 244 (24), 243 (100), 228 (9.5), 186 (5.2), 168 (9.9), 167 (5.0), 166 (5.6), 165 (32), 154 (9.2), 105 (14), 77 (7.9), 43 (27). <sup>1</sup>H NMR: 1.91 [3H, s, -COCH<sub>3</sub>]; 3.52 [2H, d,  $-CH_2$ OTr,  $J = 5.8$ ]; 4.16 [1H, quintuplet,  $-CH(CH_2O-)_{2}$ , *J*=6.2]; 4.58 [2H, d, –C*H*2OAc, *J*=6.8]; 7.16–7.35 [16H, m,  $H_3$  and aromatics of Tr]; 7.54 [1H, ddd,  $H_6$ , *J*=1.3, 6.3, 8.3]; 7.72 [1H, ddd, *H*7, *J*=1.3, 6.9, 8.3]; 7.98 [1H, broad d,  $H_5$ ,  $J=8.4$ ]; 8.13 [1H, broad d,  $H_8$ , *J*=8.4]; 8.82 [1H, d, H<sub>2</sub>, *J*=4.6]. <sup>13</sup>C NMR: 20.73  $[-C OCH_3]$ ; 39.18  $[-CH(CH_2O-)_2]$ ; 63.53 and 64.27 [2C,  $-CH(CH, O-)$ ; 86.92 [–*C*Ph<sub>3</sub>]; 119.16 [*C*<sub>3</sub>]; 122.88 [*C*<sub>5</sub>]; 126.59 [*C*6]; 127.04 [3C, *C para* of Tr]; 127.36 [*C*4a]; 127.76 and 128.51 [12C, *C ortho* and *meta* of Tr]; 129.07 and 130.39 [2C, *C*<sup>7</sup> and *C*8]; 143.53 [3C, *C ipso* of Tr]; 145.43 [*C*4]; 148.40 [*C*8a]; 149.82 [*C*2]; 170.81 [*C*O].

**4.9.2. (***S***)-Acetic acid 3-{[(***p***-methoxyphenyl) diphenyl]methoxy}-2-(quinolin-4-yl)propyl ester, 6c**. *R*<sup>f</sup> 0.30 (PE/Et<sub>2</sub>O 2:8, **A**, **B**).  $[\alpha]_D = +30.4$  (*c* 1.08, CHCl<sub>3</sub>). IR:  $v_{\text{max}}$  2953, 1732, 1591, 1075, 1030. GC–MS: (usual conditions, but with fin. temp. 290°C)  $R_1$  18.73;  $m/z$  517 (M<sup>+</sup> , 2.9), 273 (100), 228 (11), 195 (5.2), 186 (5.3), 168 (10), 165 (8.9), 154 (6.2), 105 (12), 77 (5.4), 43 (18). <sup>1</sup> H NMR: 1.92 [3H, s, –COC*H*3]; 3.52 [2H, d, –C*H*2OMMTr, *J*=5.8]; 3.77 [3H, s, –OC*H*3]; 4.11 [1H, quintuplet,  $-CH(CH_2O_{2})$ ,  $J=6.2$ ]; 4.59 [2H, d,  $-CH<sub>2</sub>OAc$ ,  $J=6.6$ ]; 6.76 [2H, dt, *H ortho* to  $-OMe$ ,  $J=3.0, 9.2$ ; 7.14–7.36 [13H, m,  $H_3$  and aromatics of MMTr]; 7.55 [1H, ddd,  $H_6$ ,  $J=1.6$ , 7.0, 8.4]; 7.72 [1H, ddd,  $H_7$ ,  $J=1.4$ , 7.0, 8.4]; 7.98 [1H, broad d,  $H_5$ , *J*=8.0]; 8.13 [1H, dd, *H*8, *J*=1.2, 8.6]; 8.82 [1H, d, *H*2, *J*=4.8]. 13C NMR: 20.73 [–CO*C*H3]; 39.17  $[-CH(CH, O<sub>-</sub>)<sub>2</sub>]$ ; 55.11  $[-OCH<sub>3</sub>]$ ; 63.42 and 64.29 [2C, –CH(*C*H2O–)2]; 86.60 [–*C*Ph2Ar]; 112.99 [2C, *C ortho* to OMe]; 119.16 [*C*3]; 122.88 [*C*5]; 126.56 [*C*6]; 126.86 [2C, *C para* of Ph]; 127.35 [*C*4a]; 127.73 and 128.19 [8C, *C* ortho and *meta* of Ph]; 129.05 and 130.33 [2C,  $C_7$  and *C*8]; 130.24 [2C, *C meta* to OMe]; 135.11 [*C para* to OMe]; 144.04 [2C, *C ipso* of Ph]; 145.49 [*C*4]; 148.36 [*C*8a]; 149.80 [*C*2]; 158.49 [*C*OMe of MMTra]; 170.82 [*C*O].

**4.9.3. (***S***)-Acetic acid 3-{[bis-(***p***-methoxyphenyl)phenyl] methoxy**}-2-(quinolin-4-yl)propyl ester, 6d.  $R_f$  0.42 (PE/ Et<sub>2</sub>O 2:8, **A**, **B**).  $[\alpha]_D = +33.0$  (*c* 1.31, CHCl<sub>3</sub>). IR:  $v_{\text{max}}$ 2953, 1730, 1604, 1299, 1196, 1029. GC–MS: (usual conditions, but with final temp. 290°C)  $R_1$ , 23.16;  $m/z$ 547 (M<sup>+</sup> , 0.79), 303 (100), 228 (9.2), 186 (5.6), 168 (10), 155 (7.3), 154 (9.6), 135 (12), 105 (13), 77 (6.2), 43 (24). <sup>1</sup>H NMR: 1.92 [3H, s, –COCH<sub>3</sub>]; 3.51 [2H, d, –C*H*2ODMTr, *J*=5.4]; 3.77 [6H, s, –OC*H*3]; 4.10 [1H, quintuplet,  $-CH(CH_2O_{2})$ ,  $J=6.2$ ]; 4.58 [2H, d, –C*H*2OAc, *J*=6.6]; 6.75 [4H, apparent dd, *H ortho* to  $-OMe$ ,  $J=1.4$ , 8.8]; 7.12–7.34 [10H, m,  $H_3$  and aromatics of DMTr]; 7.54 [1H, ddd,  $H_6$ ,  $J=1.4$ , 7.0, 8.4]; 7.72 [1H, ddd,  $H_7$ ,  $J=1.4$ , 7.0, 8.0]; 7.98 [1H, broad d,  $H_5$ , *J*=8.4]; 8.13 [1H, dd, *H*<sub>8</sub>, *J*=1.2, 8.4]; 8.82 [1H, d, *H*<sub>2</sub>, *J*=4.4]. <sup>13</sup>C NMR: 20.74 [-COCH<sub>3</sub>]; 39.20 <sup>13</sup>C NMR: 20.74 [–COCH<sub>3</sub>]; 39.20  $[-CH(CH, O<sub>-</sub>)<sub>2</sub>]$ ; 55.12 [2C,  $-OCH<sub>3</sub>$ ]; 63.36 and 64.34  $[2C, -CH(CH, O-),]$ ; 86.33  $[-CPhAr_2]$ ; 113.00  $[4C, C]$ *ortho* to OMe]; 119.18 [C<sub>3</sub>]; 122.92 [C<sub>5</sub>]; 126.55 [C<sub>6</sub>]; 126.73 [*C para* of Ph]; 127.37 [*C*4a]; 127.73 and 127.95 [4C, *C ortho* and *meta* of Ph]; 129.05 and 130.35 [2C, *C*<sup>7</sup> and *C*8]; 129.91 [4C, *C meta* to OMe]; 135.67 [2C, *C para* to OMe]; 144.56 [*C ipso* of Ph]; 145.55 [*C*4]; 148.38 [*C*8a]; 149.82 [*C*2]; 158.40 [2C, *C*OMe of DMTr]; 170.84 [*C*O].

## **4.10. (***R***)-2-(Quinolin-4-yl)-3-(trityloxy)propan-1-ol, 6e**

A solution of **6b** (1.90 g, mmol) was dissolved in dry methanol (13 ml) and cooled to 0°C. A solution of 1 M solution of KOH in MeOH (5 ml) was added and the resulting mixture was stirred at the same temperature for about 2 h. The resulting solution was treated with NH<sub>4</sub>Cl sat. sol. (10 ml) and concentrated under reduced pressure. The residue was diluted with water and extracted with AcOEt. Chromatography with  $PE/Et<sub>2</sub>O$  $3:7 \rightarrow Et_2O$  then AcOEt gave pure **6e** (1.72 g, 99%) as a

white foam.  $R_f$  0.26 (PE/Et<sub>2</sub>O 1:9, **A**, **B**).  $[\alpha]_D = +59.7$ (*c* 2.18, CHCl3). IR: max 3518, 3000, 2958, 1729, 1589, 1447, 1065. GC–MS: **6e** decomposed in the column. <sup>1</sup> H NMR: 2.50 [1H, broad s, –O*H*]; 3.62 [2H, d,  $-CH_2O$ ,  $J=6.2$ ]; 3.98 [1H, quintuplet,  $-CH(CH_2O)_{2}$ , *J*=5.9]; 4.02–4.18 [2H, m, –C*H*2O–]; 7.13–7.41 [16H, m,  $H_3$  and aromatics of Tr]; 7.53 [1H, ddd,  $H_6$ ,  $J=$ 1.2, 7.0, 8.4]; 7.68 [1H, ddd, *H*7, *J*=1.4, 7.0, 8.4]; 7.95  $[H, dd, H<sub>5</sub>, J=0.8, 8.4]; 8.08 [1H, dd, H<sub>8</sub>, J=1.2],$ 8.4]; 8.74 [1H, d,  $H_2$ ,  $J=4.6$ ]. <sup>13</sup>C NMR: 42.30 [-CH(CH<sub>2</sub>O-)<sub>2</sub>]; 64.45 and 65.20 [2C, -CH(CH<sub>2</sub>O-)<sub>2</sub>]; 87.26 [–*C*Ph3]; 119.07 [*C*3]; 123.00 [*C*5]; 126.50 [*C*6]; 127.11 [3C, *C para* of Tr]; 127.37 [*C*4a]; 127.85 and 128.50 [12C, *C ortho* and *meta* of Tr]; 129.05 and 130.05 [2C, *C*<sup>7</sup> and *C*8]; 143.56 [3C, *C ipso* of Tr]; 146.40 [C<sub>4</sub>]; 148.07 [C<sub>8a</sub>]; 149.67 [C<sub>2</sub>].

# **4.11. (***S***)-4-{1-[(Triethylsilyl)oxymethyl]-2-(trityloxy) ethyl}quinoline, 6f**

A solution of  $6e$  (523 mg, 1.17 mmol) in dry CH<sub>2</sub>Cl<sub>2</sub> (10 ml) was cooled to 0°C and treated with 2,6 lutidine (232 µl, 2.00 mmol) and triethylsilyl triflate  $(319 \mu l, 1.41 \text{ mmol})$ . After about 2 h at the same temperature, the solution was partitioned between water and  $Et<sub>2</sub>O$ . After the addition of aqueous  $NaHCO<sub>3</sub>$  until pH 8 of the aqueous layer, an usual extraction was performed. After solvent evaporation, 2,6-lutidine was azeotropically removed with *n*-octane. Chromatography with  $PE/Et_2O$  6:4 $\rightarrow$ 4:6 gave 6f as a yellow oil.  $R_f$  0.77 (PE/Et<sub>2</sub>O 1:9, **A, B**).  $[\alpha]_D = +15.6$  (*c* 2.06, CHCl<sub>3</sub>). IR:  $v_{\text{max}}$  2948, 2872, 1590, 1447, 1193, 1094, 1002. GC–MS: **6f** decomposed in the column. <sup>1</sup> <sup>1</sup>H NMR: 0.46 [6H, dq,  $-Si(CH_2CH_3)$ <sub>3</sub>,  $J=1.2, 7.8$ ]; 0.82 [9H, t,  $-Si(CH_2CH_3)_3$ ,  $J=7.4$ ]; 3.48 and 3.65 [2H, AB part of ABX system,  $-CH_2O^-$ ,  $J_{AB} = 8.8$ ,  $J_{AX}$  and  $J_{\text{BX}} = 5.6$ , 5.8]; 3.50 [1H, quintuplet,  $-CH(CH_2O-)_{2}$ , *J*=5.7]; 3.98-4.04 [2H, m, -CH<sub>2</sub>O-]; 7.13-7.34 [16H, m,  $H_3$  and aromatics of Tr]; 7.54 [1H, ddd,  $H_6$ ,  $J=$ 1.0, 7.0, 8.4]; 7.70 [1H, ddd,  $H_7$ ,  $J=1.0$ , 6.6, 8.0]; 8.08  $[H, d, H<sub>5</sub>$  or  $H<sub>8</sub>, J=8.8]$ ; 8.12 [1H, d,  $H<sub>5</sub>$  or  $H<sub>8</sub>$ , *J*=8.4]; 8.78 [1H, d, *H*2, *J*=4.8]. 13C NMR: 4.26 [3C,  $-Si(CH_2CH_3)$ ; 6.66 [3C,  $-Si(CH_2CH_3)$ <sub>3</sub>]; 42.58  $[-CH(CH, O<sub>-</sub>)<sub>2</sub>]$ ; 63.33 and 63.81 [2C,  $-CH(CH, O<sub>-</sub>)<sub>2</sub>$ ]; 86.67 [–*C*Ph3]; 119.46 [*C*3]; 123.33 [*C*5]; 126.21 [*C*6]; 126.89 [3C, *C para* of Tr]; 127.66 and 128.60 [12C, *C ortho* and *meta* of Tr]; 127.82 [*C*4a]; 128.81 and 130.25 [2C, *C*<sup>7</sup> and *C*8]; 143.86 [3C, *C ipso* of Tr]; 147.28 [*C*4]; 148.35 [C<sub>8a</sub>]; 149.76 [C<sub>2</sub>].

# **4.12. (***S***)-4-[1-(***p***-Methoxyphenoxymethyl)-2-(trityloxy) ethyl]quinoline, 6g**

The same procedure employed for the preparation of **5f**,**g** from **5a** was followed, starting from **6e** to give **6g** in 72% yield as a white foam.  $R_f$  0.56 (PE/Et<sub>2</sub>O 1:9, A, **B**).  $[\alpha]_D = -13.0$  (*c* 2.00, CHCl<sub>3</sub>). IR:  $v_{\text{max}}$  2955, 1730, 1590, 1464, 1444, 1250, 1067. GC–MS: **6g** decomposed in the column.  ${}^{1}H$  NMR: 3.60 and 3.69 [2H, AB part of ABX system,  $-CH_2O^-$ ,  $J_{AB} = 9.0$ ,  $J_{AX}$  and  $J_{BX} = 5.8$ , 5.9]; 3.79 [3H, s, –OC*H*3]; 4.20 [1H, center of m, >C*H*CH2O–]; 4.34–4.46 [2H, m, –C*H*2O–]; 6.75–6.86 [4H, m, *p*-anisyl]; 7.17–7.36 [16H, m,  $H_3$  and aromat-

ics of Tr]; 7.53 [1H, ddd,  $H_6$ ,  $J=1.4$ , 7.0, 8.4]; 7.71  $[H, ddd, H<sub>7</sub>, J=1.3, 7.0, 8.4]; 7.99 [1H, d, H<sub>5</sub>, J=$ 8.8]; 8.13 [1H, dd,  $H_8$ ,  $J=1.0$ , 8.4]; 8.81 [1H, d,  $H_2$ ,  $J=4.4$ ]. <sup>13</sup>C NMR: 40.05 [-CH(CH<sub>2</sub>O–)<sub>2</sub>]; 55.63  $[-OCH_3]$ ; 62.07 and 68.44 [2C,  $-CH(CH_2O-)_2]$ ; 86.84 [–*C*Ph3]; 114.57 [2C, *C ortho* to –OMe]; 115.56 [2C, *C meta* to –OMe]; 119.48 [*C*3]; 122.97 [*C*5]; 126.45 [*C*6]; 126.96 [3C, *C para* of Tr]; 127.42 [*C*4a]; 127.71 and 128.52 [12C, *C ortho* and *meta* of Tr]; 128.94 and 130.26 [2C, *C*<sup>7</sup> and *C*8]; 143.65 [3C, *C ipso* of Tr]; 146.18 [C<sub>4</sub>]; 148.28 [C<sub>8a</sub>]; 149.80 [C<sub>2</sub>]; 152.62 and 153.98 [2C, *C* quat. of *p*-anisyl].

## **4.13. Preparation of commercially unavailable alkynes**

**4.13.1. Prop-2-ynyloxymethylbenzene (used for the preparation of 8,9k)**. A suspension of NaH (60% in mineral oil, 672 mg, 1.68 mmol) in dry DMF (10 ml) was cooled to  $0^{\circ}$ C; then benzyl alcohol (1.65 g, 15.28) mmol) was added through an addition funnel during a period of 10 min. The funnel was washed with additional DMF (3 ml) and the mixture was vigorously stirred for 25 min at  $0^{\circ}$ C. Propargyl bromide (80%, 1.87 ml, 1.68 mmol) was added dropwise via the addition funnel during 5 min. The mixture was allowed to react at 0°C for 2.5 h and 30 min at rt. Additional 122 mg of NaH and overnight stirring at rt were necessary in order to complete the reaction. After cooling again at  $0^{\circ}$ C, the reaction was quenched with NH<sub>4</sub>Cl sat. sol. and extracted with  $Et<sub>2</sub>O$  (1:1). Chromatography with  $PE/Et_2O$  95:5 $\rightarrow$ 9:1 gave pure alkyne as a yellow oil (2.03 g, 91%).  $R_f$  0.76 (PE/Et<sub>2</sub>O 8:2, A, B). GC– MS: *R*<sup>t</sup> 2.77; *m*/*z* 146 (M<sup>+</sup> , 2.8), 145 (8.4), 118 (5.7), 117 (17), 116 (47), 115 (24), 107 (29), 106 (5.0), 105 (4.1), 92 (65), 91 (100), 89 (10), 79 (56), 78 (14), 77 (58), 65 (28), 63 (9.7), 52 (6.4), 51 (30), 50 (12), 40 (8.7), 39 (53), 38 (8.7). <sup>1</sup>H NMR: 2.48 [1H, t, -C=CH, *J*=2.4]; 4.18 [2H, d, -CH<sub>2</sub>C=CH, *J*=2.2]; 4.62 [2H, s,  $-CH_2Ph$ ]; 7.26–7.38 [5H, m, aromatics].

**4.13.2. 1-Methoxy-4-prop-2-ynyloxymethylbenzene (used for the preparation of 8,9l)**. The same procedure described above was followed, performing the reaction on 80.2 mmol of 4-methoxybenzyl alcohol. The reaction was completed over 2.5 h at 0°C and 30 min at rt. After cooling again at 0°C, quenching with 50 ml of a 0.65 M aqueous solution of  $K_2CO_3$  was performed. After extraction with  $Et<sub>2</sub>O$ , chromatography with  $PE/Et_2O$  95:1 $\rightarrow$ 9:1 gave pure alkyne as a yellow oil (10.75 g, 76%).  $R_f$  0.62 (PE/Et<sub>2</sub>O 7:3, A, B). GC– MS: *R*<sup>t</sup> 4.75; *m*/*z* 176 (M<sup>+</sup> , 24), 175 (8.4), 146 (9.2), 145 (5.7), 137 (7.1), 136 (50), 135 (61), 131 (5.2), 122 (17), 121 (100), 109 (14), 108 (6.5), 107 (8.2), 94 (12), 92 (9.5), 91 (14), 89 (7.7), 78 (20), 77 (46), 65 (11), 64 (7.7), 63 (9.4), 53 (5.5), 52 (8.3), 51 (16), 50 (7.6), 40 (6.4), 39 (40), 37 (6.6). <sup>1</sup>H NMR: 2.46 [1H, t, -C=CH,  $J=2.4$ ]; 3.80 [3H, s,  $-OCH_3$ ]; 4.14 [2H, d,  $-CH_2C=CH$ , *J*=2.5]; 4.54 [2H, s, –CH<sub>2</sub>Ar]; 6.85–6.89 [2H, m, *H ortho* to OMe]; 7.26–7.32 [2H, m, *H meta* to OMe]. <sup>13</sup>C NMR: 55.27 [–O*C*H<sub>3</sub>]; 56.68 [–*CH*<sub>2</sub>C=CH]; 71.14 [–*C*H2OAr]; 74.44 [–C*C*H]; 79.81 [–*C*CH]; 113.81 [2C, *C ortho* to OMe]; 129.31 [*C para* to OMe]; 129.68 [2C, *C meta* to OMe]; 159.34 [*C*OMe].

# **4.14. General procedure for the addition of acetylides to 5a–h and 6a–g**

**Preparation of magnesium acetylide**: a solution of the desired alkyne (4.44 mmol) in dry THF (6.03 ml, when EtMgBr was used, 5.36 ml when *i*-PrMgCl was used) was cooled to −15°C, before EtMgBr (1.33 ml of a 3 M solution in Et<sub>2</sub>O, 4.0 mmol) or  $i$ -PrMgCl (2 ml of a 2 M solution in  $Et<sub>2</sub>O$  was added. The resulting red– brown solution was stirred at rt for 1 h to give a 0.5 M solution of acetylide.

**Preparation and employment of cerous acetylide**: a suspension of anhydrous  $CeCl<sub>3</sub>$ , [previously dried by known procedure<sup>46</sup> from CeCl<sub>3</sub>·6H<sub>2</sub>O (494.6 mg 1.40) mmol)] in dry THF (1 ml) was cooled to −30°C and treated with an equimolecular amount of lithium trimethylsilyl acetylide [0.5 M solution in THF, obtained by treatment of ethynyl trimethyl silane at  $-30$ °C with a 1 M solution in THF of LiN(SiMe<sub>3</sub>)<sub>2</sub>]. The temperature was then allowed to raise to rt, before quinoline  $5b$  (105 mg, 350  $\mu$ mol) and phenyl chloroformate (176 µl, 1.40 mmol) are added. Work-up followed the same protocol described later in the general procedure.

**Addition reaction (conditions of Table 2)**: a solution of quinoline (the screening was performed on about 100 mg of derivative) in dry THF (5 ml) was cooled to −78°C. The acetylide solution was added (3 molar equiv.), followed by phenyl chloroformate (3 molar equiv.). The resulting solution was allowed to react at the reported temperature until complete (3–15 h). After quenching with NH4Cl sat. sol., the mixture was extracted as usual with  $Et<sub>2</sub>O$ . Chromatography was performed with the appropriate  $PE/Et<sub>2</sub>O$  mixture. With the exception of **8**,**9d**, the mixture of purified diastereoisomers was obtained. They were further separated, with the exceptions reported below, by preparative thin layer chromatography, as reported for every addition mixture. The yields are reported in Tables 1 and 2. For preparative purposes the same procedure was followed, reducing THF quantities (about 2 ml/ mmol of substrate) and acetylide or chloroformate, as reported in the Tables 1 and 2.

**4.14.1. Determination of d.r. by chromatographic methods (compounds 8,9a–j)**. **8**,**9a**: GC–MS (usual conditions but with final temperature 290 $^{\circ}$ C);  $R_1$  12.25 (8a) and 12.60 (**9a**). **8**,**9b**: GLC [SE-54 column, inj. temp. 300°C, det. temp. 300°C, oven temp. 250°C, carrier (He) press. 1.60 kg/cm2 ]; *R*<sup>t</sup> 3.42 (**8b**) and 3.87 (**9b**). **8**,**9c**: GLC [RSL-150 column, inj. temp. 300°C, det. temp. 250°C, oven temp. 250°C, carrier (He) press. 1.60 kg/cm<sup>2</sup>];  $R_t$ 8.90 (**8c**) and 10.70 (**9c**) or GC–MS (usual conditions but with final temperature 290 $^{\circ}$ C)  $R_1$  11.89 (8c) and 12.21 (**9c**). **8,9d**: HPLC (hexane/Et<sub>2</sub>O 4:6, flow 1.5 ml/min); *R*<sup>t</sup> 21.55 (**8d**) and 11.90 (**9d**). **8**,**9e**: HPLC (hexane/CH<sub>2</sub>Cl<sub>2</sub>/Et<sub>2</sub>O 95:3:2, flow 1.5 ml/min);  $R_1$ , 13.20 (**8e**) and 16.40 (**9e**). **8**,**9f**: GLC [SE-54 column, inj. temp. 300°C, det. temp. 300°C, oven temp. 250°C, carrier (He) press.  $1.60 \text{ kg/cm}^2$ ];  $R_t$  5.77 (8f) and 6.06 (9f). **8.9g**: HPLC (hexane/Et<sub>2</sub>O 8:2, flow 1.2 ml/min);  $R_1$ 

5.54 (**8g**) and 4.50 (**9g**). **8**,**9h**: GLC [RSL-150 column, inj. temp. 300°C, det. temp. 250°C, oven temp. 280°C, carrier (He) press. 1.60 kg/cm2 ]; *R*<sup>t</sup> 10.43 (**8h**) and 11.51 (9h). **8.9i**: HPLC (hexane/Et<sub>2</sub>O 8:2, flow 1.2 ml/min);  $R_t$ 7.30 (8i) and 10.17 (9i). 8,9j: HPLC (hexane/Et<sub>2</sub>O 95:5, flow 1.5 ml/min);  $R_1$  12.02 (8j) and 14.28 (9j).

**4.14.2. 4-{(***S***\*)-[2-(***tert***-Butyldimethylsilyloxy)-1-methyl] ethyl}-2-{(***R***\*)-[(trimethylsilyl)ethynyl]}-2***H***-quinoline-1 carboxylic acid benzyl ester 8a and its epimer, 9a**. Preparative thin layer chromatography was performed with  $PE/Et_2O$  (9:1) to give **8a** and **9a** as pale yellow oils.  $R_f$  0.55 (8a) and 0.44 (9a) (PE/Et<sub>2</sub>O 9:1, **A**, **B**). **Charac**terization of 8a: IR:  $v_{\text{max}}$  2929, 2858, 2168, 2091, 1696, 1391, 1288, 1247. GC–MS: (usual conditions, but with fin. temp. 290°C) *R*<sub>t</sub> 12.25; *m*/*z* 398 (M<sup>+</sup>−135, 9.6), 316 (6.4), 266 (14), 245 (55), 244 (26), 238 (5.6), 92 (7.1), 91 (100), 75 (7.3), 73 (47). <sup>1</sup> H NMR: −0.09 and −0.07 [6H, 2 s,  $>Si(CH_3)_2$ ]; 0.02 [9H, s,  $-Si(CH_3)_3$ ]; 0.82 [9H, s,  $-C(CH)$ <sub>3</sub>]; 1.26 [3H, d,  $>CHCH_3$ ,  $J=6.6$ ]; 3.04 [1H, center of m,  $>CHCH<sub>3</sub>$ ]; 3.30 and 3.64 [2H, AB part of ABX system, >CHC $H_2O$ –,  $J_{AB}$ =9.6,  $J_{AX}$  and  $J_{BX}$ =4.1, 7.8]; 5.22 and 5.30 [2H, AB system,  $-CH_2Ph$ ,  $J_{AB} =$ 12.4]; 5.83 and 5.83 [2H, AB system,  $H_2$  and  $H_3$ ,  $J_{AB}$ =6.9]; 7.14 [1H, dt,  $H_6$  or  $H_7$ ,  $J$ =1.5, 7.4]; 7.24 [1H, dt,  $H_6$  or  $H_7$ ,  $J=1.5$ , 7.8]; 7.31–7.44 [6H, m,  $H_5$  and Ph]; 7.64 [1H, broad d,  $H_8$ , *J*=8.2]. <sup>13</sup>C NMR: −5.45 and −5.37 [2C, >Si(*CH*<sub>3</sub>)<sub>2</sub>]; −0.13 [3C, -Si(*CH*<sub>3</sub>)<sub>3</sub>]; 15.88  $[\neg \text{CHCH}_3]$ ; 18.28  $[-C(\text{CH}_3)_3]$ ; 25.90  $[3C, -C(\text{CH}_3)_3]$ ; 36.11 [>*C*HCH3]; 44.81 [*C*2]; 67.32 and 67.89 [2C, –*C*H2O–]; 88.08 and 101.99 [2C, –*CC*–TMS]; 121.22 [ $C_8$ ]; 122.99, 124.36, 124.85 and 127.25 [4C,  $C_3$ ,  $C_5$ ,  $C_6$ , *C*7]; 127.94 and 128.49 [4C, *C ortho* and *meta* of Ph]; 128.10 [*C para* of Ph]; 127.78 [*C*4a]; 134.52 [*C*4]; 136.05 [*C ipso* of Ph]; 137.09 [*C*8a], 153.20 [*C*O]. **Characterization of 9a**: IR:  $v_{\text{max}}$  2956, 2928, 2169, 1696, 1391, 1247, 1095, 1025. GC–MS: (usual conditions, but with fin. temp. 290°C) *R*<sub>t</sub> 12.60; *m*/*z* 398 (M<sup>+</sup>−135, 12), 316 (5.0), 266 (13), 244 (10), 238 (6.1), 155 (8.2), 92 (8.7), 91 (100), 89 (7.3), 75 (6.6), 73 (46). <sup>1</sup> H NMR: 0.02 [9H, s,  $-Si(CH_3)$ ; 0.04 and 0.06 [6H, 2 s,  $>Si(CH_3)$ ]; 0.90 [9H, s, –C(C*H*)3]; 1.09 [3H, d, >CHC*H*3, *J*=6.9]; 2.98 [1H, sextuplet,  $>CHCH_3$ ,  $J=6.7$ ; 3.63 and 3.81 [2H, AB part of ABX system, >CHC $H_2O$ –,  $J_{AB}$ =9.8,  $J_{AX}$  and  $J_{\rm BX}$ =5.9, 6.8]; 5.24 and 5.30 [2H, AB system, –CH<sub>2</sub>Ph,  $J_{AB}$ =12.5]; 5.83 and 5.83 [2H, AB system,  $H_2$  and  $H_3$ ,  $J_{AB}$ =7.2]; 7.14 [1H, dt,  $H_6$  or  $H_7$ , *J*=1.4, 7.5]; 7.24 [1H, dt,  $H_6$  or  $H_7$ ,  $J=1.5$ , 7.7]; 7.32–7.44 [6H, m,  $H_5$  and Ph]; 7.64 [1H, broad d, *H*<sub>8</sub>, *J*=7.5]. <sup>13</sup>C NMR: −5.24<br>[2C, >Si(CH<sub>3</sub>)<sub>2</sub>]; −0.12 [3C, −Si(CH<sub>3</sub>)<sub>3</sub>]; 17.47  $[2\overline{C}, >\overline{Si(CH_3)_2}]; -0.12^\circ [3C,$ [>CH*C*H3]; 18.35 [–*C*(CH3)3]; 26.00 [3C, –C(*C*H3)3]; 36.11 [>*C*HCH3]; 44.62 [*C*2]; 67.43 and 67.89 [2C, –*C*H2O–]; 87.92 and 102.02 [2C, –*CC*–TMS]; 120.22 [C<sub>8</sub>]; 123.17, 124.34, 124.66 and 127.16 [4C, C<sub>3</sub>, C<sub>5</sub>, C<sub>6</sub>, *C*7]; 127.86 and 128.48 [4C, *C ortho* and *meta* of Ph]; 128.08 [*C para* of Ph]; 128.55 [*C*4a]; 134.41 [*C*4]; 136.05 [*C ipso* of Ph]; 137.95 [*C*8a], 153.29 [*C*O].

**4.14.3. 4-{(***S***\*)-[2-(***tert***-Butyldimethylsilyloxy)-1-methyl] ethyl}-2-{(***R***\*)-[(trimethylsilyl)ethynyl]}-2***H***-quinoline-1 carboxylic acid methyl ester 8b and its epimer, 9b**. Preparative thin layer chromatography was performed with  $PE/Et$ ,  $O(9:2)$  to give **8b** and **9b** as pale yellow oils.  $R_f$  0.62 (8b) and 0.53 (9b) (PE/Et<sub>2</sub>O 8:2, A, B). **Charac**terization of 8b: IR:  $v_{\text{max}}$  2954, 2927, 2169, 2089, 1702, 1381, 1332, 1096, 1035. GC–MS: (usual conditions, but with fin. temp. 290°C)  $R_1$  10.21;  $m/z$  457 (M<sup>+</sup>, 0.39), 400 (7.1), 284 (17), 180 (6.3), 155 (11), 89 (35), 75 (17), 74 (8.8), 73 (100), 59 (22), 45 (6.6). <sup>1</sup> H NMR: −0.07 and  $-0.05$  [6H, 2 s, >Si(CH<sub>3</sub>)<sub>2</sub>]; 0.01 [9H, s,  $-Si(CH_3)_{3}$ ]; 0.84 [9H, s,  $-C(CH)$ <sub>3</sub>]; 1.26 [3H, d,  $>CHCH_3$ ,  $J=6.6$ ]; 3.04 [1H, center of m, >C*H*CH3]; 3.28 and 3.64 [2H, AB part of ABX system, >CHC $H_2O$ –,  $J_{AB}$ =9.7,  $J_{AX}$  and  $J_{\text{BX}} = 4.1, 7.9$ ]; 3.80 [3H, s,  $-OCH_3$ ]; 5.80 and 5.86 [2H, AB system,  $H_2$  and  $H_3$ ,  $J_{AB} = 6.8$ ]; 7.15 [1H, dt,  $H_6$  or *H*<sub>7</sub>, *J*=1.4, 7.5]; 7.26 [1H, dt, *H*<sub>6</sub> or *H*<sub>7</sub>, *J*=1.6, 7.6]; 7.40 [1H, dd,  $H_5$ ,  $J=1.6$ , 7.6]; 7.58 [1H, broad d,  $H_8$ , *J*=7.0]. <sup>13</sup>C NMR: −5.56 and −5.44 [2C, >Si(*CH*<sub>3</sub>)<sub>2</sub>];<br>-0.20 [3C, -Si(*CH*<sub>3</sub>)<sub>3</sub>]; 15.81 [>CH*CH*<sub>3</sub>]; 18.22 −0.20 [3C, –Si(*C*H3)3]; 15.81 [>CH*C*H3]; 18.22 [–*C*(CH3)3]; 25.84 [3C, –C(*C*H3)3]; 36.08 [>*C*HCH3]; 44.64 [*C*2]; 53.18 [–O*C*H3]; 67.28 [–*C*H2O–]; 87.99 and 101.93 [2C, –*CC*–TMS]; 121.45 [*C*8]; 123.00, 124.43, 124.96 and 127.25 [4C,  $C_3$ ,  $C_5$ ,  $C_6$ ,  $C_7$ ]; 127.87 [ $C_{4a}$ ]; 134.56 [*C*4]; 137.08 [*C*8a], 153.90 [*C*O]. **Characterization** of 9b: IR:  $v_{\text{max}}$  2927, 2855, 1697, 1440, 1380, 1094, 1036. GC–MS: (usual conditions, but with fin. temp. 290°C) *R*<sup>t</sup> 10.01; *m*/*z* 457 (M<sup>+</sup> , 2.9), 443 (11), 442 (30), 402 (12), 401 (32), 400 (100), 370 (5.9), 339 (6.3), 338 (21), 310 (8.4), 285 (14), 284 (60), 180 (6.0), 278 (6.5), 268 (5.3), 266 (18), 252 (7.5), 194 (5.5), 192 (6.2), 180 (8.7), 89 (24), 75 (9.5), 73 (50), 59 (9.7). <sup>1</sup> H NMR: −0.005 [9H, s,  $-Si(CH_3)$ ; 0.04 and 0.07 [6H, 2 s,  $>Si(CH_3)$ ; 0.89 [9H, s,  $-C(CH)$ <sub>3</sub>]; 1.09 [3H, d,  $>CHCH$ <sub>3</sub>,  $J=6.9$ ]; 2.97 [1H, sextuplet,  $>CHCH_3$ ,  $J=6.6$ ; 3.62 and 3.81 [2H, AB part of ABX system, >CHC $H_2O$ –,  $J_{AB}=9.8$ ,  $J_{AX}$  and *J*<sub>BX</sub> = 6.0, 6.8]; 3.81 [3H, s, -OC*H*<sub>3</sub>]; 5.81 and 5.83 [2H, AB system,  $H_2$  and  $H_3$ ,  $J_{AB} = 7.0$ ]; 7.14 [1H, dt,  $H_6$  or *H*<sub>7</sub>,  $J=1.4$ , 7.4]; 7.25 [1H, dt,  $H_6$  or  $H_7$ ,  $J=1.7$ , 8.1]; 7.42 [1H, dd,  $H_5$ ,  $J=1.8$ , 7.8]; 7.58 [1H, broad d,  $H_8$ , *J*=8.2]. <sup>13</sup>C NMR: −5.34 and −5.29 [2C, >Si( $CH_3$ )<sub>2</sub>]; −0.19 [3C, –Si(*C*H3)3]; 17.44 [>CH*C*H3]; 18.30 [–*C*(CH3)3]; 25.96 [3C, –C(*C*H3)3]; 36.12 [>*C*HCH3]; 44.46 [C<sub>2</sub>]; 53.25 [–OCH<sub>3</sub>]; 67.50 [–CH<sub>2</sub>O–]; 87.88 and 102.00 [2C, –*CC*–TMS]; 120.45 [*C*8]; 123.22, 124.43, 124.78 and 127.18 [4C, *C*3, *C*5, *C*6, *C*7]; 128.21 [*C*4a]; 134.48 [*C*4]; 138.05 [*C*8a], 154.02 [*C*O].

**4.14.4. 4-{(***S***\*)-[2-(***tert***-Butyldimethylsilyloxy)-1-methyl] ethyl}-2-{(***R***\*)-[(trimethylsilyl)ethynyl]}-2***H***-quinoline-1 carboxylic acid phenyl ester, 8c and its epimer, 9c**. Preparative thin layer chromatography was performed with  $PE/Et_2O$  (9:1) to give **8c** and **9c** as pale yellow oils.  $R_f$  0.77 (8c) and 0.72 (9c) (PE/Et<sub>2</sub>O 8:2, A, B). **Charac**terization of 8c: IR:  $v_{\text{max}}$  2955, 2171, 1715, 1382, 1328, 1289, 1243. GC–MS: (usual conditions, but with fin. temp. 290°C) *R*<sub>t</sub> 12.06; *m*/*z* 398 (M<sup>+</sup>−121, 2.8), 347 (8.7), 346 (28), 338 (8.3), 278 (6.0), 266 (12), 252 (5.5), 222 (5.3), 180 (8.6), 151 (25), 115 (6.0), 89 (30), 77 (12), 75 (15), 74 (9.0), 73 (100), 59 (7.6). <sup>1</sup> H NMR: −0.038 and  $-0.028$  [6H, 2 s, >Si(CH<sub>3</sub>)<sub>2</sub>]; 0.04 [9H, s,  $-Si(CH_3)$ <sub>3</sub>]; 0.86 [9H, s,  $-C(CH)$ <sub>3</sub>]; 1.30 [3H, d,  $>CHCH_3$ ,  $J=6.7$ ]; 3.09 [1H, center of m, >C*H*CH3]; 3.37 and 3.69 [2H, AB part of ABX system, >CHC $H_2O$ -,  $J_{AB}$ =9.7,  $J_{AX}$ and  $J_{\text{BX}} = 4.2$ , 7.5]; 5.91 [2H, apparent s,  $H_2$  and  $H_3$ ]; 7.14–7.46 [8H, m, H<sub>5</sub>, H<sub>6</sub>, H<sub>7</sub>, Ph]; 7.73 [1H, broad d, *H*<sub>8</sub>, *J*=8.0]. <sup>13</sup>C NMR: −5.38 [2C, >Si(*C*H<sub>3</sub>)<sub>2</sub>]; −0.13  $[3C, -Si(CH_3)_3]$ ; 15.90  $[>CHCH_3]$ ; 18.31  $[-C(CH_3)_3]$ ; 25.93 [3C,  $-C(CH_3)_3$ ]; 36.14 [>*C*HCH<sub>3</sub>]; 45.17 [*C*<sub>2</sub>]; 67.31 [– $CH_2O$ –]; 88.49 and 101.54 [2C, – $C \equiv C$ –TMS]; 121.61 [2C, *C ortho* of Ph]; 121.33 [*C*8]; 123.14, 124.79, 125.62 and 127.40 [4C, *C*<sub>3</sub>, *C*<sub>5</sub>, *C*<sub>6</sub>, *C*<sub>7</sub>]; 125.66 [*C para* of Ph]; 128.05 [*C*4a]; 129.30 [2C, *C meta* of Ph]; 134.24 [*C*4]; 137.28 [*C*8a]; 151.00 [*C ipso* of Ph]; 151.77 [*C*O]. **Characterization of 9c**: IR:  $v_{\text{max}}$  2954, 2927, 2169, 1714, 1381, 1327, 1304, 1246, 1190. GC–MS: (usual conditions, but with fin. temp. 290°C)  $R_1$  12.40;  $m/z$  398 (M<sup>+</sup> −121, 2.4), 347 (6.5), 346 (22), 338 (6.1), 266 (8.1), 180 (7.5), 156 (5.4), 155 (13), 151 (20), 115 (5.3), 89 (34), 77 (14), 75 (14), 74 (7.5), 73 (100), 59 (8.0). <sup>1</sup> H NMR: 0.04 [9H, s,  $-Si(CH_3)$ <sub>3</sub>]; 0.08 and 0.06 [6H, 2 s,  $>Si(CH_3)$ ; 0.91 [9H, s, –C(CH)<sub>3</sub>]; 1.14 [3H, d, >CHC*H*3, *J*=6.9]; 3.02 [1H, sextuplet, >C*H*CH3, *J*= 6.6]; 3.67 and 3.85 [2H, AB part of ABX system,  $>CHCH<sub>2</sub>O-$ ,  $J<sub>AB</sub>=9.5$ ,  $J<sub>AX</sub>$  and  $J<sub>BX</sub>=5.7$ , 6.7]; 5.92 [2H, apparent s,  $H_2$  and  $H_3$ ]; 7.14–7.49 [8H, m,  $H_5$ ,  $H_6$ , *H*<sub>7</sub>, Ph]; 7.72 [1H, broad d, *H*<sub>8</sub>, *J*=7.0]. <sup>13</sup>C NMR:<br>-5.25 and -5.21 [2C, >Si(CH<sub>3</sub>)<sub>2</sub>]; -0.13 [3C, −5.25 and −5.21 [2C, >Si(*C*H3)2]; −0.13 [3C, –Si(*C*H3)3]; 17.52 [>CH*C*H3]; 18.36 [–*C*(CH3)3]; 26.02 [3C, –C(*C*H3)3]; 36.22 [>*C*HCH3]; 44.97 [*C*2]; 67.49 [–*C*H2O–]; 88.33 and 101.57 [2C, –*CC*–TMS]; 120.43 [*C*8]; 121.64 [2C, *C ortho* of Ph]; 123.33, 124.80, 127.32 and 129.54 [4C, *C*<sub>3</sub>, *C*<sub>5</sub>, *C*<sub>6</sub>, *C*<sub>7</sub>]; 125.64 [*C* para of Ph]; 128.36 [*C*4a]; 129.31 [2C, *C meta* of Ph]; 134.16 [*C*4]; 138.16 [*C*8a]; 151.00 [*C ipso* of Ph]; 151.90 [*C*O].

**4.14.5. 4-{(***S***\*)-[2-Hydroxy-1-methyl]ethyl}-2-{(***R***\*)- [(trimethylsilyl)ethynyl]}-2***H***-quinoline-1-carboxylic acid phenyl ester, 8d and its epimer, 9d**. Pure **8d** and **9d** have been isolated by chromatography with  $PE/Et_2O$  (4:6) as solids. Both have been crystallized from  $Et_2O/PE$  to give white solids.  $R_f$  0.39 (8d) and 0.47 (9d) (PE/Et<sub>2</sub>O 6:4, **A**, **B**). **Characterization of 8d**: Mp 109.3–110.7°C (Et<sub>2</sub>O/PE). IR:  $v_{\text{max}}$  3621, 2961, 2929, 2169, 1715, 1383, 1191, 1024. GC–MS:  $R_1$  11.62;  $m/z$  406 (9.5), 405 (M<sup>+</sup>, 28), 404 (10), 374 (11), 360 (8.2), 348 (9.7), 347 (30), 346 (100), 329 (12), 328 (53), 313 (5.7), 312 (26), 284 (12), 267 (5.5), 254 (14), 253 (6.5), 252 (15), 238 (7.9), 226 (12), 222 (5.2), 208 (9.9), 194 (11), 180 (19), 156 (5.6), 77 (22), 75 (12), 74 (5.5), 73 (57). <sup>1</sup> H NMR: 0.04 [9H, s, –Si(C*H*3)3]; 1.34 [3H, d, >CHC*H*3, *J*=6.8]; 3.16 [1H, sextuplet,  $>CHCH_3$ ,  $J=6.6$ ; 3.64 [2H, m, became an AB part of ABX system at 3.54 and 3.72 after exchange with  $D_2O$ , >CHC*H*<sub>2</sub>O–,  $J_{AB} = 10.7$ ,  $J_{AX}$  and  $J_{BX} = 4.8$ , 6.2]; 5.94 and 5.98 [2H, AB system,  $H_2$  and  $H_3$ ,  $J_{AB}$ = 7.0]; 7.16–7.46 [8H, m,  $H_5$ ,  $H_6$ ,  $H_7$ , Ph]; 7.76 [1H, broad d, *H*8, *J*=8.2]. 13C NMR: −0.22 [3C, –Si(*C*H3)3]; 15.81 [>CH*C*H3]; 36.02 [>*C*HCH3]; 45.15 [*C*2]; 66.40 [–*C*H2O–]; 88.72 and 101.26 [2C, –*CC*–TMS]; 121.52 [2C, *C ortho* of Ph]; 121.53 [*C*8]; 123.05, 124.84 [most likely 2C] and 127.66 [4C,  $C_3$ ,  $C_5$ ,  $C_6$ ,  $C_7$ ]; 125.63 [C] *para* of Ph]; 127.46 [*C*4a]; 129.27 [2C, *C meta* of Ph]; 134.32 [*C*4]; 136.69 [*C*8a]; 150.88 [*C ipso* of Ph]; 151.76 [CO]. **Characterization of 9d**: Mp  $112.3-113.0^{\circ}$ C (Et<sub>2</sub>O) PE). IR:  $v_{\text{max}}$  3611, 2963, 2929, 2393, 1715, 1484, 1300, 1207, 1024. GC-MS:  $R_1$  11.66;  $m/z$  406 (7.5), 405 (M<sup>+</sup>, 24), 404 (8.4), 374 (9.8), 348 (7.5), 347 (28), 346 (100), 330 (5.0), 329 (12), 328 (49), 313 (5.9), 312 (25), 284 (12), 282 (10), 268 (5.1), 266 (19), 254 (14), 253 (7.7),

252 (18), 238 (9.3), 236 (5.6), 226 (12), 208 (8.9), 194 (10), 180 (17), 151 (6.4), 77 (20), 75 (9.3), 73 (53). <sup>1</sup> H NMR: 0.04 [9H, s, –Si(CH<sub>3</sub>)<sub>3</sub>]; 1.19 [3H, d, >CHCH<sub>3</sub>, *J*=7.0]; 4.13 [1H, sextuplet, >C*H*CH3, *J*=6.5]; 3.77 and 3.89 [2H, AB part of ABX system,  $\geq$ CHC*H*<sub>2</sub>O–,  $J_{AB}$ =10.7,  $J_{AX}$  and  $J_{BX}$ =6.0, 6.3]; 5.95 and 5.98 [2H, AB system,  $H_2$  and  $H_3$ ,  $J_{AB} = 6.8$ ]; 7.17–7.48 [8H, m,  $H_5$ ,  $H_6$ ,  $H_7$ , Ph]; 7.75 [1H, broad d,  $H_8$ ,  $J = 7.7$ ]. <sup>13</sup>C NMR: −0.23 [3C, –Si(*C*H3)3]; 17.14 [>CH*C*H3]; 36.03 [>*C*HCH3]; 44.81 [*C*2]; 66.61 [–*C*H2O–]; 88.76 and 101.60 [2C, –*CC*–TMS]; 120.64 [*C*8]; 121.49 [2C, *C ortho* of Ph]; 122.96, 123.07 and 124.87 [most likely 2C] [4C, *C*3, *C*5, *C*6, *C*7]; 125.74 [*C para* of Ph]; 127.62 [*C*4a]; 129.25 [2C, *C meta* of Ph]; 134.23 [*C*<sub>4</sub>]; 137.44 [*C*<sub>8a</sub>]; 150.86 [*C ipso* of Ph]; 151.77 [*C*O].

**4.14.6. 4-{(***S***\*)-[2-(***tert***-Butyldiphenylsilyloxy)-1-methyl] ethyl}-2-{(***R***\*)-[(trimethylsilyl)ethynyl]}-2***H***-quinoline-1 carboxylic acid phenyl ester, 8e and its epimer, 9e**. Preparative thin layer chromatography was performed with  $PE/Et_2O$  (95:5) to give **8e** and **9e** as pale yellow oils.  $R_f$  0.48 (**8e**) and 0.44 (**9e**) (PE/CH<sub>2</sub>Cl<sub>2</sub>/Et<sub>2</sub>O 85:7:8, **A, B). Characterization of 8e**: IR:  $v_{\text{max}}$  2927, 2895, 2857, 2170, 1711, 1378, 1327, 1302, 1191, 1110. GC–MS: unsuitable for this analysis.  $^1H$  NMR: 0.04 [9H, s,  $-Si(CH_3)_3$ ; 1.00 [9H, s,  $-C(CH)_3$ ]; 1.38 [3H, d,  $>ECHCH_3$ ,  $J=7.1$ ; 3.12 [1H, center of m,  $>EHCH_3$ ]; 3.51 and 3.71 [2H, AB part of ABX system,  $>CHCH_2O_7$ ,  $J_{AB}=9.7$ ,  $J_{AX}$  and  $J_{BX}=4.2$ , 7.0]; 5.91 and 5.91 [2H, AB system,  $H_2$  and  $H_3$ ,  $J = 7.5$ ]; 7.04 [1H, dt,  $H_6$ ,  $J=1.2$ , 8.1]; 7.07–7.75 [18H, m, aromatics]. <sup>13</sup>C NMR: −0.18 [3C, –Si(*C*H3)3]; 15.80 [>CH*C*H3]; 19.26 [–*C*(CH3)3]; 26.78 [3C, –C(*C*H3)3]; 35.93 [>*C*HCH3]; 45.15 [*C*2]; 67.54 [–*C*H2O–]; 88.43 and 101.62 [2C, –*CC*–TMS]; 120.76 [*C*8]; 121.85 [2C, *C ortho* of –OPh]; 123.12, 124.81 [most likely 2C], and 127.33 [4C, *C*<sub>3</sub>, *C*<sub>5</sub>, *C*6, *C*7]; 125.59 [*C para* of –OPh]; 127.60 and 127.70 [5C, *C meta* to Si and, most likely, *C*4a]; 129.28 [2C, *C meta* of –OPh]; 129.53 and 129.62 [2C, *C para* to Si]; 133.82 [*C*4]; 135.53 and 135.61 [4C, *C ortho* to Si]; 135.70 and 135.80 [2C, *C* quat. of PhSi]; 137.00 [C<sub>8a</sub>]; 151.02 [*C ipso* of –OPh]; 151.79 [*C*O]. **Characterization of 9e**: IR: max 2953, 2926, 2854, 2170, 1724, 1378, 1326, 1304, 1233, 1107. GC–MS: unsuitable for this analysis. <sup>1</sup> <sup>1</sup>H NMR: 0.02 [9H, s,  $-Si(CH_3)_3$ ]; 1.08 [9H, s,  $-C(CH)$ <sub>3</sub>]; 1.19 [3H, d,  $>CHCH_3$ ,  $J=7.0$ ]; 3.05 [1H, sextuplet,  $>CHCH_3$ ,  $J=6.7$ ; 3.73 and 3.90 [2H, AB part of ABX system, >CHC $H_2O$ –,  $J_{AB}$ =10.0,  $J_{AX}$  and  $J_{\text{BX}}$ =6.0, 7.1]; 5.93 [2H, s,  $H_2$  and  $H_3$ ]; 7.10 [1H, dt,  $H_6$ , *J*=1.2, 6.9]; 7.17-7.77 [18H, m, aromatics]. <sup>13</sup>C NMR: −0.21 [3C, –Si(*C*H3)3]; 17.43 [>CH*C*H3]; 19.25 [–*C*(CH3)3]; 26.93 [3C, –C(*C*H3)3]; 36.28 [>*C*HCH3]; 44.95 [*C*2]; 67.84 [>*C*H2O–]; 88.37 and 101.47 [2C, –*CC*–TMS]; 120.77 [*C*8]; 121.66 [2C, *C ortho* of –OPh]; 123.22, 124.85 [most likely 2C], and 127.32 [4C, *C*<sub>3</sub>, *C*<sub>5</sub>, *C*6, *C*7]; 125.66 [*C para* of –OPh]; 127.71 and 127.76 [4C, *C meta* to Si]; 128.32 [*C*4a]; 129.35 [2C, *C meta* of –OPh]; 129.62 and 129.72 [2C, *C para* to Si]; 133.67 [*C*4]; 135.52 and 135.60 [2C, *C* quat. of PhSi]; 135.70 and 135.80 [4C, *C ortho* to Si]; 137.79 [*C*8a]; 151.04 [*C ipso* of –OPh]; 151.91 [*C*O].

**4.14.7. 4-{(***S***\*)-[2-Methoxy-1-methyl]ethyl}-2-{(***R***\*)- [(trimethylsilyl)ethynyl]}-2***H***-quinoline-1-carboxylic acid phenyl ester, 8f and its epimer, 9f**. Preparative thin layer chromatography was performed with  $PE/Et_2O$  (7:3) to give **8f** and **9f** as pale yellow oils.  $R_f$  0.68 (**8f**) and 0.72 (9f) (PE/Et<sub>2</sub>O 6:4, A, B). Characterization of 8f: IR:  $v_{\text{max}}$ 2965, 2168, 1713, 1383, 1329, 1304, 1099. GC–MS: *R*<sup>t</sup> 10.86; *m*/*z* 419 (M<sup>+</sup> , 6.8), 347 (7.8), 346 (28), 342 (16), 326 (6.7), 298 (5.1), 254 (5.4), 252 (5.0), 226 (5.1), 222 (5.5), 194 (11), 180 (14), 77 (28), 75 (5.0), 74 (5.4), 73 (62). 59 (7.8), 51 (5.5), 45 (100). <sup>1</sup> H NMR: 0.04 [9H, s,  $-Si(CH_3)$ <sub>3</sub>]; 1.34 [3H, d, >CHCH<sub>3</sub>,  $J=6.2$ ]; 3.11–3.23 (2H) and 3.45–3.56 (1H) [3H, 2 m, >CHCH<sub>2</sub>O–]; 3.32 [3H, s,  $-OCH_3$ ]; 5.92 and 5.96 [2H, AB system,  $H_2$  and *H*<sub>3</sub>, *J*<sub>AB</sub> = 6.8]; 7.17–7.47 [8H, m, *H*<sub>5</sub>, *H*<sub>6</sub>, *H*<sub>7</sub>, Ph]; 7.75 [1H, broad d,  $H_8$ ,  $J = 7.2$ ]. <sup>13</sup>C NMR: −0.25 [3C, –Si(*C*H3)3]; 16.33 [>CH*C*H3]; 33.87 [>*C*HCH3]; 45.10 [C<sub>2</sub>]; 58.84 [–OCH<sub>3</sub>]; 77.00 [–CH<sub>2</sub>O–]; 88.57 and 101.36 [2C, –*CC*–TMS]; 121.29 [*C*8]; 121.59 [2C, *C ortho* of Ph]; 122.95, 124.89 [most likely 2C] and 127.54 [4C, *C*3, *C*5, *C*6, *C*7]; 125.65 [*C para* of Ph]; 127.66 [*C*4a]; 129.31 [2C, *C meta* of Ph]; 134.30 [*C*4]; 136.96 [*C*8a]; 150.95 [*C ipso* of Ph]; 151.82 [*C*O]. **Characterization of 9f**: IR: max 2966, 2394, 1715, 1197, 1081, 1033. GC–MS: *R*<sup>t</sup> 10.99;  $m/z$  419 (M<sup>+</sup>, 6.0), 347 (6.3), 346 (22), 342 (15), 326 (6.3), 254 (5.2), 226 (5.4), 222 (5.7), 194 (11), 180 (13), 77 (28), 75 (5.1), 74 (5.6), 73 (64). 59 (7.8), 51 (5.4), 45 (100). <sup>1</sup> H NMR: 0.04 [9H, s, –Si(C*H*3)3]; 1.19 [3H, d,  $>CHCH_3$ ,  $J=6.9$ ]; 3.11 [1H, center of m, >C*H*CH3]; 3.42 [3H, s, –OC*H*3]; 3.46 and 3.66 [2H, AB part of ABX system, >CHC $H_2O$ –,  $J_{AB}$ =9.2,  $J_{AX}$  and  $J_{\text{BX}}$ =5.0, 7.4]; 5.91 [2H, apparent s,  $H_2$  and  $H_3$ ]; 7.16– 7.46 [8H, m,  $H_5$ ,  $H_6$ ,  $H_7$ , Ph]; 7.73 [1H, broad d,  $H_8$ , *J*=6.8]. <sup>13</sup>C NMR: −0.24 [3C, −Si(*C*H<sub>3</sub>)<sub>3</sub>]; 17.69<br>[>CH*C*H<sub>3</sub>]; 33.88 [>*C*HCH<sub>3</sub>]; 44.88 [*C*<sub>2</sub>]; 59.05 [ $>ECHCH<sub>3</sub>$ ]; 33.88 [ $ECHCH<sub>3</sub>$ ]; 44.88 [-OCH<sub>3</sub>]; 77.02 [-CH<sub>2</sub>O-]; 88.55 and 101.52 [2C,  $-C \equiv C$ -TMS]; 120.44 [ $C_8$ ]; 121.63 [2C, *C ortho* of Ph]; 123.10, 124.89 [most likely 2C] and 127.49 [4C, *C*<sub>3</sub>, *C*<sub>5</sub>, *C*6, *C*7]; 125.67 [*C para* of Ph]; 128.82 [*C*4a]; 129.34 [2C, *C meta* of Ph]; 134.30 [*C*4]; 137.86 [*C*8a]; 151.00 [*C ipso* of Ph]; 151.89 [*C*O].

**4.14.8. 4-{(***S***\*)-[2-Acetoxy-1-methyl]ethyl}-2-{(***R***\*)- [(trimethylsilyl)ethynyl]}-2***H***-quinoline-1-carboxylic acid phenyl ester, 8g and its epimer, 9g**. Preparative thin layer chromatography was performed with  $PE/Et<sub>2</sub>O$  $(8:2)$  to give **8g** and **9g** as pale yellow oils.  $R_f$  0.29 (8g) and 0.33 (9g) (PE/Et<sub>2</sub>O 8:2, A, B). Characterization of **8g**: IR: max 2957, 2173, 1725, 1377, 1196. GC–MS: *R*<sup>t</sup> 11.99; *m*/*z* 447 (M<sup>+</sup> , 8.1), 371 (7.2), 370 (28), 348 (6.7), 347 (28), 346 (100), 294 (17), 280 (5.1), 279 (9.5), 278 (17), 266 (19), 252 (9.7), 250 (9.7), 226 (10), 220 (5.9), 208 (7.0), 194 (17), 180 (20), 151 (6.6), 117 (12), 77 (25), 75 (20), 74 (5.9), 73 (64), 59 (5.2), 43 (52). <sup>1</sup> H NMR: 0.04 [9H, s, –Si(C*H*3)3]; 1.33 [3H, d, >CHC*H*3, *J*=6.7]; 2.00 [3H, –COC*H*3]; 3.30 [1H, sextuplet, >C*H*CH3, *J*=6.5]; 3.87 and 4.22 [2H, AB part of ABX system,  $>CHCH<sub>2</sub>O-$ ,  $J<sub>AB</sub>=10.8$ ,  $J<sub>AX</sub>$  and  $J<sub>BX</sub>=5.0$ , 8.2]; 5.94 and 5.99 [2H, AB system,  $H_2$  and  $H_3$ ,  $J_{AB} = 6.8$ ]; 7.00– 7.50 [8H, m,  $H_5$ ,  $H_6$ ,  $H_7$ , Ph]; 7.74 [1H, broad d,  $H_8$ , *J*=6.8]. <sup>13</sup>C NMR: −0.19 [3C, −Si(*C*H<sub>3</sub>)<sub>3</sub>]; 15.98 [>CH*C*H3]; 20.83 [–CO*C*H3]; 32.93 [>*C*HCH3]; 45.05  $[C_2]$ ; 68.16  $[-CH_2O-]$ ; 88.74 and 101.21  $[2C, -C\equiv C-$ 

TMS]; 121.58 [2C, *C ortho* of Ph]; 121.83 [*C*8]; 123.01, 124.97, 125.14 and 127.69 [4C,  $C_3$ ,  $C_5$ ,  $C_6$ , *C*7]; 125.64 [*C para* of Ph]; 127.53 [*C*4a]; 129.30 [2C, *C meta* of Ph]; 134.33 [*C*4]; 136.43 [*C*8a]; 150.99 [*C ipso* of Ph]; 151.81 [–O*C*OPh]; 170.91 [–*C*OCH3]. **Characterization of 9g**: IR:  $v_{\text{max}}$  2956, 2168, 1723, 1376, 1285, 1243. GC–MS: *R*<sup>t</sup> 12.28; *m*/*z* 447 (M<sup>+</sup> , 9.7), 371 (7.0), 370 (24), 347 (15), 346 (50), 295 (5.0), 294 (22), 280 (5.3), 279 (9.0), 278 (18), 266 (18), 252 (9.9), 250 (9.5), 236 (6.0), 226 (11), 220 (6.3), 208 (9.8), 194 (19), 193 (6.2), 180 (21), 156 (5.2), 154 (6.1), 151 (7.2), 117 (23), 97 (5.3), 77 (43), 75 (33), 73 (100), 65 (5.9), 59 (7.6), 51 (5.8), 45 (7.6), 43 (87). <sup>1</sup> H NMR: 0.04 [9H, s, -Si(CH<sub>3</sub>)<sub>3</sub>]; 1.20 [3H, d, >CHCH<sub>3</sub>, *J*=7.0]; 2.09 [3H, –COC*H*3]; 3.24 [1H, sextuplet,  $>ECHCH_3$ ,  $J=6.9$ ; 3.87 and 4.22 [2H, AB part of ABX system, >CHC $H_2O$ -,  $J_{AB}$ =10.8,  $J_{AX}$  and  $J_{BX}$ = 5.0, 8.2]; 5.94 and 5.99 [2H, AB system,  $H_2$  and  $H_3$ , *J*AB=6.8]; 7.00–7.50 [8H, m, *H*5, *H*6, *H*7, Ph]; 7.74 [1H, broad d, *H*8, *J*=6.8]. 13C NMR: −0.18 [3C, –Si(*C*H3)3]; 17.49 [>CH*C*H3]; 20.95 [–CO*C*H3]; 32.94 [>*C*HCH3]; 44.92 [*C*2]; 67.81 [–*C*H2O–]; 88.76 and 101.24 [2C, –*CC*–TMS]; 120.96 [*C*8]; 121.58 [2C, *C ortho* of Ph]; 122.96, 124.98 [most likely 2C] and 127.70 [4C, *C*3, *C*5, *C*6, *C*7]; 125.68 [*C para* of Ph]; 127.54 [*C*4a]; 129.32 [2C, *C meta* of Ph]; 134.33 [*C*4]; 136.88 [*C*8a]; 150.98 [*C ipso* of Ph]; 151.84 [–O*C*OPh]; 170.95 [–*C*OCH3].

**4.14.9. 4-{(***S***\*)-[2-(***p***-Methoxyphenoxy)-1-methyl]ethyl}-** 2 -  $\{(R^*)$  -  $[(\text{trimethylsilyl}) \in \text{thynyll}\}$  -  $2H$  - quinoline - 1 - car**boxylic acid phenyl ester, 8h and its epimer, 9h**. Both components of the mixture **8**,**9h**, as a pale yellow oil, could not be separated.  $R_f$  0.39 (8h and 9h) (PE/Et<sub>2</sub>O 8:2, **A**, **B**). Both compounds have however been characterized separately, by protection of **8d** and **9d** (see text and experimental below). **Characterization of 8h**: IR:  $v_{\text{max}}$  3014, 2956, 2169, 1713, 1380, 1301, 1014. GC–MS: (usual conditions, but with fin. temp. 290°C) *R*<sup>t</sup> 14.62; *m*/*z* 512 (8.2), 511 (M<sup>+</sup> , 20), 435 (14); 434 (48), 418 (14), 390 (14), 388 (7.4), 386 (6.6), 374 (8.6), 348 (7.6), 347 (30), 346 (100), 294 (8.1), 280 (6.1), 266 (12), 264 (6.4), 254 (6.2), 252 (14), 250 (8.1), 226 (11), 222 (8.0), 220 (6.1), 194 (28), 193 (6.2), 181 (6.1), 180 (19), 170 (10), 154 (6.8), 151 (11), 137 (20), 124 (15), 123 (14), 109 (18), 107 (19), 97 (6.8), 95 (6.5), 94 (6.6), 92 (7.9), 77 (52), 75 (6.7), 74 (6.7), 73 (73). <sup>1</sup> H NMR: 0.05 [9H, s, –Si(CH<sub>3</sub>)<sub>3</sub>]; 1.44 [3H, d, >CHCH<sub>3</sub>, *J*=6.5]; 3.38 [1H, center of m, >CHCH<sub>3</sub>]; 3.63 [1H, t, >CHC*H*HO–, *J*=8.8]; 3.75 [3H, –OC*H*3]; 4.04 [1H, dd, >CHCH*H*O–, *J*=3.9, 9.0]; 5.94 and 6.03 [2H, AB system,  $H_2$  and  $H_3$ ,  $J_{AB} = 6.7$ ]; 6.80 [4H, s, *p*-anisyl]; 7.15–7.48 [8H, m, *H*<sub>5</sub>, *H*<sub>6</sub>, *H*<sub>7</sub>, Ph]; 7.76 [1H, broad d, *H*<sub>8</sub>, *J*=7.8]. <sup>13</sup>C NMR: −0.17 [3C, −Si(CH<sub>3</sub>)<sub>3</sub>]; 16.28 [>CH*C*H3]; 33.79 [>*C*HCH3]; 45.14 [*C*2]; 55.71 [–O*C*H3]; 72.85 [–*C*H2O–]; 88.72 and 101.24 [2C, –*CC*–TMS]; 114.58 [2C, *C ortho* to –OMe]; 115.65 [2C, *C meta* to –OMe]; 121.59 [2C, *C ortho* of Ph]; 121.79 [*C*8]; 122.95, 124.96 [most likely 2C] and 127.66 [4C, *C*3, *C*5, *C*6, *C*7]; 125.66 [*C para* of Ph]; 129.30 [3C, *C meta* of Ph and, most likely, *C*4a]; 134.32 [*C*4]; 136.59 [*C*8a]; 150.91 [*C ipso* of Ph]; 151.79 [*C*O]; 152.99 and 153.83 [2*C* quat. of *p*-anisyl]. **Char-**

**acterization of 9h**: IR:  $v_{\text{max}}$  3002, 2958, 2168, 1714, 1379, 1324, 1287, 1013. GC–MS: (usual conditions, but with fin. temp. 290°C)  $R_t$  15.06;  $m/z$  511 (M<sup>+</sup>, 16), 435 (13); 434 (35), 418 (11), 390 (10), 388 (8.7), 374 (9.8), 347 (22), 346 (98), 294 (9.7), 266 (19), 252 (15), 250 (9.9), 226 (9.9), 222 (9.4), 204 (8.4), 194 (35), 180 (21), 179 (8.7), 170 (13), 151 (2), 137 (26), 124 (22), 123 (14), 109 (27), 107 (31), 94 (12), 92 (9.3), 77 (56), 75 (8.5), 74 (8.4), 73 (100), 43 (9.1). <sup>1</sup> H NMR: 0.05 [9H, s, -Si(CH<sub>3</sub>)<sub>3</sub>]; 1.38 [3H, d, >CHCH<sub>3</sub>, *J*=7.0]; 3.33 [1H, sextuplet, >C*H*CH3, *J*=6.6]; 3.78 [3H, –OC*H*3]; 3.99 and 4.18 [2H, AB part of ABX system, >CHC*H*<sub>2</sub>O–,  $J_{AB}$ =9.0  $J_{AX}$  and  $J_{BX}$ =5.8, 7.2]; 5.94 and 5.97 [2H, AB system,  $H_2$  and  $H_3$ ,  $J_{AB} = 6.0$ ]; 6.80–6.93 [4H, m, *p*-anisyl]; 7.16–7.48 [8H, m, *H*<sub>5</sub>, *H*<sub>6</sub>, *H*<sub>7</sub>, Ph]; 7.75 [1H, broad d, *H*<sub>8</sub>, *J*=8.2]. <sup>13</sup>C NMR:<br>-0.17 [3C, -Si(CH<sub>3</sub>)<sub>3</sub>]; 17.69 [>CHCH<sub>3</sub>]; 33.65 −0.17 [3C, –Si(*C*H3)3]; 17.69 [>CH*C*H3]; 33.65 [>*C*HCH3]; 44.95 [*C*2]; 55.71 [–O*C*H3]; 72.56 [–*C*H2O–]; 88.66 and 101.42 [2C, –*CC*–TMS]; 114.65 [2C, *C ortho* to –OMe]; 115.79 [2C, *C meta* to –OMe]; 120.79 [*C*8]; 121.59 [2C, *C ortho* of Ph]; 123.03, 124.91 [most likely 2C] and 127.56 [4C,  $C_3$ ,  $C_5$ ,  $C_6$ , *C*7]; 125.66 [*C para* of Ph]; 129.30 [3C, *C meta* of Ph and, most likely,  $C_{4a}$ ; 134.32 [ $C_4$ ]; 136.59 [ $C_{8a}$ ]; 150.91 [*C ipso* of Ph]; 151.79 [*C*O]; 153.06 and 153.96 [2*C* quat. of *p*-anisyl].

**4.14.10. 4-{(***S***\*)-[1-Methyl-2-(***p***-nitrophenoxy)]ethyl}-2-**  $\{(R^*)$  -  $[(\text{trimethylsilyl})\text{ethynyl}] \}$  - 2*H* - quinoline - 1 - car**boxylic acid phenyl ester, 8i and its epimer, 9i**. Preparative thin layer chromatography was performed with  $PE/Et_2O$  (8:2) to give **8i** and **9i** as pale yellow oils.  $R_f$  0.42 (8i) and 0.34 (9i) (PE/Et<sub>2</sub>O 7:3, A, B). **Characterization of 8i**: IR:  $v_{\text{max}}$  2956, 2171, 1721, 1592, 1379, 1332, 1216, 1111, 986, 840. GC–MS: unsuitable for this analysis.  $^1H$  NMR: 0.05 [9H, s, –Si(C*H*3)3]; 1.47 [3H, d, >CHC*H*3, *J*=6.6]; 3.37–3.55 [1H, m, >C*H*CH3]; 3.79 [1H, t, >CHC*H*HO–, *J*= 8.7]; 4.15 [1H, dd, >CHCH*H*O–, *J*=3.9, 8.9]; 5.96 and 6.07 [2H, AB system,  $H_2$  and  $H_3$ ,  $J_{AB} = 6.8$ ]; 6.90 [2H, dt, *H* meta to  $-NO_2$ ,  $J=2.8$ , 9.3]; 7.15–7.48 [8H, m, *H*5, *H*6, *H*7, Ph]; 7.78 [1H, broad d, *H*8, *J*=7.7]; 8.16 [2H, dt, *H* ortho to  $-NO_2$ ,  $J=2.8$ , 9.3]. <sup>13</sup>C NMR: −0.18 [3C, −Si(*CH*<sub>3</sub>)<sub>3</sub>]; 16.09 [>CH*CH*<sub>3</sub>]; 33.52 [>*C*HCH3]; 45.08 [*C*2]; 72.70 [–*C*H2O–]; 88.98 and 100.91  $[2C, -C\equiv C\equiv TMS]$ ; 114.49  $[2C, C \text{ meta to}$  $-NO_2$ ]; 121.50 [2C, *C* ortho of Ph]; 122.35 [C<sub>8</sub>]; 122.73, 125.07, 125.21 and 127.95 [4C, *C*<sub>3</sub>, *C*<sub>5</sub>, *C*<sub>6</sub>, *C*<sub>7</sub>]; 125.78 [*C para* of Ph]; 125.83 [2C, *C ortho* to NO2]; 127.24 [*C*4a]; 129.36 [2C, *C meta* of Ph]; 134.36 [*C*4]; 135.92 [*C*8a]; 141.50 [*C*NO2]; 150.84 [*C ipso* of Ph]; 151.78 [CO]; 163.81 [C para to -NO<sub>2</sub>]. **Characterization of 9i**: IR:  $v_{\text{max}}$  3002, 2177, 1717, 1593, 1342, 1257. GC– MS: unsuitable for this analysis. <sup>1</sup>H NMR: 0.07 [9H, s,  $-Si(CH_3)_3$ ; 1.33 [3H, d,  $>CHCH_3$ ,  $J=6.9$ ]; 3.42 [1H, sextuplet,  $>CHCH_3$ ,  $J=6.6$ ]; 4.14 and 4.31 [2H, AB part of ABX system, >CHC $H_2O$ -,  $J_{AB}$ =9.0,  $J_{AX}$ and  $J_{\text{BX}} = 5.6, 7.1$ ; 5.97 [2H, apparent s,  $H_2$  and  $H_3$ ]; 6.96–7.04 [2H, *H meta* to  $-NO_2$ ]; 7.18–7.48 [8H, m, *H*<sub>5</sub>, *H*<sub>6</sub>, *H*<sub>7</sub>, Ph]; 7.70 [1H, broad d, *H*<sub>8</sub>, *J*=7.9]; 8.15– 8.26 [2H, m, *H ortho* to –NO<sub>2</sub>]. <sup>13</sup>C NMR: –0.20 [3C, –Si(*C*H3)3]; 17.62 [>CH*C*H3]; 33.52 [>*C*HCH3]; 44.91

 $[C_2]$ ; 72.47  $[-CH_2O-]$ ; 88.95 and 101.22  $[2C, -C\equiv C-$ TMS]; 114.55 [2C, *C* meta to –NO<sub>2</sub>]; 121.14 [C<sub>8</sub>]; 121.56 [2C, *C ortho* of Ph]; 122.84, 124.97, 125.15 and 127.88 [4C, *C*3, *C*5, *C*6, *C*7]; 125.75 [*C para* of Ph]; 125.92 [2C, *C* ortho to NO<sub>2</sub>]; 127.36 [C<sub>4a</sub>]; 129.36 [2C, *C meta* of Ph]; 134.44 [*C*4]; 136.86 [*C*8a]; 141.71 [*C*NO2]; 150.96 [*C ipso* of Ph]; 151.84 [*CO*]; 163.85 [*C para* to –NO<sub>2</sub>].

**4.14.11. 4-{(***S***\*)-[1-Methyl-2-trityloxy]ethyl}-2-{(***R***\*)- [(trimethylsilyl)ethynyl]}-2***H***-quinoline-1-carboxylic acid phenyl ester, 8j and its epimer, 9j**. Both components of the mixture **8**,**9j**, as a white foam, could not be separated. By crystallization with  $Et_2O/PE$  nearly pure 8j (containing only 1% of **9j** by HPLC) was obtained. Mp 126.9–125.7°C (Et<sub>2</sub>O/PE).  $R_f$  0.35 (8j and 9j) (PE/Et<sub>2</sub>O 9:1, **A**, **B**). **Characterization of 8j**: IR:  $v_{\text{max}}$  2977, 1712, 1383, 1111, 1071. GC–MS: unsuitable for this analysis. <sup>1</sup> <sup>1</sup>H NMR: 0.13 [9H, s, -Si(CH<sub>3</sub>)<sub>3</sub>]; 1.44 [3H, d, >CHC*H*3, *J*=6.5]; 3.07–3.13 [1H, m, >C*H*CH3]; 3.25– 3.36 [2H, m, >CHC $H_2O$ –]; 6.03 [2H, apparent s,  $H_2$  and *H*<sub>3</sub>]; 7.03 [1H, broad s,  $H_6$ ]; 7.17–7.47 [22H, m,  $H_5$ ,  $H_7$ , Ph, Tr]; 7.83 [1H, broad d,  $H_8$ ,  $J=7.8$ ]. <sup>13</sup>C NMR: −0.14 [3C, –Si(*C*H3)3]; 16.39 [>CH*C*H3]; 34.25 [>*C*HCH3]; 45.15 [*C*2]; 67.26 [–*C*H2O–]; 86.24 [–*C*Ph3]; 88.43 and 101.64 [2C, –*CC*–TMS]; 121.38 [*C*8]; 121.63 [2C, *C ortho* of Ph]; 123.25, 124.81, 125.04 and 127.26 [4C, *C*3, *C*5, *C*6, *C*7]; 125.47 [*C para* of Ph]; 126.74 [3C, *C para* of Tr]; 127.65 and 128.61 [12C, *C ortho* and *meta* of Tr]; 128.22 [*C*4a]; 129.15 [2C, *C meta* of Ph]; 134.29 [*C*4]; 137.72 [*C*8a]; 144.13 [3C, *C ipso* of Tr]; 150.93 [*C ipso* of Ph]; 151.80 [*C*O]. **Characterization of 9j** (this compound was obtained pure after reduction of **30**, see experimental below): IR:  $v_{\text{max}}$  2955, 2922, 1708, 1377, 1305, 1190, 1066, 1017. GC–MS: unsuitable for this analysis. <sup>1</sup> H NMR: 0.07 [9H, s, –Si(C*H*3)3]; 1.19 [3H, d,  $>CHCH_3$ ,  $J=6.6$ ]; 3.08–3.36 [3H, m,  $>CHCH<sub>2</sub>O-$ ]; 5.82 and 5.89 [2H, AB system,  $H<sub>2</sub>$  and *H*3, *J*AB=7.0]; 7.10–7.50 [23H, m, *H*5, *H*6, *H*7, Ph, Tr]; 7.72 [1H, broad d, *H*8, *J*=7.8]. 13C NMR: −0.32 [3C, –Si(*C*H3)3]; 18.06 [>CH*C*H3]; 34.46 [>*C*HCH3]; 44.92  $[C_2]$ ; 67.33 [– $CH_2O$ –]; 86.50 [– $CPh_3$ ]; 88.48 and 101.30 [2C, –*CC*–TMS]; 120.85 [*C*8]; 121.66 [2C, *C ortho* of Ph]; 123.39, 124.84 [most likely 2C] and 127.36 [4C, *C*3, *C*5, *C*6, *C*7]; 125.67 [*C para* of Ph]; 126.87 [3C, *C para* of Tr]; 127.76 and 128.85 [12C, *C ortho* and *meta* of Tr]; 128.36 [*C*4a]; 129.34 [2C, *C meta* of Ph]; 134.16 [*C*4]; 138.02 [*C*8a]; 144.15 [3C, *C ipso* of Tr]; 151.00 [*C ipso* of Ph]; 151.92 [*C*O].

**4.14.12. 2-{(***R***\*)-[3-(Benzyloxy)prop-1-ynyl]}-4-{(***S***\*)-[1 methyl-2-trityloxy]ethyl}-2***H***-quinoline-1-carboxylic acid phenyl ester, 8k and its epimer, 9k**. Both components of the mixture **8**,**9k**, together with the excess of the starting alkyne, as a white foam, could not be separated. Thus only a partial characterization of them was possible. The two diastereoisomers were however separated and fully characterized after trityl and benzyl removal to give **26** and **27**, respectively (see experimental below).  $R_f$  0.64 (8k and 9k) (PE/Et<sub>2</sub>O 6:4, **A**, **B**). **Characteriza**tion of 8,9k: GC–MS: unsuitable for this analysis. <sup>1</sup>H NMR: 1.19 [3H, d, >CHC*H*<sup>3</sup> (**9k**), *J*=6.7]; 1.34 [3H, d, >CHC*H*<sup>3</sup> (**8k**), *J*=6.6]; 2.99–3.35 [3H, m, >C*H*C*H*2O–  $(8,9k)$ ]; 3.89 [2H, d,  $-OCH_2C\equiv C-$  (9k),  $J=1.3$ ]; 4.02  $[2H, d, -OCH, C\equiv C-$  (8k),  $J=1.5$ ]; 4.27 [2H, s, –OC*H*2Ph (**9k**)]; 4.39 [2H, s, –OC*H*2Ph (**8k**)]; 5.84–6.12 [2H, m,  $H_2$  and  $H_3$  (8,9k)]; 6.93 [1H, broad s,  $H_6$ (**8**,**9k**)]; 7.08–7.48 [27H, m, *H*5, *H*6, *H*7, –OPh, Ph, Tr (8,9k)]; 7.75 [1H, broad d,  $H_8$ ,  $J=7.7$  (8,9k)].

**4.14.13. 2-{(***R***\*)-[3-(4-Methoxybenzyloxy)prop-1-ynyl]}- 4-{(***S***\*)-[1-methyl-2-trityloxy]ethyl}-2***H***-quinoline-1-carboxylic acid phenyl ester 8l and its epimer 9l**. Both purified components of the mixture **8**,**9k**, together with a small amount of the excess of the starting alkyne, as an ivory foam, could not be separated. Thus only a partial characterization of them was possible. The two diastereoisomers were however separated and fully characterized after trityl and 4-methoxybenzyl removal to give **26** and **27**, respectively (see experimental below).  $R_f$  0.50 (81 and 91) (PE/Et<sub>2</sub>O 6:4, **A**, **B**). **Characterization of 8**,**9l**: GC–MS: unsuitable for this analysis. <sup>1</sup> H NMR: 1.18 [3H, d, >CHC $H_3$  (9I),  $J=6.6$ ]; 1.35 [3H, d, >CHC*H*<sub>3</sub> (8l), *J*=6.6]; 2.95–3.35 [3H, m, >CHC*H*<sub>2</sub>O– (**8**,**9l**)]; 3.78 [3H, s, –OC*H*<sup>3</sup> (**9l**)]; 3.79 [3H, s, –OC*H*<sup>3</sup> **(8l)**]; 3.87 [2H, broad s,  $-OCH_2C=C$ – **(9l)**]; 3.98 [2H, d,  $-OCH_2C=C-(81), J=1.5$ ; 4.18 [2H, s,  $-OCH_2Ar$  (91)]; 4.28 [2H, s,  $-OCH<sub>2</sub>Ar$  (8I)]; 5.84–6.08 [2H, m,  $H<sub>2</sub>$  and *H*<sub>3</sub> (8,91)]; 6.75–7.48 [27H, m, *H*<sub>5</sub>, *H*<sub>6</sub>, *H*<sub>7</sub>, -OPh, *p*-OMeC6H4, Tr (**8**,**9l**)]; 7.73 [1H, broad d, *H*8, *J*=7.5 (**8**,**9l**)].

**4.14.14. 4-{(***S***)-[1-(Acetoxy)methyl-2-trityloxy]ethyl}-2- {(***R***)-[(trimethylsilyl)ethynyl]}-2***H***-quinoline-1-carboxylic acid phenyl ester, 8m and its epimer, 9m**. Diastereoisomers **8**,**9m** were in part separated by chromatography with  $PE/Et_2O$  95:5 $\rightarrow$ 6:4 both as white foams.  $R_f$  0.36 (8m) and 0.26 (9m) (PE/Et<sub>2</sub>O 8:2, A, B). Characteriza**tion of 8m**:  $[\alpha]_D$  = +115.1 (*c* 1.08, CHCl<sub>3</sub>). IR:  $v_{\text{max}}$  2962, 2168, 1726, 1382, 1325, 1239, 1074, 1023. GC–MS: unsuitable for this analysis. <sup>1</sup>H NMR: 0.005 [9H, s, –Si(C*H*3)3]; 1.94 [3H, s, –COC*H*3]; 3.21 [2H, apparent d,  $-CH_2$ OTr,  $J=4.8$ ]; 3.38  $-CH(CH_2O-)_2$ ,  $J=5.7$ ]; 4.45 and 4.92 [2H, AB part of ABX system,  $-CH<sub>2</sub>OAc$ ,  $J<sub>AB</sub>=11.0$ ,  $J<sub>AX</sub>$  and  $J<sub>BX</sub>=6.1$ , 6.7]; 5.93 [2H, apparent s,  $H_2$  and  $H_3$ ]; 6.91 [1H, broad d, *H*6, *J*=7.2]; 7.08–7.34 [22H, m, *H*5, *H*7, Ph, Tr]; 7.73 [1H, broad d, *H*8, *J*=7.5]. 13C NMR: −0.26 [3C,  $-Si(CH_3)$ ; 20.81 [–COCH<sub>3</sub>]; 38.75 [–CH(CH<sub>2</sub>O–)<sub>2</sub>]; 44.81 [C<sub>2</sub>]; 62.07 and 63.59 [2C, –CH(CH<sub>2</sub>O–)<sub>2</sub>]; 86.38  $[-CPh_3]$ ; 88.64 and 101.05 [2C,  $-C\equiv C-TMS$ ]; 115.28, 122.56, 123.03, 124.99 and 125.19 [5C,  $C_3$ ,  $\overline{C_5}$ ,  $\overline{C_6}$ ,  $\overline{C_7}$ , *C*8]; 121.62 [2C, *C ortho* of Ph]; 125.62 [*C para* of Ph]; 126.90 [3C, *C para* of Tr]; 127.69 [*C*4a]; 127.76 and 128.51 [12C, *C ortho* and *meta* of Tr]; 129.23 [2C, *C meta* of Ph]; 133.41 and 134.41 [2C,  $C_4$  and  $C_{8a}$ ]; 143.74 [3C, *C ipso* of Tr]; 150.83 [*C ipso* of Ph]; 151.82 [–O*C*OPh]; 170.92 [–*C*OCH3]. **Characterization of 9m**:  $[\alpha]_{\text{D}} = -83.1$  (*c* 0.79, CHCl<sub>3</sub>). IR:  $v_{\text{max}}$  2959, 2169, 1715, 1380, 1240. GC-MS: unsuitable for this analysis. <sup>1</sup>H NMR: 0.00 [9H, s,  $-Si(CH_3)_3$ ]; 2.01 [3H, s,  $-COCH_3$ ]; 3.34–3.65 [3H, m, >C*H*C*H*2OTr]; 4.31–4.46 [2H, m,  $-CH_2OAc$ ; 6.04 and 6.07 [2H, AB system,  $H_2$  and  $H_3$ , *J*AB=7.0]; 6.90–7.34 [23H, m, *H*5, *H*6, *H*7, Ph, Tr]; 7.72 [1H, broad d, *H*8, *J*=7.2]. 13C NMR: −0.37 [3C,  $-Si(CH_3)_3$ ; 20.81  $[-COCH_3]$ ; 39.18  $[-CH(CH_2O-)_2]$ ;

44.85 [C<sub>2</sub>]; 62.43 and 64.14 [2C, –CH( $CH_2O$ –)<sub>2</sub>]; 86.77  $[-CPh_3]$ ; 88.79 and 101.74  $[2C, -C \equiv C - TMS]$ ; 121.63 [2C, *C ortho* of Ph]; 122.92, 123.07, 125.01 [most likely 2C] [4C, *C*H of 1,2-dihydroquinoline]; 125.62 [*C para* of Ph]; 126.97 [3C, *C para* of Tr]; [*C*4a: overlapped with other signals]; 127.82 and 128.78 [12C, *C ortho* and *meta* of Tr]; 129.37 [2C, *C meta* of Ph]; 133.26 and 134.21 [2C, *C*<sup>4</sup> and *C*8a]; 143.80 [3C, *C ipso* of Tr]; 150.96 [*C ipso* of Ph]; 151.90 [–O*C*OPh]; 170.84  $[$ -COCH<sub>3</sub>].

**4.14.15. 4-{(***S***)-[1-(Acetoxy)methyl-2-(***p***-methoxyphenyl) diphenylmethoxy]ethyl}-2-{(***R***)-[(trimethylsilyl)ethynyl]}- 2***H***-quinoline-1-carboxylic acid phenyl ester, 8n and its epimer, 9n**. Diastereoisomers **8**,**9n** were in part separated by chromatography with  $PE/Et_2O$  7:3 $\rightarrow$ 3:7 both as yellow foams.  $R_f$  0.49 (8n) and 0.41 (9n) (PE/Et<sub>2</sub>O 6:4, **A**, **B**). **Characterization of 8n**:  $[\alpha]_D = +207.9$  (*c* 1.46, CHCl<sub>3</sub>). IR:  $v_{\text{max}}$  3006, 2393, 1722, 1379, 1189. GC– MS: unsuitable for this analysis. <sup>1</sup>H NMR: 0.03 [9H, s,  $-Si(CH_3)_3$ ; 1.97 [3H, s,  $-COCH_3$ ]; 3.16–3.28 [2H, m,  $-CH_2OMMTr$ ]; 3.39 [1H, quintuplet,  $-CH(CH_2O-)_{2}$ , *J*=5.7]; 3.72 [3H, s, –OC*H*3]; 4.47 and 4.52 [2H, AB part of ABX system,  $-CH_2OAc$ ,  $J_{AB}=10.8$ ,  $J_{AX}$  and  $J_{\text{BX}}=6.1$ , 6.7]; 5.95 [2H, s,  $H_2$  and  $H_3$ ]; 6.74 [2H, apparent d, *H* ortho to –OMe,  $J=8.8$ ]; 6.95 [1H, broad d,  $H_6$ ,  $J=7.0$ ]; 7.11–7.43 [19H, m,  $H_5$ ,  $H_7$ , Ph, 2 Ph of MMTr and *H meta* to –OMe]; 7.74 [1H, broad d,  $H_8$ , *J*=8.0]. <sup>13</sup>C NMR: −0.24 [3C, −Si(*C*H<sub>3</sub>)<sub>3</sub>]; 20.84 [–CO*C*H3]; 38.79 [–*C*H(CH2O–)2]; 44.84 [*C*2]; 55.15  $[-OCH_3]$ ; 62.03 and 63.67 [2C,  $-CH(CH_2O-)$ ]; 86.11 [–*C*Ph2Ar]; 88.66 and 101.08 [2C, –*CC*–TMS]; 113.05 [2C, *C ortho* to –OMe]; 115.29, 122.54, 123.05, 124.98 and 125.20 [5C, *C*3, *C*5, *C*6, *C*7, *C*8]; 121.65 [2C, *C ortho* of Ph]; 125.64 [*C para* of Ph]; 126.77 [2C, *C para* of Ph of MMTr]; 127.67 [*C*4a]; 127.78, 128.23 and 128.28 [8C, *C ortho* and *meta* of Ph of MMTr]; 129.25 [2C, *C meta* of Ph]; 130.21 [2C, *C meta* to –OMe]; 133.44 and 134.43 [2C, *C*<sup>4</sup> and *C*8a]; 135.43 [*C para* to –OMe]; 144.25 and 144.32 [2C, *C ipso* of MMTr]; 150.86 [*C ipso* of Ph]; 151.83 [–O*C*OPh]; 158.44 [*C*OMe of MMTr]; 170.94  $[-COCH<sub>3</sub>]$ . **Characterization of 9n**:  $[\alpha]_D = -179.3$  (*c* 0.44, CHCl<sub>3</sub>). IR:  $v_{\text{max}}$  2928, 2854, 2171, 1725, 1377, 1248, 1068, 1018. GC-MS: unsuitable for this analysis. <sup>1</sup>H NMR: −0.11 [9H, s, −Si(CH<sub>3</sub>)<sub>3</sub>]; 1.90 [3H, s, −COCH<sub>3</sub>]; 3.32–3.50 [3H, m, >C*H*C*H*2OMMTr]; 3.78 [3H, s, –OC*H*3]; 4.26 and 4.26 [2H, AB part of ABX system,  $-CH_2OAc$ ,  $J_{AB} = 11.6$ ,  $J_{AX}$  and  $J_{BX} = 6.6$ , 6.6]; 5.92 and 5.95 [2H, AB system,  $H_2$  and  $H_3$   $J_{AB} = 7.0$ ]; 6.81 [2H, dt, *H ortho* to –OMe, *J*=8.8]; 7.13–7.48 [20H, m, *H*5,  $H_6$ ,  $H_7$ , Ph, 2 Ph of MMTr and *H meta* to –OMe]; 7.30 [1H, broad d,  $H_8$ ,  $J=7.0$ ]. <sup>13</sup>C NMR: −0.33 [3C,  $-Si(CH_3)$ ; 20.83 [–COCH<sub>3</sub>]; 39.26 [–CH(CH<sub>2</sub>O–)<sub>2</sub>]; 44.87 [*C*2]; 55.19 [–O*C*H3]; 62.39 and 64.22 [2C,  $-CH(CH_2O-)$ <sub>2</sub>]; 86.52 [–*C*Ph<sub>2</sub>Ar]; 88.80 and 101.82 [2C, –*CC*–TMS]; 113.09 [2C, *C ortho* to –OMe]; 121.66 [2C, *C ortho* of Ph]; 122.97, 123.11, 125.01 [most likely 2C] [4C, *C*H of 1,2-dihydroquinoline]; 125.75 [*C para* of Ph]; 126.85 [2C, *C para* of Ph of MMTr]; 127.67 [*C*4a]; 127.82, and 128.53 [8C, *C ortho* and *meta* of Ph of MMTr]; 129.38 [2C, *C meta* of Ph]; 130.48 [2C, *C* *meta* to –OMe]; 133.35 and 134.24 [2C,  $C_4$  and  $C_{8a}$ ]; 135.55 [*C para* to –OMe]; 144.25 and 144.37 [2C, *C ipso* of MMTr]; 150.98 [*C ipso* of Ph]; 151.92 [–O*C*OPh]; 158.53 [*C*OMe of MMTr]; 170.87 [–*C*OCH3].

**4.14.16. 4-{(***S***)-[1-(Triethylsilyloxy)methyl-2-trityloxy] ethyl}-2-{(***R***)-[(trimethylsilyl)ethynyl]}-2***H***-quinoline-1 carboxylic acid phenyl ester, 8o and its epimer, 9o**. Both components of the mixture **8**,**9o**, as a yellow oil, could not be separated.  $R_f$  0.85 (**8o** and **9o**) (PE/Et<sub>2</sub>O 6:4, **A**, **B**). **Characterization of 8,9o**: IR:  $v_{\text{max}}$  2950, 2313, 2170, 1712, 1594, 1447, 1380, 1304, 1084. GC–MS: unsuitable for this analysis. <sup>1</sup>H NMR: -0.14 [9H, s, -Si(CH<sub>3</sub>)<sub>3</sub> (**9o**)]; 0.02 [9H, s, –Si(C*H*3)3 (**8o**)]; 0.50 [6H, broad q,  $-Si(CH_2CH_3)$ <sub>3</sub>,  $J=8.0$  (9o)]; 0.56 [6H, broad q,  $-Si(CH_2CH_3)_3$ ,  $J=8.0$  (80)]; 0.83 [9H, t,  $-Si(CH_2CH_3)_3$ , *J*=7.9 (90)]; 0.91 [9H, t, -Si(CH<sub>2</sub>CH<sub>3</sub>)<sub>3</sub>, *J*=7.9 (80)]; 3.12–3.38 [2H, m, –C*H*2OTr (**8**,**9o**)]; 3.53–3.81 [1H, m,  $-CH(CH_2O-)$ <sub>2</sub> (8,9o)]; 3.90–4.04 [2H, m,  $-CH_2OTES$ ]; 5.86 and 5.89 [2H, AB system,  $H_2$  and  $H_3$ ,  $J_{AB} = 6.8$ (**9o**)]; 5.93 and 5.93 [2H, AB system,  $H_2$  and  $H_3$ ,  $J_{AB}$ =6.4 (**8o**)]; 6.90 [1H, broad d,  $H_6$ ,  $J=6.6$  (**8.9o**)]; 7.08–7.49 [22H, m, *H*5, *H*7, Ph, Tr (**8**,**9o**)]; 7.73 [1H, broad d, *H*8, *J*=7.0 (**8**,**9o**)]. 13C NMR: −0.37 [3C, –Si(*C*H3)3 (**9o**)]; −0.24 [3C, –Si(*C*H3)3 (**8o**)]; 4.31 [3C, –Si(*C*H2CH3)3 (**9o**)]; 4.40 [3C, –Si(*C*H2CH3)3 (**8o**)]; 6.75 [3C, -Si(CH<sub>2</sub>CH<sub>3</sub>)<sub>3</sub> (9o)]; 6.88 [3C, -Si(CH<sub>2</sub>CH<sub>3</sub>)<sub>3</sub> (8o)]; 42.38 [–*C*H(CH2O–)2 (**8o**)]; 42.55 [–*C*H(CH2O–)2 (**9o**)]; 44.86 [C<sub>2</sub> (8o)]; 44.98 [C<sub>2</sub> (9o)]; 62.49, 62.82 and 62.98 [4C, –CH(*C*H2O–)2 (**8**,**9o**)]; 86.22 [–*C*Ph3 (**9o**)]; 86.55 [–*C*Ph3 (**8o**)]; 88.24 and 101.44 [2C, –*CC*–TMS (**8o**)]; 88.55 and 101.05 [2C, –*CC*–TMS (**9o**)]; 121.61 [2C, *C ortho* of Ph (**9o**)]; 121.67 [2C, *C ortho* of Ph (**8o**)]; 122.03, 123.33, 124.85, 127.30 and 128.88 [5C, *C*<sub>3</sub>, *C*<sub>5</sub>, *C*6, *C*7, *C*<sup>8</sup> (**8o**)]; 122.03, 123.45, 125.30, 127.46 and 128.88 [5C, *C*3, *C*5, *C*6, *C*7, *C*<sup>8</sup> (**9o**)]; 125.53 [*C para* of Ph (**8o**)]; 125.64 [*C para* of Ph (**9o**)]; 126.76 [3C, *C para* of Tr (**8**,**9o**)]; 127.66, 127.69 and 128.63 [12C, *C ortho* and *meta* of Tr (**8**,**9o**)]; 129.33 [2C, *C meta* of Ph (**8o**)]; 129.45 [2C, *C meta* of Ph (**9o**)]; 134.21 and 134.86 [2C, *C*<sup>4</sup> and *C*8a (**9o**)]; 134.26 and 134.50 [2C, *C*<sup>4</sup> and *C*8a (**8o**)]; 144.02 [3C, *C ipso* of Tr (**8o**)]; 144.11 [3C, *C ipso* of Tr (**9o**)]; 150.89 [*C ipso* of Ph (**8o**)], 151.00 [*C ipso* of Ph (**9o**)]; 151.86 [*C*O (**8**,**9o**)].

# **4.15. (2***S***\*,3***S***\*,4***R***\*)-3,4-Epoxy-4-{(***S***\*)-[1-methyl-2 trityloxy]ethyl}-2-[(trimethylsilyl)ethynyl]-3,4-dihydro-2***H***-quinoline-1-carboxylic acid phenyl ester, 11 and its epimer, 12**

A solution of  $\mathbf{8}$  or  $\mathbf{9}$  (324 mg, 500  $\mu$ mol) in dry  $\text{CH}_2\text{Cl}_2$ (10 ml) was cooled to 0°C and treated with *m*CPBA (1.0 mmol). After stirring until complete (about 3 h) the reaction was quenched with  $Me<sub>2</sub>S$  (86  $\mu$ l, 1.17 mmol) and stirred at rt in the presence of  $NAHCO<sub>3</sub>$  sat. sol. for an additional 15 min. Usual extraction with  $Et<sub>2</sub>O$  was followed by chromatography with  $PE/Et<sub>2</sub>O$  (85:15) to give **11** (92% yield) as a white solid, which was crystallized from  $PE/Et_2O$ , and 12 (90% yield) as a pale yellow oil. **Characterization of 11**: Mp: 151.5–151.9°C (PE/ Et<sub>2</sub>O).  $R_f$  0.41 (PE/Et<sub>2</sub>O 8:2, A, B). IR:  $v_{\text{max}}$  2963, 2928,

2176, 1721, 1488, 1191. GC–MS: unsuitable for this analysis. <sup>1</sup> H NMR: −0.02 [9H, s, –Si(C*H*3)3]; 1.27 [3H, d, >CHCH<sub>3</sub>,  $J=6.2$ ]; 2.94–3.06 and 3.43–3.53 [3H, m, >C*H*C*H*2O–]; 3.88 [1H, d, *H*3, *J*=3.2]; 5.71 [1H, d, *H*2,  $J=3.0$ ]; 7.08–7.50 [24H, m, aromatics]. <sup>13</sup>C NMR: −0.44 [3C, –Si(*C*H3)3]; 15.25 [>CH*C*H3]; 33.02 [>*C*HCH3]; 44.75 [*C*2]; 58.65 [*C*4]; 63.82 [*C*3]; 64.01  $[-CH_2O-]; 86.92$   $[-CPh_3]; 91.71$  and  $99.22$   $[2C, -C=C-$ TMS]; 121.55 [2C, *C ortho* of Ph]; 125.49, 125.55, 127.29, 128.08 and 128.76 [5C,  $C_5$ ,  $C_6$ ,  $C_7$ ,  $C_8$ ,  $C_9$  *para* of Ph]; 127.00 [3C, *C para* of Tr]; 127.49 [*C*4a]; 127.73 and 128.85 [12C, *C ortho* and *meta* of Tr]; 129.24 [2C, *C meta* of Ph]; 135.54 [*C*8a]; 144.20 [3C, *C ipso* of Tr]; 151.20 [*C ipso* of Ph]; 153.43 [*C*O]. **Characterization of 12**:  $R_f$  0.41 (PE/Et<sub>2</sub>O 8:2, **A, B**). IR:  $v_{\text{max}}$  3005, 2396, 1716, 1379, 1323, 1190, 1065. GC–MS: unsuitable for this analysis. <sup>1</sup> H NMR: −0.01 [9H, s, –Si(C*H*3)3]; 1.16 [3H, d,  $\geq$ CHCH<sub>3</sub>,  $J=6.8$ ]; 2.81–2.91 [1H, m,  $>CHCH<sub>2</sub>O$ -]; 3.16 and 3.30 [2H, AB part of ABX system, >CHCH<sub>2</sub>O–,  $J_{AB}$ =9.7,  $J_{AX}$  and  $J_{BX}$ =5.5, 7.2]; 3.85 [1H, d, *H*3, *J*=3.1]; 5.72 [1H, d, *H*2, *J*=2.9]; 7.05–7.49 [24H, m, aromatics]. 13C NMR: −0.49 [3C, –Si(*C*H3)3]; 12.76 [>CH*C*H3]; 33.84 [>*C*HCH3]; 44.80  $[C_2]$ ; 57.18  $[C_4]$ ; 63.52  $[C_3]$ ; 64.82  $[-CH_2O-]$ ; 86.88 [–*C*Ph3]; 91.35 and 98.97 [2C, –*CC*–TMS]; 121.58 [2C, *C ortho* of Ph]; 125.57 [most likely 2C], 127.02, and 127.27 [4C, 3C (between  $C_5$ ,  $C_6$ ,  $C_7$ ,  $C_8$ , while one is overlapped with other signals), *C para* of Ph]; 127.50 [*C*4a]; 127.86 [3C, *C para* of Tr]; 127.94 and 128.71 [12C, *C ortho* and *meta* of Tr]; 129.26 [2C, *C meta* of Ph]; 135.44 [*C*8a]; 144.12 [3C, *C ipso* of Tr]; 146.88 [*C ipso* of Ph]; 151.11 [*C*O].

# **4.16. (2***S***\*,3***S***\*,4***S***\*)-3-Hydroxy-4-{(***R***\*)-[2-hydroxy-1 methyl]ethyl}-4-methoxy-2-[(trimethylsilyl)ethynyl]-3,4 dihydro-2***H***-quinoline-1-carboxylic acid phenyl ester, 13 and its epimer, 14**

A suspension of  $11$  or  $12$  ( $122$  mg,  $184$   $\mu$ mol) in dry methanol (5 ml) was cooled to 0°C and treated with  $p$ TSA (35 mg, 184  $\mu$ mol). After 5 min the reaction flask was allowed to stir at rt for 2 h and gradually a pale yellow solution was obtained. Neutralization with solid  $NaHCO<sub>3</sub>$  (16 mg, 184  $\mu$ mol) was followed by careful evaporation of methanol under reduced pressure at  $\approx$  15°C. The residue was partitioned between water and  $Et<sub>2</sub>O$  and extracted as usual. Chromatography with  $PE/Et<sub>2</sub>O (1:1)$  gave the corresponding diols 13 (50 mg, 60% yield) or **14** (42 mg, 51% yield) as pale yellow oils. **Characterization of 13:**  $R_f$  0.36 (PE/Et<sub>2</sub>O 1:1, **A**, **B**). IR:  $v_{\text{max}}$  3309, 2964, 2174, 1723, 1368, 1324, 1098. GC–MS: *R*<sub>t</sub> 13.48; *m*/*z* 404 (M<sup>+</sup>−49, 5.6), 394 (5.3), 377 (8.5), 376 (30), 300 (14), 256 (7.5), 216 (6.1), 202 (13), 190 (6.6), 170 (5.6), 169 (6.6), 168 (44), 162 (7.8), 160 (5.1), 158 (15), 146 (14), 144 (5.5), 140 (5.6), 132 (6.5), 130 (24), 97 (9.8), 95 (8.0), 94 (28), 89 (8.2), 83 (7.7), 77 (31), 75 (16), 74 (9.1), 73 (100), 66 (5.2), 65 (7.7), 59 (12), 51 (5.4), 45 (8.7), 43 (7.3), 39 (6.1). <sup>1</sup> H NMR: 0.17 [9H, s,  $-Si(CH_3)$ <sub>3</sub>]; 0.68 [3H, d, >CHC*H*<sub>3</sub>, *J*=7.0]; 2.01 [1H, center of m, >C*H*CH2O–]; 2.04 [1H, s, –O*H*]; 3.44 [3H, s, –OC*H*3]; 3.50–3.88 [1H, m; became a dd at 3.52,

 $J=2.5$ , 10.9, after exchange with D<sub>2</sub>O, –C*H*HOH]; 3.98–4.10 [2H, m; became a t at 4.00, *J*=9.6, –CHHOH and a d at 4.02,  $J=9.4$ ,  $H<sub>3</sub>$ , after exchange with D<sub>2</sub>O]; 4.96 [1H, d,  $H_2$ ,  $J=9.4$ ]; 5.97 [1H, broad s, –O*H*]; 7.16–7.42 [8H, m,  $H_5$ ,  $H_6$ ,  $H_7$  and Ph]; 7.72 [1H, broad d, *H*8, *J*=7.3]. 13C NMR: −0.01 [3C, –Si(*C*H3)3]; 13.69 [>CH*C*H3]; 38.45 [>*C*HCH3]; 50.55 [–O*C*H3]; 52.03 [*C*2]; 63.62 [–*C*H2O–]; 74.32 [*C*3]; 80.71 [*C*4]; 87.62 and 104.59 [2C, –*CC*–TMS]; 121.58 [2C, *C ortho* of Ph]; 124.82, 125.46, 126.02, and 127.38 [4C,  $C_5$ ,  $C_6$ ,  $C_7$ , *C*8]; 125.71 [*C para* of Ph]; 129.34 [2C, *C meta* of Ph]; 133.07 and 134.40 [2C,  $C_{4a}$  and  $C_{8a}$ ]; 151.14 [*C ipso* of Ph]; 152.56 [CO]. **Characterization of 14**:  $R_f$  0.39 (PE/ Et<sub>2</sub>O 4:6, **A, B**). IR:  $v_{\text{max}}$  3302, 2170, 1725, 1365, 1088. GC–MS: *R*<sup>t</sup> 13.52; *m*/*z* 404 (M<sup>+</sup> −49, 6.1), 394 (13), 378 (8.4), 377 (27), 376 (100), 334 (9.2), 301 (5.3), 300 (18), 256 (19), 244 (5.4), 242 (6.8), 240 (5.3), 226 (5.6), 216 (6.1), 210 (6.8), 202 (13), 190 (6.9), 170 (6.7), 169 (6.8), 168 (49), 162 (8.1), 160 (5.8), 158 (14), 146 (16), 144 (6.7), 140 (5.9), 132 (5.9), 130 (23), 115 (5.5), 97 (8.6), 95 (7.6), 94 (29), 91 (5.3), 90 (5.2), 89 (8.6), 83 (7.5), 77 (30), 75 (15), 74 (7.6), 73 (89), 66 (6.0), 65 (7.6), 59 (11), 55 (5.2), 45 (7.5), 43 (6.4), 39 (5.9). <sup>1</sup> H NMR: 0.13 [9H, s,  $-Si(CH_3)$ <sub>3</sub>]; 1.15 [3H, d,  $>CHCH_3$ ,  $J=7.3$ ]; 2.15 [1H, center of m,  $>CHCH<sub>2</sub>O-$ ]; 2.87 [1H, broad s,  $-OH$ ]; 3.36 [3H, s, –OC*H*3]; 3.32–3.43 [1H, m; became a dd at 3.38,  $J=5.5$ , 11.1, after exchange with  $D_2O$ , –C*H*HOH]; 3.82 [1H, broad d, *J*=10.7; became a dd,  $J=2.6$ , 11.1, after exchange with D<sub>2</sub>O, –CH*HOH*]; 4.05 [1H, broad s, –OH]; 4.21–4.25 [1H, m, became a d at 4.23,  $J=7.6$ , after exchange with D<sub>2</sub>O,  $H_3$ ]; 5.07 [1H, d, *H*<sub>2</sub>, *J*=7.7]; 7.14–7.43 [8H, m, *H*<sub>5</sub>, *H*<sub>6</sub>, *H*<sub>7</sub> and Ph]; 7.75 [1H, broad d,  $H_8$ ,  $J=8.3$ ]. <sup>13</sup>C NMR: −0.16 [3C, –Si(*C*H3)3]; 12.63 [>CH*C*H3]; 41.09 [>*C*HCH3]; 50.80 [–O*C*H3]; 52.66 [*C*2]; 64.79 [–*C*H2O–]; 75.36 [*C*3]; 81.86 [*C*4]; 88.98 and 103.82 [2C, –*CC*–TMS]; 121.57 [2C, *C ortho* of Ph]; 124.91, 125.58, 125.72 [most likely 2C] and 127.80, [5C, *C<sub>5</sub>, C<sub>6</sub>, C<sub>7</sub>, C<sub>8</sub>, <i>C* para of Ph]; 129.35 [2C, *C* meta of Ph]; 131.12 and 135.58 [2C,  $C_{4a}$  and  $C_{8a}$ ]; 151.15 [*C ipso* of Ph]; 152.42 [*C*O].

# **4.17. (2***S***\*,3***S***\*,4***S***\*)-2-Ethynyl-3-hydroxy-4-{(***R***\*)-[2 hydroxy-1-methyl]ethyl}-4-methoxy-3,4-dihydro-2***H***quinoline-1-carboxylic acid methyl ester, 15**

The oxirane opening was performed under the same conditions described for the preparation of **13** and **14**, starting from 301 mg  $(453 \text{ µmol})$  of 11. Crude product (or either chromatographed as previously reported), obtained after solvent removal, was dissolved in dry methanol (5 ml) and heated at 40°C for 3 h, after solid  $NaHCO<sub>3</sub>$  (241 mg, 2.87 mmol) has been added. The mixture was concentrated under reduced pressure, partitioned between water and  $Et<sub>2</sub>O$  and extracted as usual. Chromatography with  $PE/Et_2O$  1:1 $\rightarrow Et_2O$  gave, as prevailing product, **15** (70 mg, 50% from **11**) as a pale yellow oil.  $R_f$  0.14 (PE/Et<sub>2</sub>O 4:6, **A, B**). IR:  $v_{\text{max}}$ 3303, 2955, 1702, 1440, 1337, 1192, 1097. GC–MS: *R*<sup>t</sup> 9.48;  $m/z$  319 (M<sup>+</sup>, 3.6), 271 (5.0), 270 (28), 261 (15), 260 (92), 258 (10), 243 (13), 242 (84), 232 (5.8), 228 (24),

209 (6.9), 202 (14), 201 (16), 200 (100), 192 (11), 187 (6.6), 186 (8.8), 185 (23), 184 (7.4), 177 (5.6), 174 (6.3), 172 (6.2), 170 (8.4), 169 (5.7), 168 (17), 167 (8.9), 162 (5.5), 158 (7.4), 156 (8.3), 154 (10), 146 (25), 144 (9.7), 143 (11), 142 (7.1), 141 (18), 140 (7.4), 132 (11), 130 (17), 129 (11), 128 (12), 117 (6.8), 116 (5.9), 115 (18), 91 (5.1), 77 (7.2), 59 (10). <sup>1</sup> H NMR: 0.66 [3H, d, >CHC*H*3, *J*=7.1]; 1.90 [1H, broad s, –O*H*]; 1.93 [1H, center of m, >C*H*CH2O–]; 2.41 [1H, d,  $-C=CH$ ,  $J=2.0$ ]; 3.41 [3H, s,  $-OCH_3$ ]; 3.58 [1H, dd,  $-CHHOH$ ,  $J=2.9$ , 10.8]; 3.82 [3H, s,  $-CO_2CH_3$ ]; 3.90–4.10 [2H, m; became a d at 4.02,  $J=9.5$ ,  $H_3$  and a t at 4.65, *J*=9.7, –CH*H*OH, after exchange with D<sub>2</sub>O]; 4.81 [1H, dd, H<sub>2</sub>, J=1.7, 9.5]; 6.13 [1H, broad s, –O*H*]; 7.14–7.38 [3H, m, *H*5, *H*6, *H*7]; 7.59 [1H, broad d,  $H_8$ ,  $J=7.7$ ]. <sup>13</sup>C NMR: 13.56 [>CHCH<sub>3</sub>]; 38.14 [>*C*HCH3]; 50.45 [–O*C*H3]; 50.85 [*C*2]; 53.44 [–CO2*C*H3]; 63.41 [–*C*H2O–]; 70.51 [–C*C*H]; 74.27 [*C*3]; 80.43 [*C*4]; 83.45 [–*C*CH]; 124.58, 125.41, 125.90 and 127.27, [4C,  $C_5$ ,  $C_6$ ,  $C_7$ ,  $C_8$ ]; 132.78 and 134.28 [2C, *C*4a and *C*8a]; 154.68 [*C*O].

# **4.18. (6***R***\*,6a***S***\*,11***R***\*,11a***S***\*)-6-Ethynyl-11a-methoxy-8,8,11-trimethyl-6a,10,11,11a-tetrahydro-6***H***-7,9-dioxa-5-aza-cyclohepta[***a***]naphthalene-5-carboxylic acid methyl ester, 16**

A solution of  $15$  (54 mg, 170  $\mu$ mol) in dry CH<sub>2</sub>Cl<sub>2</sub> (4 ml) was cooled to 0°C and treated with 2 methoxypropene  $(33 \mu l, 341 \mu mol)$  and  $p$ -TSA  $(34$ -ml of a 0.1 M sol. in THF). After 10 min the solution was diluted with  $5\%$  aq NaHCO<sub>3</sub> and extracted with Et<sub>2</sub>O. Chromatography with PE/Et<sub>2</sub>O 7:3 furnished pure **16** as a pale yellow oil.  $R_f$  0.14 (PE/Et<sub>2</sub>O) 4:6, **A**, **B**). IR: max 3303, 2991, 2955, 2124, 1709, 1439, 1383, 1329, 1133, 1046. GC–MS: *R*<sup>t</sup> 8.94; *m*/*z* 359 (M<sup>+</sup> , 29), 301 (9.8), 286 (10), 273 (17), 272 (77), 258 (10), 242 (23), 240 (20), 234 (58), 232 (16), 231 (12), 230 (83), 229 (10), 218 (21), 215 (12), 212 (16), 205 (59), 202 (45), 198 (14), 192 (16), 187 (9.6), 184 (16), 182 (18), 181 (16), 180 (22), 174 (9.6), 172 (9.7), 170 (21), 168 (14), 167 (26), 160 (11), 156 (12), 154 (15), 146 (27), 144 (9.6), 143 (12), 142 (9.6), 141 (10), 140 (11), 130 (14), 129 (12), 128 (19), 127 (10), 115 (21), 101 (12), 91 (13), 90 (13), 77 (24), 73 (14), 69 (22), 59 (100), 51 (15), 45 (17), 43 (62), 42 (12), 41 (34), 39 (27). <sup>1</sup> H NMR: 0.59 [3H, d, >CHC*H*3, *J*= 6.8]; 1.45 and 1.43 [6H, 2 s,  $>C(CH_3)_2$ ]; 2.17 [1H, d,  $-C\equiv CH, J=2.5$ ]; 2.37 [1H, center of m,  $>CHCH_2O-$ ]; 3.30 [3H, s, –OC*H*3]; 3.65 and 3.67 [2H, AB part of ABX system,  $-CH_2O^-$ ,  $J_{AB} = 12.0$ ,  $J_{AX}$  and  $J_{BX} = 3.9$ , 13.2]; 3.83 [3H, s,  $-CO_2CH_3$ ]; 4.48 [1H, d,  $H_{6a}$ ,  $J=$ 3.0]; 5.44 [1H, t,  $H_6$ ,  $J=2.7$ ]; 7.15 [1H, dt,  $H_2$ ,  $J=1.4$ , 7.4]; 7.24–7.34 [2H, m, *H*1, *H*3]; 7.53 [1H, broad d, *H*4, *J*=8.5]. 13C NMR: 12.64 [>CH*C*H3]; 24.97 and 25.40 [2C, >C(*C*H3)2]; 39.82 [>*C*HCH3]; 50.56 [*C*6]; 51.46 [-OCH<sub>3</sub>]; 53.14 [-CO<sub>2</sub>CH<sub>3</sub>]; 66.06 [-CH<sub>2</sub>O-]; 73.21 [-C=CH]; 77.24 [C<sub>6a</sub>]; 79.70 [C<sub>11a</sub>]; 80.74  $[-C\equiv CH]$ ; 101.87  $[>C(CH_3)_2]$ ; 123.57, 125.01, 127.42 and 128.20, [4C, *C*1, *C*2, *C*3, *C*4]; 130.66 and 136.59 [2C,  $C_{4a}$  and  $C_{11b}$ ]; 154.43 [*CO*].

# **4.19. (2***S***\*,3***S***\*,4***S***\*)-3-Acetoxy-4-{(***R***\*)-[2-acetoxy-1 methyl]ethyl}-4-methoxy]-2-[(trimethylsilyl)ethynyl]-3,4 dihydro-2***H***-quinoline-1-carboxylic acid phenyl ester, 17**

A solution of **13** (16 mg, 35  $\mu$ mol) in dry CH<sub>2</sub>Cl<sub>2</sub> (3 ml) was treated, at  $0^{\circ}$ C, with triethylamine (15 µl, 106 µmol), 4-dimethylaminopyridine (1.0 mg, 8.2 µmol) and acetic anhydride (10  $\mu$ l, 106  $\mu$ mol). After 5 min, the solution was stirred at rt until the reaction was complete (about 3 h), and then partitioned between water and  $Et<sub>2</sub>O$  and finally extracted with the same solvent. Chromatography with  $PE/Et_2O$  (7:3) gave pure 17 (12) mg,  $63\%$ ) as a pale yellow oil.  $R_f$  0.57 (PE/Et<sub>2</sub>O 1:1, A, **B**). IR:  $v_{\text{max}}$  3015, 2962, 2177, 1728, 1374, 1323, 1194, 1030. GC–MS: *R*<sup>t</sup> 13.57; *m*/*z* 446 (M<sup>+</sup> −91, 2.5), 376 (100), 268 (6.9), 210 (5.2), 202 (12), 170 (6.0), 117 (6.4), 77 (14), 75 (10), 73 (30), 43 (5.2). <sup>1</sup> H NMR: 0.07 [9H, s,  $-Si(CH_3)$ ; 0.90 [3H, d, >CHCH<sub>3</sub>, J=6.9]; 2.03 and 2.19 [6H, 2 s, –COC*H*3]; 2.28–2.45 [1H, m, >C*H*CH2O–]; 3.24 [3H, s, –OC*H*3]; 3.87 [1H, dd, –C*H*HOAc, *J*=9.3, 10.9]; 4.68 [1H, dd, –CH*H*OAc, *J*=3.7, 11.0]; 5.30 [1H, d, *H*3, *J*=6.4]; 5.61 [1H, d, *H*2, *J*=6.4]; 7.17–7.43 [8H, m,  $H_5$ ,  $H_6$ ,  $H_7$  and Ph]; 7.72 [1H, broad d,  $H_8$ ,  $J=8.0$ ].

# **4.20. (1***S***\*,3a***S***\*,4***S***\*,9b***S***\*)-9b-Methoxy-1-methyl-2 oxo-4-[(trimethylsilyl)ethynyl]-1,3a,4,9b-tetrahydro-2***H***furo[2,3-***c***]quinoline-5-carboxylic acid phenyl ester, 18 and its epimer, 19**

A solution of 13 or 14 (35 mg, 77  $\mu$ mol) in dry  $CH_2Cl_2$ (2 ml) was stirred for 5 min at rt in the presence of powdered 4 A molecular sieves (10 mg). Then 4-methylmorpholine- $N$ -oxide (25  $\mu$ l, 232  $\mu$ mol) and tetrapropylammonium perruthenate  $(2.7 \text{ mg}, 7.7 \text{ µmol})$  were added and the resulting mixture was stirred for 30 min. After concentration under reduced pressure, direct chromatography with  $PE/Et_2O$  (8:2) gave 18 (22 mg, 62% yield) or **19** (21 mg, 61% yield) as pale yellow oils. **Characterization of 18:**  $R_f$  0.41 (PE/Et<sub>2</sub>O 7:3, A, B). IR: max 2960, 2180, 1784, 1727, 1381, 1327, 1246, 1045. GC–MS: *R*<sub>t</sub> 12.90; *m*/*z* 449 (M<sup>+</sup>, 18), 357 (13), 356 (48), 322 (13), 321 (37), 300 (24), 268 (8.8), 228 (11), 196 (19), 190 (18), 180 (9.5), 173 (11), 172 (8.2), 168 (9.1), 167 (14), 158 (8.2), 157 (7.7), 146 (40), 142 (9.7), 141 (29), 130 (11), 97 (58), 90 (9.1), 89 (15), 83 (12), 77 (47), 75 (13), 74 (8.6), 73 (100), 69 (7.3), 65 (9.6), 59 (19), 55 (8.1), 51 (8.3), 45 (9.0), 43 (13), 39 (8.2). <sup>1</sup> H NMR: −0.06 [9H, s, –Si(C*H*3)3]; 1.21 [3H, d, >CHC*H*3, *J*=7.4]; 2.52 [1H, q,  $>CHCH_3$ ,  $J=7.4$ ]; 3.38 [3H, s,  $-OCH_3$ ]; 5.10 [1H, s, *H*3a]; 5.73 [1H, s, *H*4]; 7.13–7.47 [8H, m, *H*7,  $H_8$ ,  $H_9$  and Ph]; 7.66 [1H, broad d,  $H_6$ ,  $J=7.4$ ]. <sup>13</sup>C NMR: −0.65 [3C, –Si(*C*H3)3]; 7.20 [>CH*C*H3]; 47.58 [>*C*HCH3]; 50.82 [*C*4]; 52.24 [–O*C*H3]; 79.74 [*C*9b]; 85.93  $[C_{3a}]$ ; 93.71 and 99.16  $[2\overline{C}, -\overline{C} = \overline{C}$ -TMS]; 121.49 [2C, *C ortho* of Ph]; 124.43, 125.87, 125.97, 126.17 and 128.26 [5C, *C*6, *C*7, *C*8, *C*9, *C para* of Ph]; 129.46 [2C, *C meta* of Ph]; 131.68 and 135.27 [2C,  $C_{5a}$  and  $C_{9a}$ ]; 151.14 [*C ipso* of Ph]; 152.56 [*C*O]; 176.12 [*C*O of lactone]. **Characterization of 19**:  $R_f$  0.38 (PE/Et<sub>2</sub>O 6:4, **A**, **B**). IR: max 2425, 2182, 1784, 1727, 1381, 1330, 1314, 1250, 1053. GC–MS (this lactone, unlike **18**, partially epimerizes in the column):  $R_t$  13.07;  $m/z$  449 (M<sup>+</sup> , 17), 357 (11), 356 (41), 324 (12), 321 (21), 300 (18),

268 (8.0), 228 (11), 196 (18), 190 (18), 180 (11), 173 (8.4), 172 (6.8), 168 (9.8), 167 (12), 158 (8.2), 146 (35), 142 (8.2), 141 (28), 130 (9.5), 97 (50), 90 (9.0), 89 (13), 83 (13), 77 (42), 75 (12), 74 (8.1), 73 (100), 65 (7.9), 59 (18), 50 (9.4), 45 (8.8), 43 (10). <sup>1</sup> H NMR: −0.05 [9H, s,  $-Si(CH_3)$ ; 0.77 [3H, d,  $>CHCH_3$ ,  $J=7.8$ ]; 3.15 [1H, q, >C*H*CH3, *J*=7.8]; 3.35 [3H, s, –OC*H*3]; 5.12 [1H, d,  $H_{3a}$ ,  $J=1.5$ ]; 5.73 [1H, d,  $H_4$ ,  $J=1.5$ ]; 7.12–7.42 [8H, m, *H*<sub>7</sub>, *H*<sub>8</sub>, *H*<sub>9</sub> and Ph]; 7.65 [1H, broad d, *H*<sub>6</sub>, *J*=7.7]. <sup>13</sup>C NMR: −0.72 [3C, –Si(*C*H3)3]; 12.68 [>CH*C*H3]; 48.03 [>*C*HCH3]; 49.69 [*C*4]; 52.80 [–O*C*H3]; 82.56 [*C*9b]; 85.67 [*C*3a]; 93.61 and 99.03 [2C, –*CC*–TMS]; 121.63 [2C, *C ortho* of Ph]; 125.58, 125.90, 126.10, 127.00 and 128.65 [5C,  $C_6$ ,  $C_7$ ,  $C_8$ ,  $C_9$ ,  $C$  para of Ph]; 129.44 [2C,  $C$ *meta* of Ph]; 127.62 and 136.60 [2C,  $C_{5a}$  and  $C_{9a}$ ]; 150.88 [*C ipso* of Ph]; 152.04 [*C*O]; 175.86 [*C*O of lactone].

#### **4.21. 8,9d by deprotection of 8,9c, 8,9e and 8,9j**

**From 8,9c and 8,9e**: protective group has been removed in both cases on **8c**,**e** and **9c**,**e**. A solution of substrate (100 µmol) in acetonitrile (1 ml) was cooled to  $-20^{\circ}$ C and treated with  $\approx$  11% HF in acetonitrile. After 1.5 h a stoichiometric amount of solid  $NAHCO<sub>3</sub>$  with respect to HF was added and the mixture was diluted with water and extracted with  $Et<sub>2</sub>O$ . Chromatography with  $PE/Et_2O$  (1:1) gave the corresponding alcohol **8d** or **9d** (yields, in the range 73–74% for both stereoisomers and whatever the starting silyl ether, have been improved up to 91%, performing the reaction on a 10 mmol scale of **8**,**9c** mixture).

From  $8.9$ j: A solution of  $8.9$ j (65 mg, 100  $\mu$ mol) in dry methanol (2 ml) was cooled to 0°C and treated with  $p$ TSA (19 mg, 100  $\mu$ mol), then, after 5 min, the solution was allowed to stir at rt for 1.5 h. A stoichiometric amount of solid NaHCO<sub>3</sub> with respect to  $p$ TSA was added and the mixture was carefully concentrated under reduced pressure at  $\approx 15^{\circ}$ C. Then it was diluted with water and extracted with  $Et<sub>2</sub>O$ . Chromatography with  $PE/Et_2O$  (1:1) gave the corresponding alcohols **8d** or **9d** with an overall 99% yield.

## **4.22. 8f by protection of 8d**

A solution of  $8d$  (15 mg, 37  $\mu$ mol) in chloroform (2 ml) was treated with 10 mg of powdered 4 Å molecular sieves, 2,6-di-*t*butyl-4-methylpyridine (61 mg, 296  $\mu$ mol) and methyl triflate (17  $\mu$ l, 148  $\mu$ mol) and then heated at 60°C for 48 h (a second addition of an equivalent amount of both reagents was performed after 24 h). After dilution with water, the mixture was extracted with  $Et<sub>2</sub>O$  and crude product was chromatographed with  $PE/Et_2O$  (6:4) to give **8f** (11 mg, 73%) yield).

#### **4.23. 8g by protection of 8d**

The same procedure reported for the preparation of **5e** was followed starting from pure **8d** and the corresponding **8g** was obtained in 87% yield.

## **4.24. 8,9h by protection of 8,9d**

The same procedure reported for the preparation of **5f** was followed starting from pure **8d** or **9d** and the corresponding **8h** and **9h** were obtained in 67–68% yield.

# **4.25. 2-[(***R***\*)-Ethynyl]-4-{(***S***\*)-[1-methyl-2-trityloxy] ethyl}-2***H***-quinoline-1-carboxylic acid phenyl ester, 20**

A solution of **8**,**9j** (with a 82:18 d.r.) (247 mg, 381  $\mu$ mol) in dry MeOH (4 ml) was treated with NaHCO<sub>3</sub>  $(128 \text{ mg}, 1.52 \text{ mmol})$  and heated at 40 $^{\circ}$ C for 2.5 h. After addition of water, the mixture was extracted with Et<sub>2</sub>O. Chromatography with PE/Et<sub>2</sub>O  $9:1 \rightarrow 7:3$  gave desired product (199 mg, 91%) as a white foam. Crystallization from  $Et<sub>2</sub>O/PE$  gave diastereomerically pure **20** as a white solid. Mp 160.7–161.6°C (Et<sub>2</sub>O/PE).  $R_f$ 0.34 (PE/Et<sub>2</sub>O 8:2, A, B). IR:  $v_{\text{max}}$  3303, 2962, 2037, 1712, 1447, 1377, 1302, 1067. GC–MS: unsuitable for this analysis. <sup>1</sup>H NMR: 1.34 [3H, d, >CHC $H_3$ ,  $J=6.5$ ]; 2.19 [1H, d,  $-C\equiv CH$ ,  $J=1.8$ ]; 2.97-3.06 [1H, m, >CHCH<sub>3</sub>]; 3.13–3.29 [2H, m, >CHCH<sub>2</sub>O–]; 5.93–6.01 [2H, m,  $H_2$  and  $H_3$ ]; 6.92 [1H, broad s,  $H_6$ ]; 7.09–7.38 [22H, m,  $H_5$ ,  $H_7$ , Ph, Tr]; 7.74 [1H, broad d,  $H_8$ ,  $J=8.4$ ]. <sup>13</sup>C NMR 16.28 [>CHCH<sub>3</sub>]; 34.15 [>*C*HCH<sub>3</sub>]; 44.09 [*C*2]; 67.14 [–*C*H2O–]; 71.55 [–C*C*H]; 80.20 [–*C*CH]; 86.19 [–*C*Ph3]; 120.82 [*C*8]; 123.44, 125.08 [most likely 2C] and 127.60 [4C,  $C_3$ ,  $C_5$ ,  $C_6$ ,  $C_7$ ]; 121.60 [2C, *C ortho* of Ph]; 125.63 [*C para* of Ph]; 126.73 [3C, *C para* of Tr]; 127.70 and 128.58 [12C, *C ortho* and *meta* of Tr]; 128.83 [*C*4a]; 129.24 [2C, *C meta* of Ph]; 134.11 [*C*4]; 138.28 [*C*8a]; 144.10 [3C, *C ipso* of Tr]; 150.83 [*C ipso* of Ph], 151.83 [*C*O].

**4.26. 2-{(***R***\*)-[3-(Benzyloxy)prop-1-ynyl]}-4-{(***S***\*)-[(2 hydroxy-1-methyl)ethyl]}-2***H***-quinoline-1-carboxylic acid phenyl ester 22 and its epimer 23 and 4-** $\{(S^*)$ **-** $[(2$ **hydroxy-1-methyl)ethyl]}-2-{(***R***\*)-[3-(***p***-methoxybenzyloxy)prop-1-ynyl]}-2***H***-quinoline-1-carboxylic acid phenyl ester, 24 and its epimer, 25**

**By reaction with** *p***-TSA**: the same procedure used for the transformation of **8**,**9j** into **8**,**9d**, but working at rt, was employed for the preparation of **22**,**23** and **24**,**25**, starting from **8**,**9k** and **8**,**9l**, respectively. Chromatography with the appropriate eluent based on  $PE/Et_2O$  gave the corresponding products as pale yellow oils and as inseparable diastereoisomeric mixtures (**22**,**23** in 63% yield and **24**,**25** in 78% yield). **By reaction with EtSH and boron trifluoride**: a solution of **8**,**9k** or **8**,**9l** (500  $\mu$ mol) in dry CH<sub>2</sub>Cl<sub>2</sub> (10 ml) was cooled to 0°C and treated with EtSH (185  $\mu$ l, 2.5 mmol) and BF<sub>3</sub>·Et<sub>2</sub>O  $(127 \mu l, 1.0 \text{ mmol})$ . After 1 h the reaction is usually complete. Quenching with  $NH<sub>4</sub>Cl$  sat. sol. and extraction with  $Et<sub>2</sub>O$  was followed by chromatography with  $PE/Et<sub>2</sub>O$  to give the inseparable diastereoisomeric mixtures **22**,**23** in 89% yield from **8**,**9k**. When the reaction was performed on **8**,**9l** the 4-methoxybenzyl ether was also cleaved and **26**,**27** were isolated in 66% overall yield. **Characterization of 22,23**:  $R_f$  0.31 (22,23) (PE/ Et<sub>2</sub>O 4:6, **A**, **B**). IR (22,23):  $v_{\text{max}}$  3472, 2400, 1712, 1380, 1327, 1304, 1068. GC–MS: unsuitable for this analysis. <sup>1</sup> H NMR: 1.19 [3H, d, >CHC*H*3, *J*=7.0 (**23**)]; 1.34 [3H, d,  $>CHCH_3$ ,  $J=6.8$  (22)]; 3.15 [1H, sextuplet,  $>CHCH_3O_6$ ,  $J=6.8$  (23)]; 3.16 [1H, sextuplet,  $>CHCH<sub>2</sub>O<sub>-</sub>, J=6.8 (23)$ ];  $>CHCH<sub>2</sub>O-, J=6.6 (22)$ ]; 3.50–3.94 [2H, m, –CHC*H*<sub>2</sub>O–  $(22,23)$ ]; 4.03 [2H, s,  $-OCH_2C=C$ <sup> $-$ </sup> (23)]; 4.04 [2H, s, –OC*H*2CC– (**22**)]; 4.27 [2H, s, –OC*H*2Ph (**22**,**23**)]; 6.02 [2H, apparent s,  $H_2$  and  $H_3$  (22,23)]; 7.14–7.49 [13H, m, *H*<sub>5</sub>, *H*<sub>6</sub>, *H*<sub>7</sub>, -OPh, Ph (22,23)]; 7.79 [1H, broad d, *H*<sub>8</sub>, *J*=7.8 (**22**,**23**)]. 13C NMR: 15.91 [>CH*C*H3, (**22**)]; 17.25 [>CH*C*H3, (**23**)]; 36.08 [>*C*HCH3, (**22**)]; 36.22 [>*C*HCH<sub>3</sub>, (23)]; 44.25 [*C*<sub>2</sub> (23)]; 44.56 [*C*<sub>2</sub> (22)]; 57.13 [–*C*H2CC– (**22**,**23**)]; 66.51 [–*C*H2OH (**22**,**23**)]; 71.04  $[-OCH_2Ph (22,23)]$ ; 79.62 and 82.90 [2C,  $-C\equiv C$ (**22**,**23**)]; 121.58 [2C, *C ortho* of Ph (carbamate) (**22**,**23**)]; 123.27, 124.96, 125.07, 127.83, 127.92 and 127.98 [5C, aromatics (**22**,**23**)]; 125.78 [*C para* of Ph (carbamate) (**22**,**23**)]; 127.56 [*C*4a (**22**,**23**)]; 128.23 and 128.35 [4C, *C ortho* and *meta* of Ph (Bn) (**22**,**23**)]; 129.40 [2C, *C meta* of Ph (carbamate) (**22**,**23**)]; 134.50 [*C*<sup>4</sup> (**22**,**23**)]; 137.13 and 137.22 [C<sub>8a</sub> and *C ipso* of Ph (Bn) (22,23)]; 150.98 [*C ipso* of Ph (carbamate) (**22**,**23**)]; 151.83 [*C*O (**22**,**23**)]. **Characterization of 24,25**:  $R_f$  0.30 (24,25) (PE/Et<sub>2</sub>O 2:8, **A**, **B**). IR (**24**,**25**): max 3008, 2411, 1714, 1384, 1303, 1234, 1071. GC-MS: unsuitable for this analysis. <sup>1</sup>H NMR: 1.20 [3H, d, >CHCH<sub>3</sub>, *J*=7.0 (25)]; 1.35 [3H, d, >CHC*H*3, *J*=6.7 (**24**)]; 3.07–3.25 [1H, m, >C*H*CH2O– (**24**,**25**)]; 3.52–3.77 [2H, m, –CHC*H*2O– (**24**,**25**)]; 3.79 [3H, s,  $-OCH_3$  (24,25)]; 4.01 [2H, s,  $-OCH_2C=C-$ (**24**,**25**)]; 4.31 [2H, s, –OC*H*2Ar (**24**,**25**)]; 6.02 [2H, apparent s, *H*<sup>2</sup> and *H*<sup>3</sup> (**24**,**25**)]; 6.82 [2H, dt, *H ortho* to –OMe, *J*=2.4, 8.6 (**24**,**25**)]; 7.09 [2H, dt, *H meta* to –OMe, *J*=2.6, 8.7 (**24**,**25**)]; 7.18–7.48 [8H, m, *H*5, *H*6, *H*<sub>7</sub>,  $-\text{OPh}$  (24,25)]; 7.80 [1H, broad d, *H*<sub>8</sub>, *J*=6.6 (**24**,**25**)]. 13C NMR: 15.89 [>CH*C*H3, (**24**)]; 17.26 [>CHCH<sub>3</sub>, (25)]; 35.99 [>CHCH<sub>3</sub>, (24)]; [>*C*HCH3, (**25**)]; 44.16 [*C*<sup>2</sup> (**25**)]; 44.48 [*C*<sup>2</sup> (**24**)]; 55.24  $[-OCH_3 \ (24,25)]; \ 56.65 \ [-CH_2C \equiv C - \ (24,25)]; \ 66.37$ [–*C*H2OH (**24**,**25**)]; 70.55 [–O*C*H2Ar (**24**,**25**)]; 79.58 (**25**), 79.65 (**24**), 82.72 (**24**) and 82.96 (**25**) [2C, –*CC*–]; 113.68 [2C, *C ortho* to –OMe (**24**,**25**)]; 121.53 [2C, *C ortho* of Ph (**24**,**25**)]; 123.21, 124.86, 125.02, 127.82 (**24**) and 127.89 (**25**) [4C, aromatics (**24**,**25**)]; 125.74 [*C para* of Ph (**24**,**25**)]; 127.45 [*C*CH2CC– (**24**,**25**)]; 129.03 [*C*4a (**25**)]; 129.07 [*C*4a (**24**)]; 129.34 [2C, *C meta* of Ph (**24**,**25**)]; 129.85 [2C, *C meta* to –OMe (**24**,**25**)]; 134.33  $[C_4 (24,25)]$ ; 136.93  $[C_{8a} (24)]$ ; 137.73  $[C_{8a} (25)]$ ; 150.78 [*C ipso* of Ph (**24**,**25**)]; 151.76 [*C*O (**24**,**25**)]; 159.25  $[\geq C$ -OMe (24,25)].

# **4.27. 4-{(***S***\*)-[(2-Hydroxy-1-methyl)ethyl]}-2-[(***R***\*)-3 hydroxyprop-1-ynyl]-2***H***-quinoline-1-carboxylic acid phenyl ester, 26 and its epimer, 27**

**From 20**: (a) **Transformation of 20 into 21**. A solution of  $20$  (124 mg, 215  $\mu$ mol) in dry THF (5 ml) was cooled to  $0^{\circ}$ C and treated with *t*BuMgCl (2 M sol in Et<sub>2</sub>O, 107 -l). After 45 min the solution was stirred for additional 15 min at rt. Then paraformaldehyde (39 mg, 1.29 mg) was added and the slurry was stirred at rt for 17 h; after the addition of an equivalent amount of *t*BuMgCl and

paraformaldehyde the reaction was refluxed for 22 h. Quenching with  $NH<sub>4</sub>Cl$  sat. sol. was followed by extraction with Et<sub>2</sub>O. (b) **Transformation of 21 into 26**. Crude mixture was treated with *p*-TSA in methanol as described for the transformation of **8**,**9j** into **8**,**9d**. Chromatography with  $PE/Et_2O$  (1:9) furnished **26** (11) mg) in 14% overall yield from **20**.

**From 22,23 or 24,25**: the same procedure described in the previous paragraph, using EtSH and  $BF_3$ ·Et<sub>2</sub>O was followed, performing the reaction at reflux (**22**,**23**) or at 0°C (**24**,**25**). The diols **26**,**27**, that have been separated, have been obtained in an overall yield of 50 and 99%, respectively, as pale yellow oils. **Characterization of 26**:  $R_f$  0.47 (Et<sub>2</sub>O 4:6, **A**, **B**). IR:  $v_{\text{max}}$  3461, 2960, 2391, 1713, 1328, 1303. GC–MS (analysis performed on the diastereomeric mixture **26**,**27**): (usual conditions, but with fin. temp. 290°C). *R*<sub>t</sub> 12.05; *m*/*z* 363 (M<sup>+</sup>, 9.8), 345 (15), 305 (11), 304 (52), 288 (12), 286 (14), 242 (11), 222 (25), 212 (12), 210 (14), 196 (11), 195 (10), 194 (46), 193 (12), 192 (13), 184 (20), 183 (8.8), 182 (17), 181 (9.9), 180 (25), 179 (11), 178 (15), 168 (12), 167 (31), 166 (40), 165 (17), 156 (18), 155 (7.0), 154 (22), 153 (9.1), 152 (18), 144 (10), 132 (14), 130 (11), 129 (9.3), 128 (22), 127 (14), 115 (19), 94 (20), 91 (9.5), 78 (10), 77 (100), 65 (25), 55 (11), 51 (24), 44 (16), 43 (10), 41 (10), 39 (25). <sup>1</sup>H NMR: 1.33 [3H, d, >CHC*H*<sub>3</sub>, *J*=6.8] 3.11 [1H, sextuplet,  $>CHCH_2O-, J=6.2$ ; 3.48 and 3.69 [2H, AB part of ABX system,  $-CHCH<sub>2</sub>OH$ ,  $J<sub>AB</sub>=9.8$ ,  $J<sub>AX</sub>$  and  $J_{\text{BX}}=6.1, 6.9$ ; 4.09 [2H, d,  $-OCH_2C=C$ ,  $J=1.6$ ]; 5.93 [1H, d,  $H_3$ ,  $J=6.4$ ]; 5.99 [1H, dt,  $H_2$ ,  $J=1.4$ , 6.6]; 7.15–7.44 [8H, m, H<sub>5</sub>, H<sub>6</sub>, H<sub>7</sub>, Ph]; 7.75 [1H, broad d,  $H_8$ ,  $J=7.6$ ]. <sup>13</sup>C NMR: 15.71 [>CHCH<sub>3</sub>]; 35.92  $[\triangleright \text{CHCH}_3]$ ; 44.42  $[C_2]$ ; 50.97  $[\neg \text{CH}_2C \equiv C \neg]$ ; 66.35 [–*C*H2OH]; 81.56 and 81.89 [2C, –*CC*–]; 121.40 [*C*8]; 123.26, 124.87, 125.06 and 127.93 [4C,  $C_3$ ,  $C_5$ ,  $C_6$ ,  $C_7$ ]; 121.55 [2C, *C ortho* of Ph]; 125.82 [*C para* of Ph]; 127.32 [*C*4a]; 129.39 [2C, *C meta* of Ph]; 134.18 [*C*4]; 136.80 [*C*8a]; 150.81 [*C ipso* of Ph]; 151.96 [*C*O]. **Characterization of 27:**  $R_f$  0.36 (Et<sub>2</sub>O 4:6, **A, B**). IR:  $v_{\text{max}}$ 3426, 2398, 1713, 1380, 1303, 1188, 1135. GC–MS: see compound **26**. <sup>1</sup> H NMR: 1.17 [3H, d, >CHC*H*3, *J*=6.8] 3.11 [1H, sextuplet, >C*H*CH2O–, *J*=6.6]; 3.77 and 3.88 [2H, AB part of ABX system,  $-CHCH<sub>2</sub>OH$ ,  $J<sub>AB</sub>=10.8$ ,  $J_{AX}$  and  $J_{BX} = 5.8$ , 6.2]; 4.12 [2H, d,  $-OCH_2C=C_7$ , *J*=1.8]; 5.95 [1H, d, *H*3, *J*=6.6]; 6.02 [1H, dt, *H*2, *J*=1.7, 6.6]; 7.18–7.55 [8H, m, *H*5, *H*6, *H*7, Ph]; 7.76 [1H, broad d,  $H_8$ ,  $J=6.1$ ]. <sup>13</sup>C NMR: 17.25 [>CHCH<sub>3</sub>]; 36.03 [>*C*HCH3]; 44.14 [*C*2]; 51.07 [–*C*H2CC–]; 66.62  $[-CH_2OH]$ ; 81.86 and 81.97 [2C,  $-C \equiv C-\hat{]}$ ; 120.34 [C<sub>8</sub>]; 123.19, 124.93, 125.11 and 127.92 [4C,  $C_3$ ,  $C_5$ ,  $C_6$ ,  $C_7$ ]; 121.60 [2C, *C ortho* of Ph]; 125.84 [*C para* of Ph]; 127.68 [*C*4a]; 129.42 [2C, *C meta* of Ph]; 134.22 [*C*4]; 137.66 [*C*8a]; 150.90 [*C ipso* of Ph]; 152.00 [*C*O].

# **4.28. 4-{(***R***)-[1-Hydroxymethyl-2-trityloxy]ethyl}-2- {(***R***)-[(trimethylsilyl)ethynyl]}-2***H***-quinoline-1-carboxylic acid phenyl ester, 28 and its epimer at C-2, 29**

Triethylsilyl ether was removed under the same conditions used for the transformation of **8c** and **8e** into **8d**, starting from the diastereomeric mixture **8**,**9o**. After chromatography with  $PE/Et_2O$  7:3 $\rightarrow$ 4:6 the two

epimeric alcohols **28** and **29** were separated and obtained in 97% overall yield as white foams. **Characterization of 28**:  $R_f$  0.43 (PE/Et<sub>2</sub>O 1:1, **A**, **B**).  $[\alpha]_D =$ +255.0 (*c* 1.75, CHCl<sub>3</sub>). IR:  $v_{\text{max}}$  3023, 2995, 2954, 2170, 1708, 1447, 1376, 1190, 1008. GC–MS: unsuitable for this analysis. <sup>1</sup> H NMR: 0.02 [9H, s, –Si(C*H*3)3]; 2.45 [1H, broad s,  $-CH_2OH$ ]; 3.29–3.43 [3H, m,  $>CHCH<sub>2</sub>OTr$ ]; 4.00 [2H, apparent s; became a broad d,  $J=4.0$ , after exchange with D<sub>2</sub>O,  $-CH_2OH$ ; 5.88 and 5.88 [2H, AB system,  $H_2$  and  $H_3$ ,  $J_{AB} = 6.9$ ]; 7.02 [1H, broad d,  $H_6$ ,  $J=7.6$ ]; 7.13–7.41 [22H, m,  $H_5$ ,  $H_7$ , Ph, Tr]; 7.74 [1H, broad d, H<sub>8</sub>, J=7.8]. <sup>13</sup>C NMR: −0.27  $[3C, -Si(CH_3)_3]$ ; 41.55  $[-CH(CH_2O-)_2]$ ; 44.72  $[C_2]$ ; 64.78 and 65.17 [2C, -CH( $CH_2O$ -)<sub>2</sub>]; 87.16 [-CPh<sub>3</sub>]; 88.90 and 101.20 [2C, –*CC*–TMS]; 121.62 [2C, *C ortho* of Ph]; 122.35 [C<sub>8</sub>]; 123.06, 125.04 [most likely 2C]; and 127.83 [4C, *C*3, *C*5, *C*6, *C*7]; 125.68 [*C para* of Ph]; 127.20 [3C, *C para* of Tr]; 127.48 [*C*4a]; 127.96 and 128.50 [12C, *C ortho* and *meta* of Tr]; 129.28 [2C, *C meta* of Ph]; 133.69 [*C*4]; 134.34 [*C*8a]; 144.61 [3C, *C ipso* of Tr]; 150.85 [*C ipso* of Ph], 151.80 [*C*O]. **Characterization of 29**:  $R_f$  0.29 (PE/Et<sub>2</sub>O 2:8, A, B).  $[\alpha]_D =$ −227.8 (*c* 1.16, CHCl<sub>3</sub>). IR: v<sub>max</sub> 3044, 2981, 2305, 1714, 1379, 1327, 1255. GC–MS: unsuitable for this analysis. <sup>1</sup> <sup>1</sup>H NMR: 0.08 [9H, s,  $-Si(CH_3)_3$ ]; 2.22 [1H, broad s, –CH2O*H*]; 3.27 [1H, quintuplet, >C*H*CH2OTr, *J*=6.0]; 3.51 and 3.57 [2H, AB part of ABX system, –CHCH<sub>2</sub>OTr,  $J_{AB} = 9.2$ ,  $J_{AX}$  and  $J_{BX} = 5.0$ , 8.1]; 3.77– 3.84 [2H, m, -CH<sub>2</sub>OH]; 5.82 and 5.88 [2H, AB system, *H*<sub>2</sub> and *H*<sub>3</sub>,  $J_{AB}$ =6.6]; 7.08–7.51 [23H, m, *H*<sub>5</sub>, *H*<sub>6</sub>, *H*<sub>7</sub>, Ph, Tr]; 7.73 [1H, broad d,  $H_8$ ,  $J=7.7$ ]. <sup>13</sup>C NMR: −0.37 [3C, –Si(*C*H3)3]; 41.92 [–*C*H(CH2O–)2]; 44.85  $[C_2]$ ; 64.46 and 64.90 [2C, –CH( $CH_2O$ –)<sub>2</sub>]; 87.29  $\left[-\overline{C}Ph_3\right]$ ; 87.29 and 101.90  $\left[2C, -C\right] = C$ –TMS]; 121.58 [2C, *C ortho* of Ph]; 122.46 [C<sub>8</sub>]; 123.12, 124.94 [most likely 2C]; and 127.78 [4C, *C*3, *C*5, *C*6, *C*7]; 125.71 [*C para* of Ph]; 127.11 [3C, *C para* of Tr]; 127.54 [*C*4a]; 127.94 and 128.68 [12C, *C ortho* and *meta* of Tr]; 129.33 [2C, *C meta* of Ph]; 133.49 [*C*4]; 134.20 [*C*8a]; 144.68 [3C, *C ipso* of Tr]; 150.87 [*C ipso* of Ph], 151.83 [*C*O].

#### **4.29. 8m by acetylation of 29**

The same procedure reported for the preparation of **5f** was followed starting from **29** and the corresponding **8m** was obtained in 82% yield.

# **4.30. 4-{(***S***)-[1-(Toluene-4-sulfonyloxymethyl)-2-trityloxy]ethyl}-2-{(***S***)-[(trimethylsilyl)ethynyl]}-2***H***-quinoline-1-carboxylic acid phenyl ester, 30**

A solution of  $29$  (202 mg, 304  $\mu$ mol) in dry CH<sub>2</sub>Cl<sub>2</sub> (3 ml) was cooled a t 0°C and treated with 4-dimethylaminopyridine (4 mg, 33 µmol), triethylamine (148 µl, 106 mmol) and freshly distilled *p*-TsCl (87 mg, 456 µmol). After 5 min the reaction was stirred at rt for 7 h. After addition of 5% aq  $NH<sub>4</sub>H<sub>2</sub>PO<sub>4</sub>$ , the reaction was extracted as usual with  $Et<sub>2</sub>O$ . Chromatography with  $PE/Et_2O \ 8:2 \rightarrow 7:3$  gave pure **30** (176 mg, 71% yield) as a white foam.  $R_f$  0.44 (PE/Et<sub>2</sub>O 6:4, **A**, **B**).  $[\alpha]_D = -60.5$ (*c* 1.32, CHCl3). IR: max 2954, 2170, 1720, 1377, 1326, 1291, 1174. GC-MS: unsuitable for this analysis. <sup>1</sup>H

NMR: 0.13 [9H, s, –Si(CH<sub>3</sub>)<sub>3</sub>]; 2.36 [3H, s, –CH<sub>3</sub>]; 3.27 [1H, quintuplet,  $-CHCH_2OTr$ ,  $J=5.7$ ]; 3.27 and 3.43 [2H, AB part of ABX system, >CHC $H_2$ OTr,  $J_{AB}$ =9.0,  $J_{AX}$  and  $J_{BX} = 5.0, 6.8$ ]; 4.18 and 4.24 [2H, AB part of ABX system, >CHC*H*<sub>2</sub>OTs,  $J_{AB}$ =9.6,  $J_{AX}$  and  $J_{BX}$ = 5.0, 5.9]; 5.77 and 5.84 [2H, AB system,  $H_2$  and  $H_3$ , *J*AB=6.8]; 7.06–7.42 [25H, m, *H*5, *H*6, *H*7, Ph, Tr, *H ortho* to –SO<sub>2</sub>–]; 7.64 [2H, apparent d, *H ortho* to  $-SO<sub>2</sub>$ ,  $J=8.4$ ]; 7.72 [1H, broad d,  $H<sub>8</sub>$ ,  $J=8.2$ ]. <sup>13</sup>C NMR: −0.41 [3C, −Si(*CH*<sub>3</sub>)<sub>3</sub>]; 39.37 [−*CH*(CH<sub>2</sub>O−)<sub>2</sub>]; 44.76 [*C*2]; 62.11 and 69.18 [2C, –CH(*C*H2O–)2]; 86.96  $[-CPh_3]$ ; 88.94 and 100.48  $[2C, -C \equiv C - TMS]$ ; 121.61 [2C, *C ortho* of Ph]; 122.69, 123.30, and 124.87 [most likely 2C] [4C, 4C (between  $C_3$ ,  $C_5$ ,  $C_6$ ,  $C_7$ ,  $C_8$ , while one is overlapped with another signal)]; 125.77 [*C para* of Ph]; 127.05 [3C, *C para* of Tr]; 127.79 [2C, *C ortho* to –SO2–]; 127.85 and 128.69 [12C, *C ortho* and *meta* of Tr]; 128.47 [*C*4a]; 129.38 [2C, *C meta* of Ph]; 129.80 [2C, *C* meta to  $-SO_2$ -]; 131.87 [*C*<sub>4</sub>]; 132.50 [-*C*SO<sub>2</sub>-]; 134.16 [*C*8a]; 143.58 [3C, *C ipso* of Tr]; 144.73 [–*C*Me of Ts]; 150.88 [*C ipso* of Ph], 151.78 [*C*O].

#### **4.31. 9j by reduction of 30**

A solution of  $30$  (48 mg,  $59$   $\mu$ mol) in dry THF (3 ml) was treated with NaI (198 mg, 132 µmol) and AIBN  $[2,2'-azobis(2-methylpropionitrile)]$  (2.5 mg, 15  $\mu$ mol). Then  $nBu_3SnH$  (24  $\mu$ l, 89  $\mu$ mol) was added via syringe and the resulting mixture was heated at 85°C in a sealed tube for 19 h. After diluting with water, an extraction was performed with  $Et<sub>2</sub>O$ . Chromatography with  $PE/Et_2O \t 9:1 \rightarrow 6:4$  gave **9**j as a pale yellow oil [11] mg, 29% yield (39% on unrecovered starting material)].

#### **4.32. NOE difference experiments on 16, 18, 19**

The samples were prepared  $(20 \text{ mg/ml})$  using CDCl<sub>3</sub> previously dried on 3 A molecular sieves, and freshly passed through alumina. The tubes were placed under nitrogen and kept for 30 min in an ultrasound bath, in order to remove oxygen. NOEDIF spectra were acquired at 200 MHz, using a delay of 3.9 s and 256 transients. A decoupling power so that  $\gamma H_2 = 3-4$  Hz was used. The NOEs were calculated on the basis of the integrals for the 'normal' spectrum and the decoupled one. **Compound 16**: irradiation of OMe gave, at 49% saturation, NOEs of 5.5, 2.5 and 1.0 for  $H_{6a}$ ,  $\equiv$ C-*H* and  $H<sub>1</sub>$ , respectively, corresponding to effective NOEs of 11.2, 5.2 and 2.1. **Compound 18**: (a) irradiation of OMe gave, at 44% saturation, NOEs of 8.0, 3.4 and 0.8 for  $H_{3a}$ ,  $H_9$ , and CH<sub>3</sub>CHCO, respectively, corresponding to effective NOEs of 18.3, 7.8 and 1.8; (b) Irradiation of C*H*3CHCO gave, at 31 saturation, NOEs of 1.3, 0.7 and 0.6 for  $H_9$ ,  $H_{3a}$  and OC $H_3$ , respectively, corresponding to effective NOEs of 4.2, 2.2 and 1.8; (c) Irradiation of CH<sub>3</sub>CHCO gave, at 43 saturation, NOEs of 0.9, 0.5 and 0.5 for  $H_9$ ,  $H_4$  and  $H_{3a}$ , respectively, corresponding to effective NOEs of 2.0, 1.2 and 1.1. **Compound 19**: (a) irradiation of OMe gave, at 50% saturation, NOEs of 8.6, and 3.8 for  $H_{3a}$  and  $H_9$ , respectively, corresponding to effective NOEs of 17.1 and 7.6; (b) Irradiation of  $CH<sub>3</sub>CHCO$  gave, at 73 saturation, a NOE of 4.5 for  $H<sub>9</sub>$ , corresponding to an

effective NOE of 6.2; (c) Irradiation of CH<sub>3</sub>CHCO gave, at 73 saturation, NOEs of 5.3 and 1.9 for  $H_{3a}$  and *H*<sub>9</sub>, respectively, corresponding to effective NOEs of 7.3 and 2.6.

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